Datasheet: STAR132F

BATCH NUMBER 173082

Description:	GOAT ANTI MOUSE IgG1:FITC
Specificity:	IgG1
Format:	FITC
Product Type:	Polyclonal Antibody
Isotype:	Polyclonal IgG
Quantity:	0.5 mg

Product Details

Applications

This product has been reported to work in the following applications. This information is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for further information. For general protocol recommendations, please visit www.bio-rad-antibodies.com/protocols.

	Yes	No	Not Determined	Suggested Dilution
Flow Cytometry	▪			1/1000
Immunohistology - Frozen	▪			
Immunohistology - Paraffin			▪	
Immunofluorescence	▪			Neat - 1/10
Rare Cell / CTC Enumeration	▪			1/100

Where this antibody has not been tested for use in a particular technique this does not necessarily exclude its use in such procedures. Suggested working dilutions are given as a guide only. It is recommended that the user titrates the antibody for use in their own system using appropriate negative/positive controls.

Target Species

Mouse

Species Cross Reactivity

Does not react with:Human

Product Form

Purified IgG conjugated to Fluorescein Isothiocyanate Isomer 1 (FITC) - liquid

Max Ex/Em

Fluorophore	Excitation Max (nm)	Emission Max (nm)
FITC	490	525

Antiserum Preparation

Antisera to mouse IgG1 were raised by repeated immunisation of goats with purified antigen. Purified IgG was prepared from whole serum by affinity chromatography.

Buffer Solution

Phosphate buffered saline

Preservative Stabilisers	<0.1% Sodium Azide (NaN_3)
Approx. Protein Concentrations	IgG concentration 1.0mg/ml
Immunogen	Mouse IgG1 paraproteins.
External Database Links	**UniProt:** P01869 Related reagents P01868 Related reagents **Entrez Gene:** 16017 Ighg1 Related reagents 16017 Ighg1 Related reagents
Synonyms	Igh-4
RRID	AB_2124271
Specificity	**Goat anti Mouse IgG1 antibody** recognizes Mouse IgG1. This antibody has been cross absorbed against mouse IgM, IgG2a, IgG2b, IgG3 and IgA, pooled human sera and purified human paraproteins. Goat anti Mouse IgG1 antibody shows minimal cross-reactivity with human immunoglobulins. Goat anti Mouse IgG1 antibody has been validated for use on the Genesis Cell Isolation System with the CelSelect Slide™ technology.
Flow Cytometry	Use 10ul of the suggested working dilution to label 10^6 cells in 100ul.
References	1. Croft, N.P. *et al.* (2009) Stage-specific inhibition of MHC class I presentation by the Epstein-Barr virus BNLF2a protein during virus lytic cycle. PLoS Pathog. 5(6): e1000490. 2. Zuo, J. *et al.* (2011) The Epstein-Barr virus-encoded BILF1 protein modulates immune recognition of endogenously processed antigen by targeting MHC class I molecules trafficking on both the exocytic and endocytic pathways. J Virol. 85: 1604-14. 3. Knipping, K. *et al.* (2011) A gastrointestinal rotavirus infection mouse model for immune modulation studies. Virol J. 8: 109. 4. Young, D. *et al.* (2012) Soy-derived di- and tripeptides alleviate colon and ileum inflammation in pigs with dextran sodium sulfate-induced colitis. J Nutr. 142 (2): 363-8. 5. Bagai, U. and Pawar, A. (2013) A blood stage fraction of *Plasmodium berghei* induces protective and long lasting immune response in BALB/c mice. Parasitol Int. 62: 329-36. 6. Anda, S. *et al.* (2014) Cell-cycle analyses using thymidine analogues in fission yeast. PLoS One. 9 (2): e88629. 7. Kamat, M.M. *et al.* (2016) Changes in Myeloid Lineage Cells in the Uterus and Peripheral Blood of Dairy Heifers During Early Pregnancy. Biol Reprod. 95 (3): 68. 8. Ramanathan, R. *et al.* (2015) Transplantation of human stem cell-derived hepatocytes in an animal model of acute liver failure. Surgery. 158 (2): 349-59.

