

## Datasheet: MCA81SBB765

<b>Description:</b>	MOUSE ANTI HUMAN HLA ABC:StarBright Blue 765
<b>Specificity:</b>	HLA ABC
<b>Format:</b>	StarBright Blue 765
<b>Product Type:</b>	Monoclonal Antibody
<b>Clone:</b>	W6/32
<b>Isotype:</b>	IgG2a
<b>Quantity:</b>	100 TESTS/0.5ml

## Product Details

### Applications

This product has been reported to work in the following applications. This information is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for further information. For general protocol recommendations, please visit [www.bio-rad-antibodies.com/protocols](http://www.bio-rad-antibodies.com/protocols).

	Yes	No	Not Determined	Suggested Dilution
Flow Cytometry	▪			Neat

Where this product has not been tested for use in a particular technique this does not necessarily exclude its use in such procedures. Suggested working dilutions are given as a guide only. It is recommended that the user titrates the product for use in their own system using appropriate negative/positive controls.

**Target Species** Human

**Species Cross Reactivity** Reacts with: Macaque, Bovine, Cynomolgus monkey, Baboon, Rhesus Monkey, Chimpanzee, Gorilla, Shrew  
Does not react with: Goat, Dog, Guinea Pig, Rabbit, Mouse, Chicken, Amphibia  
**N.B.** Antibody reactivity and working conditions may vary between species. Cross reactivity is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for further information.

**Product Form** Purified IgG conjugated to StarBright Blue 765 - liquid

Max Ex/Em	Fluorophore	Excitation Max (nm)	Emission Max (nm)
	StarBright Blue 765	476	764

**Preparation** Purified IgG prepared by affinity chromatography on Protein G from tissue culture supernatant

<b>Buffer Solution</b>	Phosphate buffered saline
<b>Preservative Stabilisers</b>	0.09% Sodium Azide (NaN <sub>3</sub> ) 1% Bovine Serum Albumin 0.1% Pluronic F68 0.1% PEG 3350 0.05% Tween 20
<b>Immunogen</b>	Purified human tonsil lymphocyte membranes.
<b>Fusion Partners</b>	Spleen cells from immunized BALB/c mice were fused with cells of the mouse NS1/1-Ag4.1 myeloma cell line.
<b>Specificity</b>	<p><b>Mouse anti Human HLA ABC antibody, clone W6/32</b> recognizes an antigenic determinant shared among products of the HLA A, B and C loci. Clone W6/32 recognizes a conformational epitope, reacting with HLA class I alpha3 and alpha2 domains. The major histocompatibility complex (MHC) is a cluster of genes that are important in the immune response to infections. In humans, this complex is referred to as the human leukocyte antigen (HLA) region. There are 3 major MHC class I proteins encoded by the HLA which are HLA A, HLA B and HLA C. These proteins are found on the surface of almost all nucleated somatic cells.</p> <p>Mouse anti Human HLA ABC antibody, clone W6/32 is routinely tested in flow cytometry on human peripheral blood lymphocytes.</p>
<b>Flow Cytometry</b>	Use 5ul of the suggested working dilution to label 10 <sup>6</sup> cells in 100ul. Best practices suggest a 5 minutes centrifugation at 6,000g prior to sample application.
<b>References</b>	<ol style="list-style-type: none"> <li>1. Brodsky, F.M. &amp; Parham, P. (1982) Evolution of HLA antigenic determinants: species cross-reactions of monoclonal antibodies. <a href="#">Immunogenetics. 15 (2): 151-66.</a></li> <li>2. Neefjes, J.J. <i>et al.</i> (1986) A biochemical characterization of feline MHC products: unusually high expression of class II antigens on peripheral blood lymphocytes. <a href="#">Immunogenetics. 23 (5): 341-7.</a></li> <li>3. Stern, P.L. <i>et al.</i> (1987) Class I-like MHC molecules expressed by baboon placental syncytiotrophoblast. <a href="#">J Immunol. 138 (4): 1088-91.</a></li> <li>4. Jacobsen, C.N. <i>et al.</i> (1993) Reactivities of 20 anti-human monoclonal antibodies with leucocytes from ten different animal species. <a href="#">Vet Immunol Immunopathol. 39 (4): 461-6.</a></li> <li>5. Verbeek, M.M. <i>et al.</i> (1995) T lymphocyte adhesion to human brain pericytes is mediated via very late antigen-4/vascular cell adhesion molecule-1 interactions. <a href="#">J Immunol. 154 (11): 5876-84.</a></li> <li>6. Raftery, M.J. <i>et al.</i> (2002) Hantavirus infection of dendritic cells. <a href="#">J Virol. 76: 10724-33.</a></li> <li>7. Dressel, R. <i>et al.</i> (2003) Differential effect of acute and permanent heat shock protein 70 overexpression in tumor cells on lysability by cytotoxic T lymphocytes. <a href="#">Cancer Res. 63 (23): 8212-20.</a></li> <li>8. Ishitani, A. <i>et al.</i> (2003) Protein expression and peptide binding suggest unique and interacting functional roles for HLA-E, F, and G in maternal-placental immune recognition. <a href="#">J Immunol. 171 (3): 1376-84.</a></li> <li>9. Giuliani, F. <i>et al.</i> (2003) Vulnerability of human neurons to T cell-mediated cytotoxicity. <a href="#">J</a></li> </ol>

