

Datasheet: MCA834A488

Description:	MOUSE ANTI BOVINE CD4:Alexa Fluor® 488
Specificity:	CD4
Format:	ALEXA FLUOR® 488
Product Type:	Monoclonal Antibody
Clone:	CC30
Isotype:	IgG1
Quantity:	100 TESTS/1ml

Product Details

Applications

This product has been reported to work in the following applications. This information is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for further information. For general protocol recommendations, please visit www.bio-rad-antibodies.com/protocols.

	Yes	No	Not Determined	Suggested Dilution
Flow Cytometry	•			Neat - 1/10

Where this antibody has not been tested for use in a particular technique this does not necessarily exclude its use in such procedures. Suggested working dilutions are given as a guide only. It is recommended that the user titrates the antibody for use in their own system using appropriate negative/positive controls.

Target Species	Bovine						
Species Cross Reactivity	reactivity is derived	N.B. Antibody reactivity and working conditions may vary between species. Cross reactivity is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for					
Product Form	Purified IgG conjug	ated to Alexa Fluor 488	- liquid				
Max Ex/Em	Fluorophore Alexa Fluor®488	Excitation Max (nm) 495	Emission Max (nm) 519				
Preparation	Purified IgG prepar supernatant	ed by affinity chromatog	raphy on Protein A from tissue culture				
Buffer Solution	Phosphate buffered	d saline					

Preservative	0.09% Sodium Azide (NaN ₃)		
Stabilisers	1% Bovine Serum Albumin		
Approx. Protein Concentrations	IgG concentration 0.05 mg/ml		
Immunogen	Bovine thymocytes.		
External Database Links	UniProt: A7YY52 Related reagents		
Fusion Partners	Spleen cells from an immunized mouse were fused with cells of the mouse NS1 myeloma cell line.		
Specificity	Mouse anti Bovine CD4 antibody, clone CC30 recognizes a ~50 kDa transmembrane molecule considered to be the bovine homologue of human CD4. The phenotype, tissue distribution and function of T-cells expressing the bovine CD4 antigen are similar to those in other species. However, expression on macrophages has not yet been detected.		
	Mouse anti Bovine CD4, clone CC30 has successfully been used for immunohistochemical localization of CD4 on paraffin embedded material using zinc salt fixation (Cantón et al. 2013). Additionally, clone CC30 has been reported as being suitable for use on formal dichromate (FD5) fixed paraffin embedded tissue with amplification and antigen retrieval techniques (Gutierrez et al. 1999).		
Flow Cytometry	Use 10ul of the suggested working dilution to label 10 ⁶ cells in 100ul.		
References	 Bensaid, A. & Hadam, M. (1991) Individual antigens of cattle. Bovine CD4 (BoCD4). <u>Vet Immunol Immunopathol. 27 (1-3): 51-4.</u> Eskra, L. <i>et al.</i> (1991) Effect of monoclonal antibodies on <i>in vitro</i>. function of T-cell subsets. <u>Vet Immunol Immunopathol. 27 (1-3): 215-22.</u> Gutierrez, M. <i>et al.</i> (1999) The detection of CD2+, CD4+, CD8+, and WC1+ T lymphocytes, B cells and macrophages in fixed and paraffin embedded bovine tissue using a range of antigen recovery and signal amplification techniques. <u>Vet Immunol Immunopathol. 71 (3-4): 321-34.</u> Winkler, M.T. <i>et al.</i> (1999) Bovine herpesvirus 1 can infect CD4(+) T lymphocytes and 		

- 4. Winkler, M.T. *et al.* (1999) Bovine herpesvirus 1 can infect CD4(+) T lymphocytes and induce programmed cell death during acute infection of cattle. J Virol. 73 (10): 8657-68.
- 5. Winkler, M.T. *et al.* (2000) Persistence and reactivation of bovine herpesvirus 1 in the tonsils of latently infected calves. J Virol. 74 (11): 5337-46.
- 6. Riondato, F. *et al.* (2008) Effects of road transportation on lymphocyte subsets in calves <u>Vet J. 175: 364-8.</u>
- 7. Collins, R.A. *et al.* (1999) Bovine interleukin-12 and modulation of IFNgamma production. Vet Immunol Immunopathol. 68: 193-207.
- 8. Liebana, E. *et al.* (2007) Distribution and activation of T-lymphocyte subsets in tuberculous bovine lymph-node granulomas. Vet Pathol. 44: 366-72.
- 9. Sühwold, A. *et al.* (2010) T cell reactions of *Eimeria bovis* primary and challenge-infected calves. Parasitol Res. 106: 595-605.

