

Datasheet: MCA874APC

## **BATCH NUMBER 161592**

Description:	MOUSE ANTI HUMAN MACROPHAGES:APC
Specificity:	MACROPHAGES/MONOCYTES/GRANULOCYTES
Other names:	CALPROTECTIN
Format:	APC
<b>Product Type:</b>	Monoclonal Antibody
Clone:	MAC387
Isotype:	lgG1
Quantity:	100 TESTS

# **Product Details**

## **Applications**

This product has been reported to work in the following applications. This information is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for further information. For general protocol recommendations, please visit <a href="www.bio-rad-antibodies.com/protocols">www.bio-rad-antibodies.com/protocols</a>.

	Yes	No	Not Determined	Suggested Dilution
Flow Cytometry (1)	-			Neat - 1/5

Where this product has not been tested for use in a particular technique this does not necessarily exclude its use in such procedures. Suggested working dilutions are given as a guide only. It is recommended that the user titrates the product for use in their own system using appropriate negative/positive controls.

(1) Membrane permeabilisation is required for this application. Bio-Rad recommends the use of Leucoperm<sup>™</sup> (Product Code <u>BUF09</u>) for this purpose.

Target Species	Human
Species Cross Reactivity	Reacts with: Horse, Pig, Dog, Rabbit, Baboon, Bovine, Guinea Pig, Rat, Cat, Cynomolgus monkey, Rhesus Monkey, Goat, Fallow deer, Pygmy hippopotamus, Mink, Marmoset <b>N.B.</b> Antibody reactivity and working conditions may vary between species. Cross reactivity is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for further information.
Product Form	Purified IgG conjugated to Allophycocyanin (APC) - lyophilized
Reconstitution	Reconstitute with 1.0 ml distilled water  Care should be taken during reconstitution as the protein may appear as a film at the bottom of the vial. Bio-Rad recommend that the vial is gently mixed after reconstitution.

Max Ex/Em	Fluorophore Excitation Max (nm) Emission Max (nm)  APC 650 661
Preparation	Purified IgG prepared by affinity chromatography on Protein A from tissue culture supernatant
Buffer Solution	Phosphate buffered saline
Preservative Stabilisers	<ul><li>0.09% Sodium Azide</li><li>1% Bovine Serum Albumin</li><li>5% Sucrose</li></ul>
Immunogen	Human monocytes.
External Database Links	UniProt: P06702 Related reagents  Entrez Gene:
	6280 S100A9 Related reagents
Synonyms	CAGB, CFAG, MRP14
Fusion Partners	Spleen cells from immunised BALB/c mice were fused with cells of the mouse NS1 myeloma cell line.
Specificity	Mouse anti Human macrophages, clone MAC387 recognizes the L1 or Calprotectin molecule, an intracytoplasmic antigen comprised of a 12 kDa alpha chain and a 14 kDa beta chain. Although originally described as binding to epitopes common to both the alphand beta chains (Flavell et al. 1987) subsequent evidence indicates that the antibody detects an epitope exclusively expressed on the beta chain (Goebeler et al. 1994) demonstrated by immunofluorescent and western blotting on both naturally expressing and transfected targets. In addition, Mouse anti Human macrophages, clone MAC387 detects the beta chain in complex with the alpha.
	The antigen recognized by Mouse anti Human macrophages, clone MAC387 is expressed by granulocytes, monocytes and by tissue macrophages. Variable results have been reported for staining brain macrophages and microglia. The epitope recognized appears be well conserved and the antibody is routinely used for the detection of myeloid cells in wide range of species.
Flow Cytometry	Use 10ul of the suggested working dilution to label 10 <sup>6</sup> cells in 100ul
References	Ueland, T. et al. (2009) Dickkopf-1 enhances inflammatory interaction between platelet and endothelial cells and shows increased expression in atherosclerosis. <a href="Arterioscler Thromb Vasc Biol. 29: 1228-34">Arterioscler Thromb Vasc Biol. 29: 1228-34</a> Brandtzaeg, P. et al. (1992) The leucocyte protein L1 (calprotectin): usefulness as an

immunohistochemical marker antigen and putative biological function. <u>Histopathology. 21:</u>

