

## Datasheet: MCA928

**BATCH NUMBER 172199**

|                      |                             |
|----------------------|-----------------------------|
| <b>Description:</b>  | MOUSE IgG1 NEGATIVE CONTROL |
| <b>Specificity:</b>  | MOUSE IgG1 NEGATIVE CONTROL |
| <b>Format:</b>       | Purified                    |
| <b>Product Type:</b> | Negative/Isotype Control    |
| <b>Isotype:</b>      | IgG1                        |
| <b>Quantity:</b>     | 100 TESTS                   |

### Product Details

#### Applications

This product has been reported to work in the following applications. This information is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for further information. For general protocol recommendations, please visit [www.bio-rad-antibodies.com/protocols](http://www.bio-rad-antibodies.com/protocols).

|                | Yes | No | Not Determined | Suggested Dilution |
|----------------|-----|----|----------------|--------------------|
| Flow Cytometry | ■   |    |                | *                  |

Where this antibody has not been tested for use in a particular technique this does not necessarily exclude its use in such procedures. Suggested working dilutions are given as a guide only. \*It is recommended that the user titrates the antibody for use in their own system to a concentration equivalent to that of their test reagents.

|                                       |   |
|---------------------------------------|---|
| <b>Target Species</b>                 | Negative Control  |
| <b>Product Form</b>                   | Purified IgG - liquid   |
| <b>Preparation</b>                    | Purified IgG prepared by affinity chromatography on Protein A from tissue culture supernatant |
| <b>Buffer Solution</b>                | Phosphate buffered saline   |
| <b>Preservative Stabilisers</b>       | 0.09% Sodium Azide<br>1% Bovine Serum Albumin   |
| <b>Approx. Protein Concentrations</b> | IgG concentration 0.1mg/ml  |
| <b>RRID</b>                           | AB_322259   |
| <b>Specificity</b>                    | <b>Mouse IgG1 negative control</b> is negative by flow cytometry on all human cells and cell  |

lines tested. Further tests have also shown that this reagent is also suitable for use as a negative control with bovine (Maslanka *et al*, 2012), ovine, porcine ([Kapetanovic \*et al\*, 2012](#)), equine ([Jacks \*et al\*, 2007](#)), canine ([Maiolini \*et al\*, 2012](#)), lapine ([Pakandl \*et al\*, 2008](#)) and guinea-pig tissues.

***This reagent recognizes a rat cell surface marker, and therefore cannot be used as a negative control in this species.***

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**Flow Cytometry** Use 10ul of the suggested working dilution to label 10<sup>6</sup> cells or 100ul whole blood

