

Datasheet: MCA643PE BATCH NUMBER 156540

Description:	MOUSE ANTI RAT CD44:RPE
Specificity:	CD44
Other names:	H-CAM, PGP-1
Format:	RPE
Product Type:	Monoclonal Antibody
Clone:	OX-50
lsotype:	lgG1
Quantity:	100 TESTS

Product Details

Applications	This product has been reported to work in the following applications. This information is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for further information. For general protocol recommendations, please visit <u>www.bio-</u>					
	rad-antibodies.com/protocols.					
		Yes	No	Not Determined	Suggested Dilution	
	Flow Cytometry	•			Neat	
	Where this antibody h necessarily exclude its a guide only. It is reco system using appropri	s use in such mmended tha	procedu It the use	res. Suggested workin er titrates the antibody	g dilutions are given as	
Target Species	Rat					
Product Form	Purified IgG conjugate	ed to R. Phyco	berythrin	(RPE) - lyophilized		
Reconstitution	Reconstitute with 1 ml	l distilled wate	r			
Max Ex/Em	Fluorophore	Excitation M	ax (nm)	Emission Max (nm)		
	RPE 488nm laser	496		578		
Preparation	Purified IgG prepared by affinity chromatography on Protein A from tissue culture supernatant					
Buffer Solution	Phosphate buffered sa	aline				
Preservative Stabilisers	0.09% Sodium Azide 1% Bovine Serum	Albumin				

	5% Sucrose
Immunogen	Rat T cell blasts.
External Database Links	UniProt: <u>P26051</u> <u>Related reagents</u> Entrez Gene: <u>25406</u> Cd44 <u>Related reagents</u>
RRID	AB_321723
Fusion Partners	Spleen cells from immunised BALB/c mice were fused with cells of the mouse NS0/1 myeloma cell line.
Specificity	Mouse anti Rat CD44 antibody, clone OX-50 recognizes the rat CD44 cell surface antigen, also known as Extracellular matrix receptor III, P90 lymphocyte homing/adhesion receptor, HUTCH-I, Hermes antigen, Hyaluronate receptor, Phagocytic glycoprotein 1, PGP-1 or Phagocytic glycoprotein I.
	CD44 is a 482 amino acid ~85 kDa single pass type I transmembrane glycoprotein, expressed by T cells, B cells, macrophages and thymocytes, with expression being increased following activation.
Flow Cytometry	Use 10ul of the suggested working dilution to label 10 ⁶ cells in 100ul.
References	 Stevenson, K.S. <i>et al.</i> (2009) Isolation, characterization, and differentiation of thy1.1-sorted pancreatic adult progenitor cell populations. <u>Stem Cells Dev. 18:1389-98.</u> Jiang, T.S. <i>et al.</i> (2010) Reconstruction of the corneal epithelium with induced marrow mesenchymal stem cells in rats. <u>Mol Vis. 16: 1304-16.</u> Kanellis, J. <i>et al.</i> (2010) JNK signalling in human and experimental renal ischaemia/reperfusion injury. <u>Nephrol Dial Transplant. 25: 2898-908.</u> Li, S. <i>et al.</i> (2010) Upregulation of CXCR4 favoring neural-like cells migration via AKT activation. <u>Neurosci Res. 67: 293-9.</u> Stephens, L.A. <i>et al.</i> (2004) Phenotypic characterization of regulatory CD4+CD25+ T cells in rats. <u>Int Immunol. 16: 365-75.</u> Rice, C.M. <i>et al.</i> (2010) Multipotent adult progenitor cell isolation and proliferation in cytokine and serum-free medium conditioned by rat B104 cells. <u>Br J Haematol. 148:</u> 441-4.

10. Walther, M. et al. (2001) Exogenous antigen containing perivascular phagocytes induce a non-encephalitogenic extravasation of primed lymphocytes. J Neuroimmunol. 117: 30-42.

11. Suzuki, A. et al. (2006) Localization of CD44 and hyaluronan in the synovial membrane of the rat temporomandibular joint. Anat Rec A Discov Mol Cell Evol Biol. 288: 646-52.

12. Goransson, V. et al. (2004) Renal hyaluronan accumulation and hyaluronan synthase expression after ischaemia-reperfusion injury in the rat. Nephrol Dial Transplant.19: 823-30.