[Immunol. 171: 368-79.](#)

10. Smith, A.C. *et al.* (2005) Interaction of the *Salmonella*-containing vacuole with the endocytic recycling system. [J Biol Chem. 280: 24634-41.](#)
11. Zuo, J. *et al.* (2008) The DNase of gammaherpesviruses impairs recognition by virus-specific CD8+ T cells through an additional host shutoff function. [J Virol. 82: 2385-93.](#)
12. Shi, J. *et al.* (2008) Bortezomib down-regulates the cell-surface expression of HLA class I and enhances natural killer cell-mediated lysis of myeloma. [Blood. 111: 1309-17.](#)
13. Watling, D. *et al.* (2008) Multiple kinases in the interferon-gamma response. [Proc Natl Acad Sci U S A. 105: 6051-6.](#)
14. Grotzke, J.E. *et al.* (2009) The *Mycobacterium tuberculosis* phagosome is a HLA-I processing competent organelle. [PLoS Pathog. 5: e1000374.](#)
15. Koch, N. *et al.* (2009) IL-10 protects monocytes and macrophages from complement-mediated lysis. [J Leukoc Biol. 86: 155-66.](#)
16. Spentzou, A. *et al.* (2010) Viral inhibition assay: a CD8 T cell neutralization assay for use in clinical trials of HIV-1 vaccine candidates. [J Infect Dis. 201: 720-9.](#)
17. Vitadello, M. *et al.* (2010) Myofiber stress-response in myositis: parallel investigations on patients and experimental animal models of muscle regeneration and systemic inflammation. [Arthritis Res Ther. 12: R52.](#)
18. Hinrichs, J. *et al.* (2010) The nature of peptides presented by an HLA class I low expression allele. [Haematologica. 95: 1373-80.](#)
19. Fujita, Y. *et al.* (2010) Bone marrow transplantation restores epidermal basement membrane protein expression and rescues epidermolysis bullosa model mice. [Proc Natl Acad Sci U S A. 107: 14345-50.](#)
20. Narita, M. *et al.* (2010) WT1 peptide vaccination in combination with imatinib therapy for a patient with CML in the chronic phase. [Int J Med Sci. 7: 72-81.](#)
21. Zuo, J. *et al.* (2011) The Epstein-Barr virus-encoded BILF1 protein modulates immune recognition of endogenously processed antigen by targeting major histocompatibility complex class I molecules trafficking on both the exocytic and endocytic pathways. [J Virol. 85: 1604-14.](#)
22. Lask, A. *et al.* (2011) TCR-Independent Killing of B Cell Malignancies by Anti-Third-Party CTLs: The Critical Role of MHC-CD8 Engagement. [J Immunol. 187: 2006-14.](#)
23. Jones, D.C. *et al.* (2011) HLA Class I Allelic Sequence and Conformation Regulate Leukocyte Ig-Like Receptor Binding. [J Immunol. 186: 2990-7.](#)
24. Enose-Akahata, Y. *et al.* (2012) Minocycline modulates antigen-specific CTL activity through inactivation of mononuclear phagocytes in patients with HTLV-I associated neurologic disease. [Retrovirology. 9: 16.](#)
25. Badrinath, S. *et al.* (2012) Position 156 influences the peptide repertoire and tapasin dependency of human leukocyte antigen B\*44 allotypes. [Haematologica. 97: 98-106.](#)
26. Tannetta, D.S. *et al.* (2013) Characterisation of syncytiotrophoblast vesicles in normal pregnancy and pre-eclampsia: expression of Flt-1 and endoglin. [PLoS One. 8 \(2\): e56754.](#)
27. Dragovic, R.A. *et al.* (2015) Isolation of syncytiotrophoblast microvesicles and exosomes and their characterisation by multicolour flow cytometry and fluorescence Nanoparticle Tracking Analysis. [Methods. 87: 64-74.](#)
28. Tischer, S. *et al.* (2016) Discovery of immunodominant T-cell epitopes reveals penton protein as a second immunodominant target in human adenovirus infection. [J Transl Med. 14 \(1\): 286.](#)

