## Datasheet: MCA43FA BATCH NUMBER 165738

| Description:  | MOUSE ANTI RAT CD45:FITC |
|---------------|--------------------------|
| Specificity:  | CD45                     |
| Other names:  | LCA                      |
| Format:       | FITC                     |
| Product Type: | Monoclonal Antibody      |
| Clone:        | OX-1                     |
| lsotype:      | lgG1                     |
| Quantity:     | 50 µg                    |

# **Product Details**

| Applications                      | This product has been reported to work in the following applications. This information is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for further information. For general protocol recommendations, please visit <u>www.bio-rad-antibodies.com/protocols</u> . |                     |           |               |                    |  |
|-----------------------------------|--|---------------------|-----------|---------------|--------------------|--|
|                                   |  | Yes No              | o No      | t Determined  | Suggested Dilution |  |
|                                   | Flow Cytometry   | •                   |           |               | Neat - 1/10        |  |
|                                   | Where this antibody has not been tested for use in a particular technique this does not necessarily exclude its use in such procedures. Suggested working dilutions are given as a guide only. It is recommended that the user titrates the antibody for use in their own system using appropriate negative/positive controls.   |                     |           |               |                    |  |
| Target Species                    | Rat  |                     |           |               |                    |  |
| Product Form                      | Purified IgG conjugated to Fluorescein Isothiocyanate Isomer 1 (FITC) - liquid   |                     |           |               |                    |  |
| Max Ex/Em                         | Fluorophore  | Excitation Max (    | nm) Emis  | sion Max (nm) |                    |  |
|                                   | FITC   | 490                 |           | 525           |                    |  |
| Preparation                       | Antibody purified from   | i tissue culture su | pernatant |               |                    |  |
| Buffer Solution                   | Phosphate buffered saline  |                     |           |               |                    |  |
| Preservative<br>Stabilisers       | 0.09% Sodium Azide<br>1% Bovine Serum Albumin  |                     |           |               |                    |  |
| Approx. Protein<br>Concentrations | IgG concentration 0.1  | mg/ml               |           |               |                    |  |

| Immunogen         | Rat thymocyte membrane glycoproteins.   |
|-------------------|---|
| External Database |   |
| Links             | UniProt:  |
|                   | P04157 Related reagents   |
|                   | Entrez Gene:  |
|                   | 24699 Ptprc Related reagents  |
| RRID              | AB_566765   |
| Fusion Partners   | Spleen cells from immunized BALB/c mice were fused with cells of the NS1 mouse myeloma cell line.   |
| Specificity       | <b>Mouse anti Rat CD45 antibody, clone OX-1</b> recognizes CD45, also known as the leucocyte common antigen (LCA). The leucocyte common antigen consists of a family of heavily glycosylated membrane glycoproteins of molecular weight 180 – 240kDa.                   |
|                   | Antibodies recognising a common epitope on all of these isoforms are termed CD45,<br>whilst those recognising only individual isoforms are termed CD45RA, CD45RO etc. OX-1<br>reacts with all forms of CD45 expressed by all haematopoietic cells, except erythrocytes. |
|                   | CD45 isoforms play complex roles in T-cell and B-cell antigen receptor signal transduction.   |
|                   | This product is routinely tested in flow cytometry on rat splenocytes   |
| Flow Cytometry    | Use 10ul of the suggested working dilution to label 10 <sup>6</sup> cells in 100ul.   |
| References        | 1. Standring, R. et al. (1978) The predominant heavily glycosylated glycoproteins at the  |
|                   | surface of rat lymphoid cells are differentiation antigens. <u>Eur J Immunol. 8 (12): 832-9.</u>  |
|                   | 2. Sunderland, C.A. <i>et al.</i> (1979) Purification with monoclonal antibody of a predominant leukocyte-common antigen and glycoprotein from rat thymocytes. <u>Eur J Immunol. 9 (2):</u>   |
|                   | <u>155-9.</u><br>3. Woollett, G.R. <i>et al.</i> (1985) Molecular and antigenic heterogeneity of the rat leukocyte-   |
|                   | common antigen from thymocytes and T and B lymphocytes. <u>Eur J Immunol. 15 (2):</u><br><u>168-73.</u>   |
|                   | 4. Martín, A. <i>et al.</i> (1995) Passive dual immunization against tumour necrosis factor-alpha   |
|                   | (TNF-alpha) and IL-1 beta maximally ameliorates acute aminonucleoside nephrosis. Clin   |
|                   | Exp Immunol. 99 (2): 283-8.   |
|                   | 5. Giezeman-Smits, K.M. <i>et al.</i> (1999) The regulatory role of CD45 on rat NK cells in   |
|                   | target cell lysis. <u>J Immunol. 163 (1): 71-6.</u>   |
|                   | 6. Murakami, K. <i>et al.</i> (2000) Regulation of mast cell signaling through high-affinity IgE  |
|                   | receptor by CD45 protein tyrosine phosphatase. <u>Int Immunol. 12 (2): 169-76.</u>  |
|                   | 7 Front L at al (2001) Comparison of different data the most bed in months that   |
|                   | 7. Ermert, L. <i>et al.</i> (2001) Comparison of different detection methods in quantitative microdonsitemetry. Am J Pathol. 158: 407-17  |
|                   | microdensitometry. Am J Pathol. 158: 407-17.  |
|                   |   |