13. Campbell, N.G. et al. (2016) Cell Size Critically Determines Initial Retention of Bone Marrow Mononuclear Cells in the Heart after Intracoronary Injection: Evidence from a Rat Model. PLoS One. 11 (7): e0158232.

14. Bejar, M.T. et al. (2016) Inhibition of Notch rescues the angiogenic potential impaired by cardiovascular risk factors in epicardial adipose stem cells. FASEB J. 30 (8): 2849-59. 15. Rochefort, G.Y. et al. (2006) Multipotential mesenchymal stem cells are mobilized into peripheral blood by hypoxia. Stem Cells. 24 (10): 2202-8.

16. Redondo, J. et al. (2015) Reductions in kinesin expression are associated with nitric oxide-induced axonal damage. J Neurosci Res. 93 (6): 882-92.

17. Huang, X. et al. (2019) MRI Tracking of SPIO- and Fth1-Labeled Bone Marrow Mesenchymal Stromal Cell Transplantation for Treatment of Stroke. Contrast Media Mol Imaging. 2019: 5184105.

18. Aminzadeh, A. et al. (2020) Investigating The Alterations of Oxidative Stress Status, Antioxidant Defense Mechanisms, MAP Kinase and Mitochondrial Apoptotic Pathway in Adipose-Derived Mesenchymal Stem Cells from STZ Diabetic Rats. Cell J. 22 (Suppl 1): 38-48.

19. Paiva, R.G. et al. (2020) Stem cells in end-to-side neurorrhaphy. Experimental study in rats Acta Cirúrgica Brasileira. 35 (12) [Epub ahead of print].

20. Hou, B. et al. (2018) Comparison of the Effects of BMSC-derived Schwann Cells and Autologous Schwann Cells on Remyelination Using a Rat Sciatic Nerve Defect Model. Int J Biol Sci. 14 (13): 1910-22.

21. Porwal, K. et al.kfz172 (2019) Increased bone marrow-specific adipogenesis by clofazimine causes impaired fracture healing, osteopenia and osteonecrosis without extraskeletal effects in rats. Toxicol Sci. kfz172.

22. Chang, H.H. et al. (2019) Intrarenal Transplantation of Hypoxic Preconditioned Mesenchymal Stem Cells Improves Glomerulonephritis through Anti-Oxidation, Anti-ER Stress, Anti-Inflammation, Anti-Apoptosis, and Anti-Autophagy. Antioxidants (Basel). 9 (1): <u>2.</u>

23. Wu, J. et al. (2022) Reinforcing the function of bone graft via the Ca-P ceramics dynamic behavior-enhanced osteogenic microenvironment for optimal bone regeneration and reconstruction Applied Materials Today. 27: 101465.

24. Cheng, Y.H. et al. (2022) Intrarenal Arterial Transplantation of Dexmedetomidine Preconditioning Adipose Stem-Cell-Derived Microvesicles Confers Further Therapeutic Potential to Attenuate Renal Ischemia/Reperfusion Injury through

miR-122-5p/Erythropoietin/Apoptosis Axis. Antioxidants (Basel). 11(9): 1702.

Storage

Prior to reconstitution store at +4°C. Following reconstitution store at +4°C.

DO NOT FREEZE.

	This product should be stored undiluted. This product is photosensitive and should be protected from light. Should this product contain a precipitate we recommend microcentrifugation before use.
Guarantee	12 months from date of despatch
Health And Safety Information	Material Safety Datasheet documentation #20487 available at: https://www.bio-rad-antibodies.com/SDS/MCA643PE 20487
Regulatory	For research purposes only

Related Products

Recommended Negative Controls

MOUSE IgG1 NEGATIVE CONTROL:RPE (MCA1209PE)

North & South	Tel: +1 800 265 7376	Worldwide	Tel: +44 (0)1865 852 700	Europe	Tel: +49 (0) 89 8090 95 21
America	Fax: +1 919 878 3751		Fax: +44 (0)1865 852 739		Fax: +49 (0) 89 8090 95 50
	Email: antibody_sales_us@	bio-rad.com	Email: antibody_sales_uk@bic	-rad.com	Email: antibody_sales_de@bio-rad.com

To find a batch/lot specific datasheet for this product, please use our online search tool at: bio-rad-antibodies.com/datasheets 'M376231:210126'

Printed on 14 Apr 2025

© 2025 Bio-Rad Laboratories Inc | Legal | Imprint