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**References**

1. Kupatt, C. *et al*. (2000) c7E3Fab reduces postischemic leukocyte-thrombocyte interaction mediated by fibrinogen. Implications for myocardial reperfusion injury. [Arterioscler Thromb Vasc Biol. 20 \(10\): 2226-32.](#)
2. Jacks, S. *et al*. (2007) Experimental infection of neonatal foals with *Rhodococcus equi* triggers adult-like gamma interferon induction. [Clin Vaccine Immunol. 14:669-77](#)
3. Pakandl, M. *et al*. (2008) Immune response to rabbit coccidiosis: a comparison between infections with *Eimeria flavescens* and *E. intestinalis*. [Folia Parasitol \(Praha\). 55:1-6.](#)
4. Dalli, J. *et al*. (2008) Annexin 1 mediates the rapid anti-inflammatory effects of neutrophil-derived microparticles. [Blood. 112 \(6\): 2512-9.](#)
5. Barratt-Due, A. *et al*. (2011) *Ornithodoros moubata* Complement Inhibitor Is an Equally Effective C5 Inhibitor in Pigs and Humans. [J Immunol. 187: 4913-9.](#)
6. Maślanka, T. *et al*. (2012) The presence of CD25 on bovine WC1+ gammadelta T cells is positively correlated with their production of IL-10 and TGF-beta, but not IFN-gamma. [Pol J Vet Sci. 15 \(1\): 11-20.](#)
7. Maiolini, A. *et al*. (2012) Toll-like receptors 4 and 9 are responsible for the maintenance of the inflammatory reaction in canine steroid-responsive meningitis-arteritis, a large animal model for neutrophilic meningitis. [J Neuroinflammation. 9: 226.](#)
8. Kapetanovic, R. *et al*. (2012) Pig bone marrow-derived macrophages resemble human macrophages in their response to bacterial lipopolysaccharide. [J Immunol. 188: 3382-94.](#)
9. Kamble, N.M. *et al*. (2016) Interaction of a live attenuated *Salmonella Gallinarum* vaccine candidate with chicken bone marrow-derived dendritic cells. [Avian Pathol. 45 \(2\): 235-43.](#)
10. Iwaszko-Simonik, A. *et al*. (2015) Expression of surface platelet receptors (CD62P and CD41/61) in horses with recurrent airway obstruction (RAO). [Vet Immunol Immunopathol. 164 \(1-2\): 87-92.](#)
11. Brace, P.T. *et al*. (2017) *Mycobacterium tuberculosis* subverts negative regulatory pathways in human macrophages to drive immunopathology. [PLoS Pathog. 13 \(6\): e1006367.](#)
12. Topoluk, N. *et al*. (2017) Amniotic Mesenchymal Stromal Cells Exhibit Preferential Osteogenic and Chondrogenic Differentiation and Enhanced Matrix Production Compared With Adipose Mesenchymal Stromal Cells. [Am J Sports Med. 45 \(11\): 2637-46.](#)
13. Arzi, B. *et al*. (2017) Therapeutic Efficacy of Fresh, Allogeneic Mesenchymal Stem Cells for Severe Refractory Feline Chronic Gingivostomatitis. [Stem Cells Transl Med. 6 \(8\): 1710-22.](#)
14. Taechangam, N. *et al*. (2021) Feline adipose-derived mesenchymal stem cells induce effector phenotype and enhance cytolytic function of CD8+ T cells. [Stem Cell Res Ther. 12 \(1\): 495.](#)

15. do Prado Duzanski, A. *et al.* (2022) Cell-mediated immunity and expression of MHC class I and class II molecules in dogs naturally infected by canine transmissible venereal tumor: Is there complete spontaneous regression outside the experimental CTVT? [Research in Veterinary Science. 145: 193-204.](#)
16. Tolstova, T. *et al.* (2023) The effect of TLR3 priming conditions on MSC immunosuppressive properties. [Stem Cell Res Ther. 14 \(1\): 344.](#)
17. Geng, Y. *et al.* (2018) Dietary vitamin D(3) supplementation protects laying hens against lipopolysaccharide-induced immunological stress. [Nutr Metab \(Lond\). 15: 58.](#)
18. Dan-Jumbo, S.O. *et al.* (2024) Derivation and long-term maintenance of porcine skeletal muscle progenitor cells. [Sci Rep. 14 \(1\): 9370.](#)
19. Maciag, S. *et al.* (2022) Effects of freezing storage on the stability of maternal cellular and humoral immune components in porcine colostrum. [Vet Immunol Immunopathol. 254: 110520.](#)
20. Forner, R. *et al.* (2021) Distribution difference of colostrum-derived B and T cells subsets in gilts and sows. [PLoS One. 16 \(5\): e0249366.](#)
21. Rogato, F. *et al.* (2024) Leukemia cutis as a prominent clinical sign in a dog with acute myeloid leukemia. [Vet Clin Pathol. 53 \(4\): 448-57.](#)

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**Storage** This product is shipped at ambient temperature. It is recommended to aliquot and store at -20°C on receipt. When thawed, aliquot the sample as needed. Keep aliquots at 2-8°C for short term use (up to 4 weeks) and store the remaining aliquots at -20°C.

Avoid repeated freezing and thawing as this may denature the antibody. Storage in frost-free freezers is not recommended.

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**Guarantee** 12 months from date of despatch

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**Health And Safety Information** Material Safety Datasheet documentation #10041 available at: <https://www.bio-rad-antibodies.com/SDS/MCA928>

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**Regulatory** For research purposes only

## Related Products

### Recommended Negative Controls

[MOUSE IgG1 NEGATIVE CONTROL \(MCA1209\)](#)

**Product inquiries:** [www.bio-rad-antibodies.com/technical-support](http://www.bio-rad-antibodies.com/technical-support)

To find a batch/lot specific datasheet for this product, please use our online search tool at: [bio-rad-antibodies.com/datasheets](http://bio-rad-antibodies.com/datasheets)

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