- 10. Rhodes, S.G. *et al.* (2000) Bovine tuberculosis: immune responses in the peripheral blood and at the site of active disease. Immunology. 99: 195-202.
- 11. Machugh, N.D. *et al.* (1997) Identification of two distinct subsets of bovine gamma delta T cells with unique cell surface phenotype and tissue distribution. <u>Immunology. 92:</u> 340-5.
- 12. Childerstone, A.J. *et al.* (1999) Demonstration of bovine CD8+ T-cell responses to foot-and-mouth disease virus. J Gen Virol. 80: 663-9.
- 13. Rhodes, S.G. *et al.* (2001) Antigen recognition and immunomodulation by gamma delta T cells in bovine tuberculosis. <u>J Immunol. 166: 5604-10.</u>
- 14. Knowles, G. *et al.* (1996) Phenotypical characterization of lymphocytes infiltrating regressing papillomas. J Virol. 70: 8451-8.
- 15. Smyth, A.J. *et al.* (2001) *In vitro* responsiveness of gammadelta T cells from *Mycobacterium bovis*-infected cattle to mycobacterial antigens: predominant involvement of WC1(+) cells. <u>Infect Immun. 69: 89-96.</u>
- 16. Hein, W.R. and Dudler, L. (1997) TCR gamma delta+ cells are prominent in normal bovine skin and express a diverse repertoire of antigen receptors. lmmunology.91:58-64.
- 17. Juleff, N. *et al.* (2009) Foot-and-mouth disease virus can induce a specific and rapid CD4+ T-cell-independent neutralizing and isotype class-switched antibody response in naïve cattle. J Virol. 83: 3626-36.
- 18. Hope, J.C. *et al.* (2005) Exposure to *Mycobacterium avium* induces low-level protection from *Mycobacterium bovis* infection but compromises diagnosis of disease in cattle. Clin Exp Immunol. 141: 432-9.
- 19. Sopp, P. *et al.* (2006) Flow cytometric detection of gamma interferon can effectively discriminate *Mycobacterium bovis* BCG-vaccinated cattle from *M. bovis*-infected cattle. Clin Vaccine Immunol. 13: 1343-8.
- 20. Sacchini, F. *et al.* (2011) A minor role of CD4+ T lymphocytes in the control of a primary infection of cattle with *Mycoplasma mycoides* subsp. *mycoides*. Vet Res. 42: 77.
- 21. Skyberg, J.A. *et al.* (2011) Murine and bovine γδ T cells enhance innate immunity against *Brucella abortus* infections. <u>PLoS One. 6(7): e21978.</u>
- 22. Cantón, G.J. *et al.* (2013) Phenotypic characterisation of the cellular immune infiltrate in placentas of cattle following experimental inoculation with *Neospora caninum* in late gestation. <u>Vet Res. 44: 60.</u>
- 23. Reid, E. *et al.* (2011) Bovine plasmacytoid dendritic cells are the major source of type I interferon in response to foot-and-mouth disease virus *in vitro* and *in vivo*. <u>J Virol. 85:</u> 4297-308.
- 24. Arranz-Solís D *et al.* (2016) Systemic and local immune responses in sheep after *Neospora caninum* experimental infection at early, mid and late gestation. <u>Vet Res. 47 (1): 2.</u>
- 25. Clapp, B. *et al.* (2011) DNA vaccination of bison to brucellar antigens elicits elevated antibody and IFN-γ responses. <u>J Wildl Dis. 47 (3): 501-10.</u>
- 26. Aranday-Cortes, E. *et al.* (2013) Upregulation of IL-17A, CXCL9 and CXCL10 in early-stage granulomas induced by *Mycobacterium bovis* in cattle. <u>Transbound Emerg</u> Dis. 60 (6): 525-37.
- 27. Pirson, C. *et al.* (2015) Highly purified mycobacterial phosphatidylinositol mannosides drive cell-mediated responses and activate NKT cells in cattle. <u>Clin Vaccine Immunol. 22</u> (2): 178-84.
- 28. Fuertes, M. et al. (2015) Immunohistochemical study and mRNA cytokine profile of the

local immune response in cattle naturally infected with *Calicophoron daubneyi*. <u>Vet</u> Parasitol. 214 (1-2): 178-83.

