Datasheet: MMT2

Description:	MOUSE MONOCLONAL ANTIBODY ISOTYPING TEST KIT
Name:	MOUSE ISOTYPING KIT
Format:	Kit
Product Type:	Kits
Quantity:	10 TESTS

Product Details

Applications

This product has been reported to work in the following applications. This information is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for further information. For general protocol recommendations, please visit www.bio-rad-antibodies.com/protocols.

	Yes	No	Not Determined	Suggested Dilution
Isotyping Assay	▪			

Where this product has not been tested for use in a particular technique this does not necessarily exclude its use in such procedures. Suggested working dilutions are given as a guide only. It is recommended that the user titrates the product for use in their own system using appropriate negative/positive controls.

Target Species

Mouse

Preservative Stabilisers

Reagent B contains 0.05% Sodium Azide (NaN_3)

Specificity

The mouse monoclonal antibody isotyping test kit shows no cross-reactivity with bovine IgG (<0.1 %).

Product Information

The Mouse Monoclonal Antibody Isotyping Test Kit provides rapid and reliable identification of mouse monoclonal antibody isotypes in 5–10 minutes using a simple dip-stick format. It is well suited for routine screening of hybridoma supernatants or purified antibodies without the need for specialized instrumentation.

The assay detects all major mouse immunoglobulin classes and subclasses (IgG1, IgG2a, IgG2b, IgG3, IgM, IgA) as well as kappa and lambda light chains, with built-in flow control bands to confirm correct assay performance.

The mouse monoclonal antibody isotyping test kit shows no cross-reactivity with bovine IgG (<0.1 %).

Test Principle

The assay principle is based on anti-mouse kappa and anti-mouse lambda antibodies

coupled onto coloured micro particles and equally reactive to any mouse monoclonal antibody regardless of its isotype. The isotyping strip has immobilized bands of goat anti-mouse antibodies corresponding to each of the common mouse antibody isotypes (IgG1, IgG2a, IgG2b, IgG3, IgM, and IgA) and to the kappa and lambda light chains. Both sides of the strip bear a positive flow control band, which indicates that the antibody-coated coloured micro particles have migrated through the strip. By using these two components, a mouse monoclonal antibody can be screened for isotype by simply diluting the antibody sample, pipetting the diluted sample into the development tube where it forms a complex with the antibody coated micro particles, and inserting the strip. This complex flows through the strip until it is bound by the immobilized goat anti-mouse antibody specific for the monoclonal's isotype and its light chain. In approximately 5-10 minutes, the micro particle complex will aggregate as blue bands in the two sections corresponding to the monoclonal antibody's isotype and its light chain. Development of the strip is complete when the positive flow control band on each side of the test strip turns blue. Please note, this kit is not suitable for use with antibodies of the isotype IgG2c; some cross-reactivity may be observed with isotype IgG2a.

Reagents In The Kit	1 Desiccant vial containing mouse isotyping test strips, 10 tests. Reagent B dropper bottle containing anti-mouse IgG coated blue microparticles, 1.0 mL. Ready-to-use solution containing 0.05% sodium azide.
Additional Reagents Required	1% BSA in Phosphate-buffered saline (PBS) pH 7.2-7.6 Microtiter plate or small glass test tube

Instructions For Use

Note: All reagents should be brought to room temperature before use.

Sample Preparation:

Dilute all monoclonal antibody samples to a concentration of 1.0 ug/ml in PBS containing 1% w/v bovine serum albumin (BSA). If the concentration of the sample is entirely unknown, make dilutions based on the following estimates:

Typically, serum contains between 10-15 mg/ml IgG and ascites can be as high as 10 mg/ml. Hollow fibre bioreactor culture supernatants contain approximately 0.5-1.0 mg/ml, whereas static flask tissue culture supernatants usually contain 10-50 ug/ml. Using these estimates, the appropriate dilutions can be made.

50µl of the diluted sample will be added to the microtiter plate well or test tube.

Assay Protocol:

1. Remove the required number of isotyping strips from the desiccant vial and replace the cap.

2. Gently mix **Reagent B** (dropper bottle) by inverting and rotating the bottle to avoid the formation of bubbles or foam. Carefully dispense three (3) drops of Reagent B by completely inverting the dropper bottle over each of the microtiter wells or test tubes. 3. Pipette 50µl of the freshly diluted sample into each well or tube and incubate at room

temperature for at least 30 seconds.

4. Place one isotyping strip, with the solid red end at the bottom, into each development tube.

Interpretation of Results:

Interpret the results at 5-10 minutes once the positive flow control bands have appeared.

Within 5-10 minutes, a blue band will appear above the letters in one of the class or subclass windows as well as in the kappa window of the strip, indicating the heavy and light-chain composition of the monoclonal antibody (lambda light chains are very rare in rat monoclonal antibodies, but if present no positive band will be seen in the kappa window). The intensity of the blue bands will increase as the sample continues to flow up the strip. The positive flow control bands on one side of the isotyping test strip should also appear, indicating that the antibody-coated micro particles are functional and have flowed up the strip. In cases where the sample is very dilute, the development time may take up to 10 minutes.

Note: For a permanent experimental record or for an easier interpretation of results when testing multiple samples, the solid red area may be cut off the bottom of the strip to prevent further band development once the positive flow control bands have appeared. A gentle stream of air can be applied to the membrane portion of the strip to assist in drying the membrane and preventing any further development.

Do not wash the strip to stop the reaction.

References

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Storage	Please refer to batch-specific Certificate of Analysis
Guarantee	Guaranteed until date of expiry. Please see product label.
Health And Safety Information	Material Safety Datasheet documentation #20521 #20511 available at: https://www.bio-rad-antibodies.com/SDS/MMT2
Regulatory	For research purposes only
Technical Advice	PROBLEM **No heavy and or light-chain band appeared on the strip, but the positive flow control bands appeared.** *Possible causes:* 1. The antibody concentration was too low - prepare a less dilute sample and re-test. 2. No antibody was in the sample - the hybridoma is either not secreting or is not a mouse monoclonal. If possible sub-clone the hybridoma and re-test.

3. Freshly diluted samples were not used - prepare fresh dilutions and re-test.

PROBLEM

Multiple heavy and light-chain bands appear on the strip.

Possible causes:

1. Antibody concentration was too high - dilute sample further and re-test.
2. For ascites, there may be small amounts of contaminating antibodies produced - dilute sample further and re-test.
3. For tissue culture supernatant, a mixed culture may be present - re-clone the hybridoma and re-test.

PROBLEM

No positive flow control bands appear.

Possible causes:

1. Sample volume was too low (<150ul) - carefully dilute a fresh sample and pipette 150ul into a new development tube and re-test.
2. Strip removed from development tube too early - re-test and allow strip to react for at least 10 minutes.

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