

Datasheet: RMT1 BATCH NUMBER 159031

Description:	RAT MONOCLONAL ANTIBODY ISOTYPING TEST KIT
Name:	RAT ISOTYPING KIT
Format:	Kit
Product Type:	Kits
Quantity:	10 TESTS

Product Details

Applications	This product has been reported to work in the following applications. This information is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for further information. For general protocol recommendations, please visit <u>www.bio-rad-antibodies.com/protocols</u> .					
		Yes	Νο	Not Determined	Suggested Dilution	
	Isotyping Assay	•				
	Where this product has n necessarily exclude its us a guide only. It is recomm system using appropriate	not been te se in such nended tha e negative/j	sted for us procedure at the user positive co	se in a particular techn s. Suggested working titrates the product fo ontrols.	ique this does not dilutions are given as r use in their own	
Target Species	Rat					
Product Information	The rat monoclonal antib (<0.1 %).	ody isotypi	ing test kit	shows no cross-react	ivity with bovine IgG	
Test Principle	The assay principle is bac coloured micro particles a its isotype. The isotyping antibodies corresponding IgG2b, IgG2c and IgM) a positive flow control band particles have migrated the monoclonal antibody can pipetting the diluted samp antibody coated micro par strip until it is bound by the monoclonal's isotype and complex will aggregate a monoclonal antibody's iso	sed on mo and equally strip has in to each o nd to kapp d, which ind hrough the be screen ple into the articles, and he immobil d its light ch s blue ban otype and i	use polycl y reactive mmobilize f the comm a light cha dicates tha strip. By u ed for isot dinserting ized mous nain. In ap ds in the t its light cha	onal anti-rat antibodie to any rat monoclonal d bands of monoclonal non rat antibody isotyp ains only. One side of the at the antibody-coated using these two compo- type by simply diluting nent tube where it form the strip. This comple- te anti-rat antibody spe- proximately 5-10 minu- wo sections correspor- ain. Development of the	s coupled onto antibody regardless of al mouse anti-rat bes (IgG1, IgG2a, the strip bears a coloured micro onents, a rat the antibody sample, ns a complex with the ex flows through the ecific for the rat ates, the micro particle nding to the me strip is complete	

Reagents In The Kit	1 Desiccant vial containing rat isotyping test strips, 10 tests.
	10 Capped ready-to-use lyophilized microparticle development tubes.

Instructions For Use Note: All reagents should be brought to room temperature before use.

Sample Preparation:

Dilute all monoclonal antibody samples to a concentration of 1.0 ug/ml in PBS containing 1% w/v bovine serum albumin (BSA). If the concentration of the sample is entirely unknown, make dilutions based on the following estimates:

Typically, serum contains between 10-15 mg/ml IgG and ascites can be as high as 10 mg/ml . Hollow fibre bioreactor culture supernatants contain approximately 0.5-1.0 mg/ml, whereas static flask tissue culture supernatants usually contain 10-50 ug/ml. Using these estimates, the appropriate dilutions can be made.

150ul of the diluted sample will be added to the development tubes.

Assay Protocol:

1. Remove the required number of isotyping strips from the desiccant vial and replace thecap. Remove the caps from an equal number of development tubes. **Note:** the tubes may be labeled with a marker for identification.

2. Pipette 150ul of the freshly diluted sample into each development tube and incubate at room temperature for 30 seconds. Vortex the tube briefly to ensure that the coloured micro particle solution is completely re-suspended.

3. Place one isotyping strip, with the solid red end at the bottom, into each development tube.

Interpretation of Results:

Interpret the results at 5-10 minutes once the positive flow control bands have appeared. Within 5-10 minutes, a blue band will appear above the letters in one of the class or subclass windows as well as in the kappa window of the strip, indicating the heavy and light-chain composition of the monoclonal antibody (lambda light chains are very rare in rat monoclonal antibodies, but if present no positive band will be seen in the kappa window). The intensity of the blue bands will increase as the sample continues to flow up the strip. The positive flow control bands on one side of the isotyping test strip should also appear, indicating that the antibody-coated micro particles are functional and have flowed up the strip. In cases where the sample is very dilute, the development time may take up to 10 minutes.