- 29. Çomakli, S. & Özdemir, S. (2019) Comparative Evaluation of the Immune Responses in Cattle Mammary Tissues Naturally Infected with Bovine Parainfluenza Virus Type 3 and Bovine Alphaherpesvirus-1. <u>Pathogens.8 (1): 26.</u>
- 30. Bozkurt, Y.A. *et al.* (2019) The architecture of the lymph nodes in the abdominal and thoracic cavities of wild boar <u>Ind J Anim Res. 53 609-15.</u>
- 31. Hecker, Y.P. *et al.* (2015) Cell mediated immune responses in the placenta following challenge of vaccinated pregnant heifers with *Neospora caninum*. <u>Vet Parasitol. 214 (3-4):</u> 247-54.
- 32. Cunha, P. *et al.* (2019) Expansion, isolation and first characterization of bovine Th17 lymphocytes. Sci Rep. 9 (1): 16115.
- 33. Sirak, A. *et al.* (2021) Cellular and Cytokine Responses in Lymph Node Granulomas of Bacillus Calmette Guérin (BCG)-Vaccinated and Non-vaccinated Cross-Breed Calves Naturally Infected With *Mycobacterium bovis*. <u>Front Vet Sci. 8: 698800</u>.
- 34. Korbonits, L. *et al.* (2022) Mycobacterium avium subsp. paratuberculosis Infected Cows Reveal Divergent Immune Response in Bovine Peripheral Blood Derived Lymphocyte Proteome. <u>Metabolites. 12 (10): 924.</u>
- 35. Andrés, S. *et al.* (2024) Essential oil supplementation in milk replacers: short- and long-term impacts on feed efficiency, the faecal microbiota and the plasma metabolome in dairy calves. <u>J Dev Orig Health Dis.: 1-11.</u>
- 36. Zhang, B. *et al.* (2025) Fatty acids promote migration of CD4(+) T cells through calcium release-activated calcium modulator ORAI1 sensitive glycolysis in dairy cows. <u>J</u> Dairy Sci. 108 (1): 856-67.
- 37. Zhang, B. *et al.* (2025) Calcium Release-Activated Calcium Modulator ORAI1-Sensitive Serine Dehydratase Regulates Fatty Acid-Induced CD4+ Th17/Treg Imbalance in Dairy Cows Animals. 15 (3): 388.

Storage

This product is shipped at ambient temperature. It is recommended to aliquot and store at -20°C on receipt. When thawed, aliquot the sample as needed. Keep aliquots at 2-8°C for short term use (up to 4 weeks) and store the remaining aliquots at -20°C.

Avoid repeated freezing and thawing as this may denature the antibody. Storage in frost-free freezers is not recommended.

Guarantee

12 months from date of despatch

Acknowledgements

This product is provided under an intellectual property licence from Life Technologies Corporation. The transfer of this product is contingent on the buyer using the purchased product solely in research, excluding contract research or any fee for service research, and the buyer must not sell or otherwise transfer this product or its components for (a) diagnostic, therapeutic or prophylactic purposes; (b) testing, analysis or screening services, or information in return for compensation on a per-test basis; (c) manufacturing or quality assurance or quality control, or (d) resale, whether or not resold for use in research. For information on purchasing a license to this product for purposes other than as described above, contact Life Technologies Corporation, 5791 Van Allen Way, Carlsbad CA 92008 USA or outlicensing@thermofisher.com

Health And Safety Material Safety Datasheet documentation #10041 available at:

Information

https://www.bio-rad-antibodies.com/SDS/MCA834A488

10041

Regulatory For research purposes only

Related Products

Recommended Negative Controls

MOUSE IgG1 NEGATIVE CONTROL: Alexa Fluor® 488 (MCA928A488)

North & South Tel: +1 800 265 7376 Worldwide Tel: +44 (0)1865 852 700 Tel: +49 (0) 89 8090 95 21 Europe Fax: +44 (0)1865 852 739 America Fax: +1 919 878 3751 Fax: +49 (0) 89 8090 95 50

> Email: antibody_sales_us@bio-rad.com Email: antibody_sales_uk@bio-rad.com Email: antibody_sales_de@bio-rad.com

To find a batch/lot specific datasheet for this product, please use our online search tool at: bio-rad-antibodies.com/datasheets 'M405646:220916'

Printed on 26 Feb 2025

© 2025 Bio-Rad Laboratories Inc | Legal | Imprint