29. Praest, P. *et al.* (2019) A Flow Cytometry-Based Approach to Unravel Viral Interference with the MHC Class I Antigen Processing and Presentation Pathway. [Methods Mol Biol. 1988: 187-98.](#)
30. Juan, C.H. *et al.* (2020) *In Vitro* Differentiation of Human Placenta-Derived Multipotent Cells into Schwann-Like Cells. [Biomolecules. 10 \(12\): 1657.](#)
31. Tupova, L. *et al.* (2020) Interplay of drug transporters P-glycoprotein (MDR1), MRP1, OATP1A2 and OATP1B3 in passage of maraviroc across human placenta. [Biomed Pharmacother. 129: 110506.](#)
32. Nguyen, J. *et al.* (2021) Quantitative contributions of TNF receptor superfamily members to CD8<sup>+</sup> T-cell responses. [Mol Syst Biol. 17 \(11\): e10560.](#)
33. Xia, P. *et al.* (2022) NLRP3 inflammasome up-regulates major histocompatibility complex class I expression and promotes inflammatory infiltration in polymyositis. [BMC Immunol. 23 \(1\): 39.](#)
34. Cacciola, R. *et al.* (2022) Impact of Anti-Endothelial Cell Antibodies (AECAs) in Patients with Polycythemia Vera and Thrombosis [Diagnostics. 12 \(5\): 1077.](#)
35. Korbonits, L. *et al.* (2022) *Mycobacterium avium* subsp. *paratuberculosis* Infected Cows Reveal Divergent Immune Response in Bovine Peripheral Blood Derived Lymphocyte Proteome. [Metabolites.12 \(10\): 924.](#)
36. Lyssy, F. *et al.* (2023) Platelet-derived factors dysregulate placental sphingosine-1-phosphate receptor 2 in human trophoblasts. [Reprod Biomed Online. 47 \(2\): 103215.](#)
37. Forstner, D. *et al.* (2023) CD39 abrogates platelet-derived factors induced IL-1 $\beta$  expression in the human placenta. [Front Cell Dev Biol. 11: 1183793.](#)
38. Zheng, S. *et al.* (2022) The SARS-CoV-2 accessory factor ORF7a downregulates MHC class I surface expression. [bioRxiv 30 May \[Epub ahead of print. Preprint\]](#)
39. Fielding, C.A. *et al.* (2022) SARS-CoV-2 host-shutoff impacts innate NK cell functions, but antibody-dependent NK activity is strongly activated through non-spike antibodies. [11: e74489.](#)

<b>Storage</b>	Store at +4°C. DO NOT FREEZE. This product should be stored undiluted.
<b>Guarantee</b>	12 months from date of despatch
<b>Acknowledgements</b>	This product is covered by U.S. Patent No. 10,150,841 and related U.S. and foreign counterparts
<b>Health And Safety Information</b>	Material Safety Datasheet documentation #20471 available at: <a href="https://www.bio-rad-antibodies.com/SDS/MCA81SBB765">https://www.bio-rad-antibodies.com/SDS/MCA81SBB765</a> 20471
<b>Regulatory</b>	For research purposes only

## Related Products

### Recommended Useful Reagents

[HUMAN SEROBLOCK \(BUF070A\)](#)

[HUMAN SEROBLOCK \(BUF070B\)](#)

**North & South** Tel: +1 800 265 7376

**Worldwide** Tel: +44 (0)1865 852 700

**Europe** Tel: +49 (0) 89 8090 95 21

**America** Fax: +1 919 878 3751

Fax: +44 (0)1865 852 739

Fax: +49 (0) 89 8090 95 50

Email: [antibody\\_sales\\_us@bio-rad.com](mailto:antibody_sales_us@bio-rad.com)

Email: [antibody\\_sales\\_uk@bio-rad.com](mailto:antibody_sales_uk@bio-rad.com)

Email: [antibody\\_sales\\_de@bio-rad.com](mailto:antibody_sales_de@bio-rad.com)

To  
find a

batch/lot specific datasheet for this product, please use our online search tool at: [bio-rad-antibodies.com/datasheets](https://bio-rad-antibodies.com/datasheets)

'M409990:221024'

**Printed on 08 Mar 2024**

---

© 2024 Bio-Rad Laboratories Inc | [Legal](#) | [Imprint](#)