

## Datasheet: MCA928PE

**BATCH NUMBER 172391**

<b>Description:</b>	MOUSE IgG1 NEGATIVE CONTROL:RPE
<b>Specificity:</b>	MOUSE IgG1 NEGATIVE CONTROL
<b>Format:</b>	RPE
<b>Product Type:</b>	Negative/Isotype Control
<b>Isotype:</b>	IgG1
<b>Quantity:</b>	100 TESTS

### Product Details

#### Applications

This product has been reported to work in the following applications. This information is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for further information. For general protocol recommendations, please visit [www.bio-rad-antibodies.com/protocols](http://www.bio-rad-antibodies.com/protocols).

	Yes	No	Not Determined	Suggested Dilution
Flow Cytometry	▪			*

Where this antibody has not been tested for use in a particular technique this does not necessarily exclude its use in such procedures. Suggested working dilutions are given as a guide only. \*It is recommended that the user titrates the antibody for use in their own system using appropriate negative/positive controls.

<b>Target Species</b>	Negative Control
<b>Product Form</b>	Purified IgG conjugated to R. Phycoerythrin (RPE) - lyophilized
<b>Reconstitution</b>	Reconstitute with 1 ml distilled water

Max Ex/Em	Fluorophore	Excitation Max (nm)	Emission Max (nm)
	RPE 488nm laser	496	578

<b>Preparation</b>	Purified IgG prepared by affinity chromatography on Protein A from tissue culture supernatant
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<b>Buffer Solution</b>	Phosphate buffered saline
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<b>Preservative</b>	0.09% Sodium Azide
<b>Stabilisers</b>	1% Bovine Serum Albumin
	5% Sucrose

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<b>Specificity</b>	<p><b>Mouse IgG1 negative control</b> is negative by flow cytometry on all human cells and cell lines tested. Further tests have also shown that this reagent is also suitable for use as a negative control with bovine (Maslanka <i>et al</i>, 2012), ovine, porcine (<a href="#">Kapetanovic <i>et al</i>, 2012</a>), equine (<a href="#">Jacks <i>et al</i>, 2007</a>), canine (<a href="#">Maiolini <i>et al</i>, 2012</a>), lapine (<a href="#">Pakandl <i>et al</i>, 2008</a>) and guinea-pig tissues.</p> <p><b><i>This reagent recognizes a rat cell surface marker, and therefore cannot be used as a negative control in this species.</i></b></p>
<b>Flow Cytometry</b>	Use 10ul of the suggested working dilution to label 10 <sup>6</sup> separated cells or 100ul lysed whole peripheral blood.
<b>References</b>	<ol style="list-style-type: none"><li>1. Kupatt, C. <i>et al</i>. (2000) c7E3Fab reduces postischemic leukocyte-thrombocyte interaction mediated by fibrinogen. Implications for myocardial reperfusion injury. <a href="#">Arterioscler Thromb Vasc Biol. 20 (10): 2226-32.</a></li><li>2. Jacks, S. <i>et al</i>. (2007) Experimental infection of neonatal foals with <i>Rhodococcus equi</i> triggers adult-like gamma interferon induction. <a href="#">Clin Vaccine Immunol. 14:669-77</a></li><li>3. Pakandl, M. <i>et al</i>. (2008) Immune response to rabbit coccidiosis: a comparison between infections with <i>Eimeria flavescens</i> and <i>E. intestinalis</i>. <a href="#">Folia Parasitol (Praha). 55:1-6.</a></li><li>4. Dalli, J. <i>et al</i>. (2008) Annexin 1 mediates the rapid anti-inflammatory effects of neutrophil-derived microparticles. <a href="#">Blood. 112 (6): 2512-9.</a></li><li>5. Barratt-Due, A. <i>et al</i>. (2011) <i>Ornithodoros moubata</i> Complement Inhibitor Is an Equally Effective C5 Inhibitor in Pigs and Humans. <a href="#">J Immunol. 187: 4913-9.</a></li><li>6. Maślanka, T. <i>et al</i>. (2012) The presence of CD25 on bovine WC1+ gamma delta T cells is positively correlated with their production of IL-10 and TGF-beta, but not IFN-gamma. <a href="#">Pol J Vet Sci. 15 (1): 11-20.</a></li><li>7. Maiolini, A. <i>et al</i>. (2012) Toll-like receptors 4 and 9 are responsible for the maintenance of the inflammatory reaction in canine steroid-responsive meningitis-arteritis, a large animal model for neutrophilic meningitis. <a href="#">J Neuroinflammation. 9: 226.</a></li><li>8. Kapetanovic, R. <i>et al</i>. (2012) Pig bone marrow-derived macrophages resemble human macrophages in their response to bacterial lipopolysaccharide. <a href="#">J Immunol. 188: 3382-94.</a></li><li>9. Kamble, N.M. <i>et al</i>. (2016) Interaction of a live attenuated <i>Salmonella Gallinarum</i> vaccine candidate with chicken bone marrow-derived dendritic cells. <a href="#">Avian Pathol. 45 (2): 235-43.</a></li><li>10. Iwaszko-Simonik, A. <i>et al</i>. (2015) Expression of surface platelet receptors (CD62P and CD41/61) in horses with recurrent airway obstruction (RAO). <a href="#">Vet Immunol Immunopathol. 164 (1-2): 87-92.</a></li><li>11. Brace, P.T. <i>et al</i>. (2017) <i>Mycobacterium tuberculosis</i> subverts negative regulatory pathways in human macrophages to drive immunopathology. <a href="#">PLoS Pathog. 13 (6): e1006367.</a></li><li>12. Topoluk, N. <i>et al</i>. (2017) Amniotic Mesenchymal Stromal Cells Exhibit Preferential Osteogenic and Chondrogenic Differentiation and Enhanced Matrix Production Compared With Adipose Mesenchymal Stromal Cells. <a href="#">Am J Sports Med. 45 (11): 2637-46.</a></li><li>13. Arzi, B. <i>et al</i>. (2017) Therapeutic Efficacy of Fresh, Allogeneic Mesenchymal Stem Cells for Severe Refractory Feline Chronic Gingivostomatitis. <a href="#">Stem Cells Transl Med. 6 (8): 1710-22.</a></li></ol>