### 191-6.

- 3. Gutierrez, M. *et al.* (1999) The detection of CD2+, CD4+, CD8+, and WC1+ T lymphocytes, B cells and macrophages in fixed and paraffin embedded bovine tissue using a range of antigen recovery and signal amplification techniques. <u>Vet Immunol Immunopathol.</u> 71 (3-4): 321-34.
- 4. Ramsay, A.D. *et al.* (1991) Phenotypic analysis of malignant lymphoma in simian immunodeficiency virus infection using anti-human antibodies. J Pathol. 164 (4): 321-8.
- 5. Christgau, M. *et al.* (1998) Characterization of immunocompetent cells in the diseased canine periodontium. <u>J Histochem Cytochem. 46 (12): 1443-54.</u>
- 6. Pérez, J. et al. (1999) Immunohistochemical study of the inflammatory infiltrate associated with equine squamous cell carcinoma. J Comp Pathol. 121 (4): 385-97.
- 7. Nanney, L.B. *et al.* (2008) Calreticulin enhances porcine wound repair by diverse biological effects. Am J Pathol. 173: 610-30.
- 8. Poncelet, L. *et al.* (2008) Detection of antigenic heterogeneity in feline coronavirus nucleocapsid in feline pyogranulomatous meningoencephalitis. Vet Pathol. 45: 140-53.
- 9. Sethi, R.S. *et al.* (2010) Immunolocalization of pulmonary intravascular macrophages, TLR4, TLR9 and IL-8 in normal and Pasteurella multocida-infected lungs of water buffalo (Bubalus bubalis). J Comp Pathol. 144: 135-44.
- 10. Sanchez, J. *et al.* (2011) Microscopical and immunological features of tuberculoid granulomata and cavitary pulmonary tuberculosis in naturally infected goats. <u>J Comp</u> Pathol. 145 (2-3): 107-17.
- 11. Isling, L.K. *et al.* (2010) Pyelonephritis in slaughter pigs and sows: morphological characterization and aspects of pathogenesis and aetiology. Acta Vet Scand. 52: 48.
- 12. Vranckx, K. *et al.* (2012) Vaccination reduces macrophage infiltration in bronchus-associated lymphoid tissue in pigs infected with a highly virulent Mycoplasma hyopneumoniae strain. <u>BMC Vet Res. 8: 24.</u>
- 13. Campuzano, O. *et al.* (2012) Arrhythmogenic right ventricular cardiomyopathy: severe structural alterations are associated with inflammation. <u>J Clin Pathol. 65 (12): 1077-83.</u>
- 14. García-Jiménez, W.L. (2012) Histological and immunohistochemical characterisation of *Mycobacterium bovis* induced granulomas in naturally infected fallow deer (*Dama dama*). Vet Immunol Immunopathol. 149: 66-75.
- 15. Santana, C.H. *et al.* (2016) Relationship Between the Inflammatory Infiltrate and the Degree of Differentiation of the Canine Cutaneous Cell Carcinoma. <u>Vet Anim Sci. Oct 10</u> [Epub ahead of print]
- 16. Masure, D. *et al.* (2013) A Role for Eosinophils in the Intestinal Immunity against Infective *Ascaris suum* Larvae. <u>PLoS Negl Trop Dis. 2013 Mar;7(3): e2138.</u>
- 17. Tellez, A. *et al.* (2014) Experimental evaluation of efficacy and healing response of everolimus-eluting stents in the familial hypercholesterolemic swine model: a comparative study of bioabsorbable versus durable polymer stent platforms. <u>Coron Artery Dis. 25 (3):</u> 198-207.
- 18. Collin, N. *et al.* (2009) Sand fly salivary proteins induce strong cellular immunity in a natural reservoir of visceral leishmaniasis with adverse consequences for *Leishmania*. PLoS Pathog. 5(5):e1000441.
- 19. McCurdy, P. *et al.* (2014) Acute lymphoblastic leukemia in a pygmy hippopotamus (*Hexaprotodon liberiensis*). J Zoo Wildl Med. 45 (4): 906-10.
- 20. Marcaccini, A. *et al.* (2008) Pseudorabies virus infection in mink: a host-specific pathogenesis. <u>Vet Immunol Immunopathol. 124 (3-4): 264-73.</u>

