

Datasheet: MCA874F BATCH NUMBER 159725

| Description: | MOUSE ANTI HUMAN MACROPHAGES:FITC |
|---------------|------------------------------------|
| Specificity: | MACROPHAGES/MONOCYTES/GRANULOCYTES |
| Format: | FITC |
| Product Type: | Monoclonal Antibody |
| Clone: | MAC387 |
| Isotype: | lgG1 |
| Quantity: | 0.1 mg |
| | |

Product Details

| Applications | This product has been reported to work in the following applications. This information is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for further information. For general protocol recommendations, please visit <u>www.bio-rad-antibodies.com/protocols</u> . | | | | | |
|-----------------------------|--|--|---|-----------------------|--|--|
| | | Yes No | Not Determined | Suggested Dilution | | |
| | Flow Cytometry (1) | - | | Neat | | |
| | Where this antibody has not been tested for use in a particular technique this does not | | | | | |
| | a guide only. It is reco system using appropri (1) Membrane perme a | mmended that th ate negative/posi abilisation is req | e user titrates the antibod tive controls. | n. Bio-Rad recommends | | |
| Target Species | Human | | | | | |
| Species Cross Reactivity | Reacts with: Horse, Pig, Dog, Rabbit, Baboon, Bovine, Guinea Pig, Rat, Cat, Cynomolgus monkey, Rhesus Monkey, Goat, Fallow deer, Pygmy hippopotamus, Mink, Marmoset N.B. Antibody reactivity and working conditions may vary between species. Cross reactivity is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for further information. | | | | | |
| Product Form | Purified IgG conjugate | ed to Fluorescein | Isothiocyanate Isomer 1 (| (FITC) - liquid | | |
| Max Ex/Em | Fluorophore | Excitation Max (| nm) Emission Max (nm) | | | |
| | FITC | 490 | 525 | - | | |
| Preparation | Purified IgG prepared | by affinity chrom | atography on Protein A fro | om tissue culture | | |

| | supernatant |
|-----------------------------------|--|
| Buffer Solution | Phosphate buffered saline |
| Preservative Stabilisers | 0.09% Sodium Azide 1% Bovine Serum Albumin |
| Approx. Protein Concentrations | IgG concentration 0.1 mg/ml |
| Immunogen | Human monocytes. |
| External Database Links | UniProt: P06702 Related reagents Entrez Gene: 6280 \$100A9 Related reagents |
| Synonyms | CAGB, CFAG, MRP14 |
| RRID | AB_323431 |
| Fusion Partners | Spleen cells from immunised BALB/c mice were fused with cells of the mouse NS1 myeloma cell line. |
| Specificity | Mouse anti Human macrophages, clone MAC387 recognizes the L1 or Calprotectin molecule, an intracytoplasmic antigen comprised of a 12 kDa alpha chain and a 14 kDa beta chain. Although originally described as binding to epitopes common to both the alpha and beta chains (<u>Flavell <i>et al.</i> 1987</u>) subsequent studies indicate that the antibody detects an epitope exclusively expressed on the beta chain (<u>Goebeler <i>et al.</i> 1994</u>) demonstrated by immunofluorescent and western blotting on both naturally expressing and transfected targets. In addition Mouse anti Human macrophages, clone MAC387 detects the beta chain in complex with the alpha. |
| | The antigen recognized by Mouse anti Human macrophages, clone MAC387 is expressed by granulocytes, monocytes and by tissue macrophages. Variable results have been reported for staining brain macrophages and microglia. The epitope recognized appears to be well conserved and the antibody is routinely used for the detection of myeloid cells in a wide range of species. |
| Flow Cytometry | Use 10ul of the suggested working dilution to label 1×10^6 cells in 100ul. |
| References | Ueland, T. <i>et al.</i> (2009) Dickkopf-1 enhances inflammatory interaction between platelets and endothelial cells and shows increased expression in atherosclerosis. <u>Arterioscler</u> <u>Thromb Vasc Biol. 29: 1228-34</u> Brandtzaeg, P. <i>et al.</i> (1992) The leucocyte protein L1 (calprotectin): usefulness as an immunohistochemical marker antigen and putative biological function. <u>Histopathology. 21:</u> |