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14. Taechangam, N. *et al.* (2021) Feline adipose-derived mesenchymal stem cells induce effector phenotype and enhance cytolytic function of CD8+ T cells. [Stem Cell Res Ther. 12 \(1\): 495.](#)
15. do Prado Duzanski, A.*et al.* (2022) Cell-mediated immunity and expression of MHC class I and class II molecules in dogs naturally infected by canine transmissible venereal tumor: Is there complete spontaneous regression outside the experimental CTVT? [Research in Veterinary Science. 145: 193-204.](#)
16. Tolstova, T. *et al.* (2023) The effect of TLR3 priming conditions on MSC immunosuppressive properties. [Stem Cell Res Ther. 14 \(1\): 344.](#)
17. Geng, Y. *et al.* (2018) Dietary vitamin D(3) supplementation protects laying hens against lipopolysaccharide-induced immunological stress. [Nutr Metab \(Lond\). 15: 58.](#)
18. Dan-Jumbo, S.O. *et al.* (2024) Derivation and long-term maintenance of porcine skeletal muscle progenitor cells. [Sci Rep. 14 \(1\): 9370.](#)
19. Maciag, S. *et al.* (2022) Effects of freezing storage on the stability of maternal cellular and humoral immune components in porcine colostrum. [Vet Immunol Immunopathol. 254: 110520.](#)
20. Forner, R. *et al.* (2021) Distribution difference of colostrum-derived B and T cells subsets in gilts and sows. [PLoS One. 16 \(5\): e0249366.](#)
21. Rogato, F. *et al.* (2024) Leukemia cutis as a prominent clinical sign in a dog with acute myeloid leukemia. [Vet Clin Pathol. 53 \(4\): 448-57.](#)

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**Storage**

This product is shipped at ambient temperature.  
Prior to reconstitution store at +4°C. Following reconstitution store at +4°C.

DO NOT FREEZE.

This product should be stored undiluted. This product is photosensitive and should be protected from light. Should this product contain a precipitate we recommend microcentrifugation before use.

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**Guarantee**

12 months from date of despatch

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**Health And Safety Information**

Material Safety Datasheet documentation #20487 available at: <https://www.bio-rad-antibodies.com/SDS/MCA928PE>

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**Regulatory**

For research purposes only

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## Related Products

### Recommended Negative Controls

[MOUSE IgG1 NEGATIVE CONTROL:RPE \(MCA1209PE\)](#)

**Product inquiries:** [www.bio-rad-antibodies.com/technical-support](http://www.bio-rad-antibodies.com/technical-support)

To find a batch/lot specific datasheet for this product, please use our online search tool at: [bio-rad-antibodies.com/datasheets](http://bio-rad-antibodies.com/datasheets)

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