

Datasheet: MCA928F

Description:	MOUSE IgG1 NEGATIVE CONTROL:FITC
Specificity:	MOUSE IgG1 NEGATIVE CONTROL
Format:	FITC
Product Type:	Negative/Isotype Control
Isotype:	IgG1
Quantity:	100 TESTS

Product Details

Applications

This product has been reported to work in the following applications. This information is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for further information. For general protocol recommendations, please visit www.bio-rad-antibodies.com/protocols.

	Yes	No	Not Determined	Suggested Dilution
Flow Cytometry	▪			*

Where this antibody has not been tested for use in a particular technique this does not necessarily exclude its use in such procedures. *It is recommended that the user dilutes the antibody for use in their own system to a concentration equivalent to their test reagents.

Target Species	Negative Control		
Product Form	Purified IgG conjugated to Fluorescein Isothiocyanate Isomer 1 (FITC) - liquid.		
Max Ex/Em	Fluorophore	Excitation Max (nm)	Emission Max (nm)
	FITC	490	525
Preparation	Purified IgG prepared by affinity chromatography on Protein A from tissue culture supernatant		
Buffer Solution	Phosphate buffered saline		
Preservative	0.09% Sodium Azide		
Stabilisers	1% Bovine Serum Albumin		
Approx. Protein Concentrations	IgG concentration 0.1 mg/ml		
RRID	AB_322262		

Specificity

Mouse IgG1 negative control is negative by flow cytometry on all human cells and cell lines tested. Further tests have also shown that this reagent is also suitable for use as a negative control with bovine (Maslanka *et al*, 2012), ovine, porcine ([Kapetanovic *et al*, 2012](#)), equine ([Jacks *et al*, 2007](#)), canine ([Maiolini *et al*, 2012](#)), lapine ([Pakandl *et al*, 2008](#)) and guinea-pig tissues.

This reagent recognizes a rat cell surface marker, and therefore cannot be used as a negative control in this species.

Flow Cytometry

Use 10ul of the suggested working dilution to label 10⁶ cells or 100ul whole blood.

References

1. Kupatt, C. *et al*. (2000) c7E3Fab reduces postischemic leukocyte-thrombocyte interaction mediated by fibrinogen. Implications for myocardial reperfusion injury. [Arterioscler Thromb Vasc Biol. 20 \(10\): 2226-32.](#)
2. Dalli, J. *et al*. (2008) Annexin 1 mediates the rapid anti-inflammatory effects of neutrophil-derived microparticles. [Blood. 112 \(6\): 2512-9.](#)
3. Barratt-Due, A. *et al*. (2011) Ornithodoros moubata Complement Inhibitor Is an Equally Effective C5 Inhibitor in Pigs and Humans. [J Immunol. 187: 4913-9.](#)
4. Kapetanovic, R. *et al*. (2012) Pig bone marrow-derived macrophages resemble human macrophages in their response to bacterial lipopolysaccharide. [J Immunol. 188: 3382-94.](#)
5. Maiolini, A. *et al*. (2012) Toll-like receptors 4 and 9 are responsible for the maintenance of the inflammatory reaction in canine steroid-responsive meningitis-arteritis, a large animal model for neutrophilic meningitis. [J Neuroinflammation. 9: 226.](#)
6. Maślanka, T. *et al*. (2012) The presence of CD25 on bovine WC1+ gammadelta T cells is positively correlated with their production of IL-10 and TGF-beta, but not IFN-gamma. [Pol J Vet Sci. 15 \(1\): 11-20.](#)
7. Pakandl, M. *et al*. (2008) Immune response to rabbit coccidiosis: a comparison between infections with Eimeria flavescens and E. intestinalis. [Folia Parasitol \(Praha\). 55:1-6.](#)
8. Jacks, S. *et al*. (2007) Experimental infection of neonatal foals with Rhodococcus equi triggers adult-like gamma interferon induction. [Clin Vaccine Immunol. 14:669-77](#)
9. Kamble, N.M. *et al*. (2016) Interaction of a live attenuated *Salmonella gallinarum* vaccine candidate with chicken bone marrow-derived dendritic cells. [Avian Pathol. Jan 26:1-24. \[Epub ahead of print\]](#)
10. Brace, P.T. *et al*. (2017) *Mycobacterium tuberculosis* subverts negative regulatory pathways in human macrophages to drive immunopathology. [PLoS Pathog. 13 \(6\): e1006367.](#)
11. Topoluk, N. *et al*. (2017) Amniotic Mesenchymal Stromal Cells Exhibit Preferential Osteogenic and Chondrogenic Differentiation and Enhanced Matrix Production Compared With Adipose Mesenchymal Stromal Cells. [Am J Sports Med. 363546517706138.](#)
12. Iwaszko-Simonik, A. *et al*. (2015) Expression of surface platelet receptors (CD62P and CD41/61) in horses with recurrent airway obstruction (RAO). [Vet Immunol Immunopathol. 164 \(1-2\): 87-92.](#)
13. Arzi, B. *et al*. (2017) Therapeutic Efficacy of Fresh, Allogeneic Mesenchymal Stem Cells for Severe Refractory Feline Chronic Gingivostomatitis. [Stem Cells Transl Med. 6 \(8\): 1710-22.](#)
14. Taechangam, N. *et al*. (2021) Feline adipose-derived mesenchymal stem cells induce effector phenotype and enhance cytolytic function of CD8+ T cells. [Stem Cell Res Ther.](#)

Storage This product is shipped at ambient temperature. It is recommended to aliquot and store at -20°C on receipt. When thawed, aliquot the sample as needed. Keep aliquots at 2-8°C for short term use (up to 4 weeks) and store the remaining aliquots at -20°C.

Avoid repeated freezing and thawing as this may denature the antibody. Storage in frost-free freezers is not recommended. This product is photosensitive and should be protected from light.

Guarantee 12 months from date of despatch

Health And Safety Information Material Safety Datasheet documentation #10041 available at: 10041: <https://www.bio-rad-antibodies.com/uploads/MSDS/10041.pdf>

Regulatory For research purposes only

Related Products

Recommended Negative Controls

[MOUSE IgG1 NEGATIVE CONTROL:FITC \(MCA1209F\)](#)

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To find a batch/lot specific datasheet for this product, please use our online search tool at: [bio-rad-antibodies.com/datasheets](https://www.bio-rad-antibodies.com/datasheets)

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