- 21. Romero-Palomo, F. *et al.* (2015) Immunopathologic Changes in the Thymus of Calves Pre-infected with BVDV and Challenged with BHV-1. <u>Transbound Emerg Dis. Aug 25.</u> [Epub ahead of print]
- 22. Rossi, C.N. *et al.* (2016) *In situ* Cutaneous cellular immune response in dogs naturally infected by visceral leishmaniasis. Rev Inst Med Trop Sao Paulo. 58: .
- 23. Vrolyk V *et al.* (2016) Lung Inflammation Associated With Clinical Acute Necrotizing Pancreatitis in Dogs. Vet Pathol. May 11. pii: 0300985816646432. [Epub ahead of print]
- 24. Nelson, M. *et al.* (2014) Comparative experimental subcutaneous glanders and melioidosis in the common marmoset (*Callithrix jacchus*). <u>Int J Exp Pathol. 95 (6): 378-91.</u>
- 25. Amarilla, S.P. *et al.* (2016) Thymic depletion of lymphocytes is associated with the virulence of PRRSV-1 strains. Vet Microbiol. 188: 47-58.
- 26. García-Jiménez, W.L. *et al.* (2013) Immunopathology of granulomas produced by *Mycobacterium bovis* in naturally infected wild boar. <u>Vet Immunol Immunopathol. 156</u> (1-2): 54-63.
- 27. Pilling, D. *et al.* (2015) The long pentraxin PTX3 promotes fibrocyte differentiation. PLoS One. 10 (3): e0119709.
- 28. Zhao, L. *et al.* (2020) Reducing macrophage numbers alleviates temporomandibular joint ankylosis. <u>Cell Tissue Res. 379 (3): 521-36.</u>
- 29. Lai, H.Y. *et al.* (2017) CCAAT/enhancer-binding protein delta promotes intracellular lipid accumulation in M1 macrophages of vascular lesions. <u>Cardiovasc Res. 113 (11):</u> 1376-88.
- 30. Wacinski, P. *et al.* (2021) Anti-Inflammatory Effect of Very High Dose Local Vessel Wall Statin Administration: Poly(L,L-Lactide) Biodegradable Microspheres with Simvastatin for Drug Delivery System (DDS). <u>Int J Mol Sci. 22 (14)Jul 13 [Epub ahead of print].</u>
- 31. Edwards, J.H. *et al.* (2021) Integration and functional performance of a decellularised porcine superflexor tendon graft in an ovine model of anterior cruciate ligament reconstruction. Biomaterials. 279: 121204.
- 32. Bertolo, P.H.L. *et al.* (2022) Influence of serum progesterone levels on the inflammatory response of female dogs with visceral leishmaniosis. <u>Vet Parasitol. 302:</u> 109658.
- 33. do Prado Duzanski, A. *et al.* (2022) Cell-mediated immunity and expression of MHC class I and class II molecules in dogs naturally infected by canine transmissible venereal tumor: Is there complete spontaneous regression outside the experimental CTVT?

  Research in Veterinary Science. 145: 193-204.

#### **Further Reading**

- 1. Burk, J. *et al.* (2013) Equine cellular therapy--from stall to bench to bedside? <u>Cytometry</u> A. 83 (1): 103-13.
- 2. Piriou-Guzylack, L. (2008) Membrane markers of the immune cells in swine: an update. Vet Res. 39: 54.

### Storage

Store at +4°C. DO NOT FREEZE.

This product should be stored undiluted. This product is photosensitive and should be protected from light. Should this product contain a precipitate we recommend microcentrifugation before use.

## Guarantee

12 months from date of despatch

### **Health And Safety**

Material Safety Datasheet documentation #20487 available at:

Information https://www.bio-rad-antibodies.com/SDS/MCA874APC

20487

Regulatory For research purposes only

## Related Products

## **Recommended Negative Controls**

MOUSE IgG1 NEGATIVE CONTROL:APC (MCA928APC)

## **Recommended Useful Reagents**

**HUMAN SEROBLOCK (BUF070A) HUMAN SEROBLOCK (BUF070B)** 

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