9. Hwang, S.R. *et al.* (2015) Altered expression levels of neurodevelopmental proteins in fetal brains of BTBR T+tf/J mice with autism-like behavioral characteristics. J Toxicol Environ Health A. 78 (8): 516-23.
10. Zhao, Z. *et al.* (2015) Multiple B-cell epitope vaccine induces a Staphylococcus enterotoxin B-specific IgG1 protective response against MRSA infection. Sci Rep. 5: 12371.
11. Gutiérrez-Miranda, B. *et al.* (2020) Oleacein Attenuates the Pathogenesis of Experimental Autoimmune Encephalomyelitis through Both Antioxidant and Anti-Inflammatory Effects. Antioxidants (Basel). 9 (11): 1161.
12. Gutierrez, B. *et al.* (2020) Oleanolic acid ameliorates intestinal alterations associated with EAE. J Neuroinflammation. 17 (1): 363.
13. Apóstolo, N. *et al.* (2020) Synapse type-specific proteomic dissection identifies IgSF8 as a hippocampal CA3 microcircuit organizer. Nat Commun. 11 (1): 5171.
14. Zhuang, X. *et al.* (2020) CAR T cells targeting tumor endothelial marker CLEC14A inhibit tumor growth. JCI Insight. 5 (19): e138808.
15. Sparks, A.M. *et al.* (2018) Natural Selection on Antihelminth Antibodies in a Wild Mammal Population. Am Nat. 192 (6): 745-760.
16. Kushwaha, V. *et al.* (2019) Troponin 1 of human filarial parasite *Brugia malayi.*: cDNA cloning, expression, purification, and its immunoprophylactic potential. Parasitol Res. 118 (6): 1849-63.
17. Kim, Y.H. *et al.* (2019) Effects of ruminal pH on gene expression in the rumen epithelium, peripheral blood mononuclear cell subpopulations, and blood metabolites from Holstein calves during weaning transition. J Vet Med Sci. 81 (6): 808-816.
18. Jimbo, S. *et al.* (2019) Natural and inducible regulatory B cells are widely distributed in ovine lymphoid tissues. Vet Immunol Immunopathol. 211: 44-48.
19. Sparks, A.M. *et al.* (2019) The genetic architecture of helminth-specific immune responses in a wild population of Soay sheep (*Ovis aries.*). PLoS Genet. 15 (11): e1008461.
20. Nedumpun, T. *et al.* (2019) Negative Immunomodulatory Effects of Type 2 Porcine Reproductive and Respiratory Syndrome Virus-Induced Interleukin-1 Receptor Antagonist on Porcine Innate and Adaptive Immune Functions. Front Immunol. 10: 579.
21. Verma, R. *et al.* (2018) *Leishmania donovani.* molecules recognized by sera of filaria infected host facilitate filarial infection. Parasitol Res. 117 (9): 2901-12.
22. Gatkowska, J. *et al.* (2019) The Impact of the Antigenic Composition of Chimeric Proteins on Their Immunoprotective Activity against Chronic Toxoplasmosis in Mice. Vaccines (Basel). 7(4):154.
23. Han, H.J. *et al.* (2022) Metal arsenic mediated enhancement of type-2 immunity in brains with altered locomotive activities in mice with autism-like behavioral characteristics. Toxicol Res. 38 (1): 27-33.
24. Anda, S. *et al.* (2021) Cosegregation of asymmetric features during cell division. Open Biol. 11 (8): 210116.
25. Minaei, S. *et al.* (2018) Propranolol efficacy as a novel adjuvant for immunization against *Toxoplasma gondii.* tachyzoites. Exp Parasitol. 194: 60-66.
26. Ngoepe, T.A. *et al.* (2023) Th1 and Th2 epitopes of Cowdria polymorphic gene 1 of *Ehrlichia ruminantium*. Onderstepoort J Vet Res. 90 (1): e1-e15.
27. Jo, J. *et al.* (2023) Immunodysregulatory potentials of polyethylene or polytetrafluorethylene microplastics to mice subacutely exposed via intragastric intubation.

Toxicol Res. 39 (3): 419-27.
28. Sessevmez, M. *et al.* (2023) Induction of humoral and cell-mediated immunity in mice by chitosan-curdlan composite nanoparticles administered intranasally and subcutaneously J Drug Deliv Sci Technol. 86: 104704.
29. Veisi, R. *et al.* (2024) Expression and purification of TolC as a recombinant protein vaccine against *Shigella flexneri* and evaluation of immunogenic response in mice. Microb Pathog. : 106539 [Epub ahead of print]
30. Faber, E. *et al.* (2024) Identification of T cell and linear B cell epitopes on African horse sickness virus serotype 4 proteins VP1-1, VP2, VP4, VP7 and NS3. Vaccine. 42 (2): 136-145.
31. Pretorius, A. *et al.* (2024) Screening for immune biomarkers associated with infection or protection against *Ehrlichia ruminantium* by RNA-sequencing analysis. Microb Pathog. : 106588.
32. Kusma, S. *et al.* (2024) Oral subacute polypropylene microplastics administration effect on potential immunotoxicity in ICR mice. J Toxicol Environ Health A. : 1-10 [Epub ahead of print].
33. Wu, Y. *et al.* (2020) Sterilizing immunity against SARS-CoV-2 in hamsters conferred by a novel recombinant subunit vaccine bioRxiv 20 Dec [Epub ahead of print].
34. Karabowicz, J. *et al.* (2024) Analysis of the role of *Dirofilaria repens* macrophage migration inhibitory factors in host-parasite interactions. J Vet Res. 68 (3): 381-8.
35. Amaral, R. *et al.* (2025) Chitosan Nanoparticles for Enhanced Immune Response and Delivery of Multi-Epitope *Helicobacter pylori* Vaccines in a BALB/c Mouse Model. Pharmaceutics. 17 (1): 132.
36. Corripio-Miyar, Y. *et al.* (2025) T-helper cell phenotypes are repeatable, positively correlated, and associated with helminth infection in wild Soay sheep. Discov Immunol. 4 (1): kyae017.

Storage	This product is shipped at ambient temperature. It is recommended to aliquot and store at -20°C on receipt. When thawed, aliquot the sample as needed. Keep aliquots at 2-8°C for short term use (up to 4 weeks) and store the remaining aliquots at -20°C. Avoid repeated freezing and thawing as this may denature the antibody. Storage in frost-free freezers is not recommended. This product is photosensitive and should be protected from light.
Guarantee	12 months from date of despatch
Acknowledgements	CelSelect Slides is a trademark of Bio-Rad Laboratories, Inc. in certain jurisdictions.
Health And Safety Information	Material Safety Datasheet documentation #10040 available at: https://www.bio-rad-antibodies.com/SDS/STAR132F
Regulatory	For research purposes only

Product inquiries: www.bio-rad-antibodies.com/technical-support

To find a batch/lot specific datasheet for this product, please use our online search tool at: bio-rad-antibodies.com/datasheets

'M428947:240301'