<u>191-196.</u>

3. Gutierrez, M. *et al.* (1999) The detection of CD2+, CD4+, CD8+, and WC1+ T lymphocytes, B cells and macrophages in fixed and paraffin embedded bovine tissue using a range of antigen recovery and signal amplification techniques. <u>Vet Immunol Immunopathol. 71 (3-4): 321-34.</u>

 Ramsay, A.D. *et al.* (1991) Phenotypic analysis of malignant lymphoma in simian immunodeficiency virus infection using anti-human antibodies. <u>J Pathol. 164 (4): 321-8.</u>
 Christgau, M. *et al.* (1998) Characterization of immunocompetent cells in the diseased canine periodontium. <u>J Histochem Cytochem. 46 (12): 1443-54.</u>

Pérez, J. *et al.* (1999) Immunohistochemical study of the inflammatory infiltrate associated with equine squamous cell carcinoma. <u>J Comp Pathol. 121 (4): 385-97.</u>
 Nanney, L.B. *et al.* (2008) Calreticulin enhances porcine wound repair by diverse biological effects. <u>Am J Pathol. 173: 610-30.</u>

 Poncelet, L. *et al.* (2008) Detection of antigenic heterogeneity in feline coronavirus nucleocapsid in feline pyogranulomatous meningoencephalitis. <u>Vet Pathol. 45: 140-53.</u>
 Sethi, R.S. *et al.* (2010) Immunolocalization of pulmonary intravascular macrophages,

TLR4, TLR9 and IL-8 in normal and Pasteurella multocida-infected lungs of water buffalo (Bubalus bubalis). <u>J Comp Pathol. 144: 135-44.</u>

10. Sanchez, J. *et al.* (2011) Microscopical and immunological features of tuberculoid granulomata and cavitary pulmonary tuberculosis in naturally infected goats. <u>J Comp</u> Pathol. 145 (2-3): 107-17.

Isling, L.K. *et al.* (2010) Pyelonephritis in slaughter pigs and sows: morphological characterization and aspects of pathogenesis and aetiology. <u>Acta Vet Scand. 52: 48.</u>
 Vranckx, K. *et al.* (2012) Vaccination reduces macrophage infiltration in bronchus-associated lymphoid tissue in pigs infected with a highly virulent Mycoplasma hyopneumoniae strain. <u>BMC Vet Res. 8: 24.</u>

Campuzano, O. *et al.* (2012) Arrhythmogenic right ventricular cardiomyopathy: severe structural alterations are associated with inflammation. <u>J Clin Pathol. 65 (12): 1077-83.</u>
 García-Jiménez, W.L. (2012) Histological and immunohistochemical characterisation of *Mycobacterium bovis* induced granulomas in naturally infected fallow deer (*Dama dama*). Vet Immunol Immunopathol. 149: 66-75.

15. Santana, C.H. *et al.* (2016) Relationship Between the Inflammatory Infiltrate and the Degree of Differentiation of the Canine Cutaneous Cell Carcinoma. <u>Vet Anim Sci. Oct 10</u> [Epub ahead of print]

16. Masure, D. *et al.* (2013) A Role for Eosinophils in the Intestinal Immunity against Infective *Ascaris suum* Larvae. <u>PLoS Negl Trop Dis. 2013 Mar;7(3): e2138.</u>

17. Tellez, A. *et al.* (2014) Experimental evaluation of efficacy and healing response of everolimus-eluting stents in the familial hypercholesterolemic swine model: a comparative study of bioabsorbable versus durable polymer stent platforms. <u>Coron Artery Dis. 25 (3):</u> 198-207.

18. Collin, N. *et al.* (2009) Sand fly salivary proteins induce strong cellular immunity in a natural reservoir of visceral leishmaniasis with adverse consequences for *Leishmania*. <u>PLoS Pathog. 5(5):e1000441</u>.

19. McCurdy, P. *et al.* (2014) Acute lymphoblastic leukemia in a pygmy hippopotamus (*Hexaprotodon liberiensis*). J Zoo Wildl Med. 45 (4): 906-10.