9. Sato, K. *et al.* (2001) Carbon monoxide generated by heme oxygenase-1 suppresses the rejection of mouse-to-rat cardiac transplants. <u>J Immunol. 166 (6): 4185-94.</u>

10. Kurozumi, K. *et al.* (2007) Effect of tumor microenvironment modulation on the efficacy of oncolytic virus therapy. <u>J Natl Cancer Inst. 99: 1768-81.</u>

11. Leonardo, C.C. *et al.* (2009) Inhibition of gelatinase activity reduces neural injury in an ex vivo model of hypoxia-ischemia. <u>Neuroscience. 160: 755-66.</u>

12. Vaschetto, R. *et al.* (2010) Renal hypoperfusion and impaired endothelium-dependent vasodilation in an animal model of VILI: the role of the peroxynitrite-PARP pathway <u>Crit</u> <u>Care. 14: R45.</u>

13. Ladhoff, J. *et al.* (2010) Immune privilege of endothelial cells differentiated from endothelial progenitor cells. <u>Cardiovasc Res. 88: 121-9.</u>

14. Jeong, H.K. *et al* (2010) Inflammatory responses are not sufficient to cause delayed neuronal death in ATP-induced acute brain injury. <u>PLoS One. 5: e13756.</u>

15. Schupp, N. *et al.* (2011) Mineralocorticoid receptor-mediated DNA damage in kidneys of DOCA-salt hypertensive rats. <u>FASEB J. 25 (3): 968-78.</u>

16. Markusic, D.M. *et al.* (2010) Separating lentiviral vector injection and induction of gene expression in time, does not prevent an immune response to rtTA in rats. <u>PLoS One. 5: e9974.</u>

17. Runesson, E. *et al.* (2015) Nucleostemin- and Oct 3/4-positive stem/progenitor cells exhibit disparate anatomical and temporal expression during rat Achilles tendon healing. <u>BMC Musculoskelet Disord. 16: 212.</u>

18. Tanner, D.C. *et al.* (2015) cFLIP is critical for oligodendrocyte protection from inflammation. <u>Cell Death Differ. 22 (9): 1489-501.</u>

Wang, C. *et al.* (2015) Small activating RNA induces myogenic differentiation of rat adipose-derived stem cells by upregulating MyoD. <u>Int Braz J Urol. 41 (4): 764-72.</u>
 Yao, Y. *et al.* (2016) Alendronate Attenuates Spinal Microglial Activation and Neuropathic Pain. J Pain. 17 (8): 889-903.