Note: For a permanent experimental record or for an easier interpretation of results when testing multiple samples, the solid red area may be cut off the bottom of the strip to

	prevent further band development once the positive flow control bands have appeared. A gentle stream of air can be applied to the membrane portion of the strip to assist in drying the membrane and preventing any further development. Do not wash the strip to stop the reaction.
References	 Esashi, E. <i>et al</i> (2004) Development of CD4+ macrophages from intrathymic T cell progenitors is induced by thymic epithelial cells. J Immunol. 173: 4360-67 Taylor, P.R. <i>et al</i>. (2005) Dectin-2 is predominantly myeloid restricted and exhibits unique activation-dependent expression on maturing inflammatory monocytes elicited <i>in vivo</i>. Eur J Immunol.35: 2163-74. Ohashi, S. <i>et al</i>. (2010) Preparation of Anti–fragrant Monoclonal Antibodies by the Rat Lymph Node Method and Their Characterization Using Enzyme–linked Immunosorbent Assay J. Fac. Agr., Kyushu Univ., 55 (1), 91–96 Balyasnikova, I.V. <i>et al</i>. (2005) Monoclonal antibodies to native mouse angiotensinconverting enzyme (CD143): ACE expression quantification, lung endothelial cell targeting and gene delivery. Tissue Antigens. 67: 10-29. Kato, M. <i>et al</i>. (2009) Production of monoclonal antibody specific for bottlenose dolphin neutrophils and its application to cell separation. Dev Comp Immunol. 33 (1): 14-7. Sawano, S. <i>et al</i>. (2016) A One-Step Immunostaining Method to Visualize Rodent Muscle Fiber Type within a Single Specimen. PLoS One. 11 (11): e0166080. Fichou, N. <i>et al</i>. (2009) Splicing variations in the ligand-binding domain of ApoER2 results in functional differences in the binding properties to Reelin. Neurosci Res. 63 (4): 251-8. Hibi, T. <i>et al</i>. (2006) Functional analysis of nuclear pore complex protein Nup62/p62 using monoclonal antibodies. Hybridoma (Larchmt). 25 (2): 51-9. Koide, A. & Koide, S. (2007) Monobodies: antibody mimics based on the scaffold of the fibronectin type III domain. Methods Mol Biol. 352: 95-109. Tanaka, M. <i>et al</i>. (2016) Identification of low-abundance proteins in serum via the isolation of HSP72 complexes. J Proteomics. 136: 214-21.
Storage	Store at +4°C. DO NOT FREEZE. Do not use components from different lots.
Guarantee	Guaranteed until date of expiry. Please see product label.
Health And Safety Information	Material Safety Datasheet documentation #qtec44-2 available at: https://www.bio-rad-antibodies.com/SDS/RMT1 qtec44-2 Not required
Regulatory	For research purposes only
Technical Advice	PROBLEM

No heavy and or light-chain band appeared on the strip, but the positive flow control bands appeared.

Possible causes:

1. The antibody concentration was too low - prepare a less dilute sample and re-test.

2. No antibody was in the sample - the hybridoma is either not secreting or is not a rat monoclonal. If possible sub-clone the hybridoma and re-test.

3. Freshly diluted samples were not used - prepare fresh dilutions and re-test.

PROBLEM

Multiple heavy and light-chain bands appear on the strip.

Possible causes:

1. Antibody concentration was too high - dilute sample further and re-test.

2. For ascites, there may be small amounts of contaminating antibodies produced - dilute sample further and re-test.

3. For tissue culture supernatant, a mixed culture may be present - re-clone the hybridoma and re-test.

PROBLEM

No positive flow control bands appear.

Possible causes:

1. Sample volume was too low (<150ul) - carefully dilute a fresh sample and pipette 150ul into a new development tube and re-test.

2. Strip removed from development tube too early - re-test and allow strip to react for at least 10 minutes.

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To find a batch/lot specific datasheet for this product, please use our online search tool at: bio-rad-antibodies.com/datasheets 'M371200:200603'

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