20. Marcaccini, A. *et al.* (2008) Pseudorabies virus infection in mink: a host-specific pathogenesis. <u>Vet Immunol Immunopathol. 124 (3-4): 264-73.</u>

| | Romero-Palomo, F. <i>et al.</i> (2015) Immunopathologic Changes in the Thymus of Calves Pre-infected with BVDV and Challenged with BHV-1. <u>Transbound Emerg Dis. Aug 25.</u> [Epub ahead of print] Rossi, C.N. <i>et al.</i> (2016) <i>In situ</i> Cutaneous cellular immune response in dogs naturally infected by visceral leishmaniasis. <u>Rev Inst Med Trop Sao Paulo. 58:.</u> Vrolyk V <i>et al.</i> (2016) Lung Inflammation Associated With Clinical Acute Necrotizing Pancreatitis in Dogs. <u>Vet Pathol. May 11. pii: 0300985816646432.</u> [Epub ahead of print] Nelson, M. <i>et al.</i> (2014) Comparative experimental subcutaneous glanders and melioidosis in the common marmoset (<i>Callithrix jacchus</i>). <u>Int J Exp Pathol. 95 (6): 378-91.</u> Amarilla, S.P. <i>et al.</i> (2016) Thymic depletion of lymphocytes is associated with the virulence of PRRSV-1 strains. <u>Vet Microbiol. 188: 47-58.</u> Gacía-Jiménez, W.L. <i>et al.</i> (2013) Immunopathology of granulomas produced by <i>Mycobacterium bovis</i> in naturally infected wild boar. <u>Vet Immunol Immunopathol. 156</u> (1-2): 54-63. Pilling, D. <i>et al.</i> (2015) The long pentraxin PTX3 promotes fibrocyte differentiation. <u>PLoS One. 10 (3): e0119709.</u> Zhao, L. <i>et al.</i> (2017) CCAAT/enhancer-binding protein delta promotes intracellular lipid accumulation in M1 macrophages of vascular lesions. <u>Cardiovasc Res. 113 (11): 1376-88.</u> Wacinski, P. <i>et al.</i> (2021) Anti-Inflammatory Effect of Very High Dose Local Vessel Wall Statin Administration: Poly(L,L-Lactide) Biodegradable Microspheres with Simvastatin for Drug Delivery System (DDS). <u>Int J Mol Sci. 22 (14)Jul 13 [Epub ahead of print].</u> |
|----------------------------------|---|
| Further Reading | Burk, J. <i>et al.</i> (2013) Equine cellular therapyfrom stall to bench to bedside? <u>Cytometry</u> <u>A. 83 (1): 103-13.</u> Piriou-Guzylack, L. (2008) Membrane markers of the immune cells in swine: an update. <u>Vet Res. 39: 54.</u> |
| Storage | This product is shipped at ambient temperature. It is recommended to aliquot and store at -20°C on receipt. When thawed, aliquot the sample as needed. Keep aliquots at 2-8°C for short term use (up to 4 weeks) and store the remaining aliquots at -20°C. Avoid repeated freezing and thawing as this may denature the antibody. Storage in frost-free freezers is not recommended. This product is photosensitive and should be protected from light. |
| Guarantee | 12 months from date of despatch |
| Health And Safety Information | Material Safety Datasheet documentation #10041 available at: https://www.bio-rad-antibodies.com/SDS/MCA874F 10041 |
| Regulatory | For research purposes only |
| | |

Related Products

Recommended Negative Controls

MOUSE IgG1 NEGATIVE CONTROL:FITC (MCA928F)

Recommended Useful Reagents

HUMAN SEROBLOCK (BUF070A) HUMAN SEROBLOCK (BUF070B)

North & South Tel: +1 800 265 7376 Tel: +44 (0)1865 852 700 Worldwide America Fax: +1 919 878 3751 Fax: +44 (0)1865 852 739 Email: antibody_sales_us@bio-rad.com

Europe Email: antibody_sales_uk@bio-rad.com

Tel: +49 (0) 89 8090 95 21 Fax: +49 (0) 89 8090 95 50 Email: antibody_sales_de@bio-rad.com

To find a batch/lot specific datasheet for this product, please use our online search tool at: bio-rad-antibodies.com/datasheets 'M385015:210513'

Printed on 15 Mar 2024

© 2024 Bio-Rad Laboratories Inc | Legal | Imprint