21. Collins, J.J.P. *et al.* (2018) Impaired Angiogenic Supportive Capacity and Altered Gene Expression Profile of Resident CD146<sup>+</sup> Mesenchymal Stromal Cells Isolated from Hyperoxia-Injured Neonatal Rat Lungs. <u>Stem Cells Dev. 27 (16): 1109-24.</u>

22. Porwal, K. *et al.* (2019) Increased bone marrow-specific adipogenesis by clofazimine causes impaired fracture healing, osteopenia and osteonecrosis without extra-skeletal effects in rats. <u>Toxicol Sci. kfz172.</u>

23. Hellenbrand, D.J. *et al.* (2019) Sustained interleukin-10 delivery reduces inflammation and improves motor function after spinal cord injury. <u>J Neuroinflammation. 16 (1): 93.</u>

24. Kuriyama, T. *et al.* (2020) A novel rat model of inflammatory bowel disease developed using a device created with a 3D printer. <u>Regen Ther. 14: 1-10.</u>

25. Pilipović, I. *et al.* (2020) Propranolol diminished severity of rat EAE by enhancing immunoregulatory/protective properties of spinal cord microglia. <u>Neurobiol Dis. 134:</u> 104665.

26. Dabrowska, S. *et al.* (2021) Neuroinflammation evoked by brain injury in a rat model of lacunar infarct. <u>Exp Neurol. 336: 113531.</u>

27. Elabi, O.F. *et al.* (2021) L-dopa-Dependent Effects of GLP-1R Agonists on the Survival of Dopaminergic Cells Transplanted into a Rat Model of Parkinson Disease. <u>Int J Mol Sci.</u> 22(22):12346.

28. Hou, Y. *et al.* (2021) Pseudoginsenoside-F11 promotes functional recovery after transient cerebral ischemia by regulating the microglia/macrophage polarization in rats. Int

|                                  | <ul> <li>Immunopharmacol. 99: 107896.</li> <li>29. Eweida, A. <i>et al.</i> (2022) Systemically injected bone marrow specifically home to axially vascularized tissue engineering core <u>e0272697.</u></li> <li>30. Yang, Q. <i>et al.</i> (2022) Electrospun aligned poly(ε-caprolacted guiding 3D organization of tendon stem/progenitor cells in tendor tendon repair. Front Bioeng Biotechnol. 10: 960694.</li> <li>31. Wu, Y. <i>et al.</i> (2018) Increased ceruloplasmin expression calleukocytes, activated microglia, and astrocytes in injured femal Neurosci Res. 96 (7): 1265-76.</li> </ul> | nstructs. <u>PLoS One. 17 (8):</u><br>one) nanofiber yarns<br>genic differentiation and<br>used by infiltrated |  |  |
|----------------------------------|---|--|--|--|
| Storage                          | This product is shipped at ambient temperature. It is recommended to aliquot and store at -20°C on receipt. When thawed, aliquot the sample as needed. Keep aliquots at 2-8°C for short term use (up to 4 weeks) and store the remaining aliquots at -20°C.<br>Avoid repeated freezing and thawing as this may denature the antibody. Storage in frost-free freezers is not recommended. This product is photosensitive and should be protected from light.   |  |  |  |
| Guarantee                        | 12 months from date of despatch   |  |  |  |
| Health And Safety<br>Information | Material Safety Datasheet documentation #10041 available at:<br>https://www.bio-rad-antibodies.com/SDS/MCA43FA<br>10041   |  |  |  |
| Regulatory                       | For research purposes only  |  |  |  |

## **Related Products**

### **Recommended Negative Controls**

MOUSE IgG1 NEGATIVE CONTROL:FITC (MCA1209F)

| North & South | Tel: +1 800 265 7376            | Worldwide | Tel: +44 (0)1865 852 700       | Europe | Tel: +49 (0) 89 8090 95 21           |
|---------------|---------------------------------|-----------|--------------------------------|--------|--------------------------------------|
| America       | Fax: +1 919 878 3751            |           | Fax: +44 (0)1865 852 739       |        | Fax: +49 (0) 89 8090 95 50           |
|               | Email: antibody_sales_us@bio-ra | ad.com    | Email: antibody_sales_uk@bio-r | ad.com | Email: antibody_sales_de@bio-rad.com |

To find a batch/lot specific datasheet for this product, please use our online search tool at: bio-rad-antibodies.com/datasheets 'M405523:220916'

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