

# Datasheet: MCA342R BATCH NUMBER 158552

Description:	MOUSE ANTI RAT CD163
Specificity:	CD163
Other names:	ED2
Format:	Purified
Product Type:	Monoclonal Antibody
Clone:	ED2
Isotype:	lgG1
Quantity:	0.25 mg

### **Product Details**

#### **Applications**

This product has been reported to work in the following applications. This information is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for further information. For general protocol recommendations, please visit <a href="www.bio-rad-antibodies.com/protocols">www.bio-rad-antibodies.com/protocols</a>.

	Yes	No	Not Determined	Suggested Dilution
Flow Cytometry	•			
Immunohistology - Frozen	•			1/50 - 1/100
Immunohistology - Paraffin (1)	-			
ELISA				
Immunoprecipitation				
Western Blotting	-			
Immunofluorescence				

Where this antibody has not been tested for use in a particular technique this does not necessarily exclude its use in such procedures. Suggested working dilutions are given as a guide only. It is recommended that the user titrates the antibody for use in their own system using appropriate negative/positive controls.

## (1) This product requires protein digestion pre-treatment of paraffin sections e.g. trypsin or pronase

Target Species	Rat	
Product Form	Purified IgG - liquid	
Preparation	Purified IgG prepared by affinity chromatography on Protein A supernatant	from tissue culture

Buffer Solution	Phosphate buffered saline
Preservative Stabilisers	0.09% Sodium Azide
Carrier Free	Yes
Approx. Protein Concentrations	IgG concentration 0.5 mg/ml
Immunogen	Rat spleen cell homogenate.
RRID	AB_321966
Fusion Partners	Spleen cells from immunized BALB/c mice were fused with cells of the SP2/0-Ag 14 mouse myeloma cell line.
Specificity	<b>Mouse anti Rat CD163, clone ED2</b> recognizes the rat ED2 cell surface glycoprotein ( <u>Dijkstra et al. 1985</u> ). A 175 kDa molecule also known as rat CD163, a member of the group B scavenger receptor cysteine-rich (SRCR) family and an erythroblast adhesion receptor ( <u>Fabriek et al. 2007</u> ).
	Mouse anti rat CD163, clone ED2 was shown to detect approximately 50% of peritoneal macrophages, a subset of splenic macrophages, and most tissue macrophages. However, no staining was observed in monocytes or alveolar macrophages ( <u>Dijkstra et al. 1985</u> , <u>Beelen et al. 1987</u> ). In freshly isolated bone marrow, expression of CD163 was limited to mature macrophages only ( <u>Barbe et al. 1990</u> ).
	Clone ED2 may be used in immunohistology using antigen retrieval, and has also been described reacting with paraffin-embedded material following PLP fixation (Periodatelysine-paraformaldehyde), see Whiteland et al.
Flow Cytometry	Use 10ul of the suggested working dilution to label 1x10 <sup>6</sup> cells in 100ul
Histology Positive Control Tissue	Liver
References	<ol> <li>Barbe, E. <i>et al.</i> (1990) Characterization and expression of the antigen present on resident rat macrophages recognized by monoclonal antibody ED2. Immunobiol. 182: 88-99.</li> <li>Dijkstra, C.D. &amp; Damoiseaux, J.G. (1993) Macrophage heterogeneity established by immunocytochemistry. Prog Histochem Cytochem. 27 (2): 1-65.</li> <li>Whiteland, J.L. <i>et al.</i> (1995) Immunohistochemical detection of T-cell subsets and other leukocytes in paraffin-embedded rat and mouse tissues with monoclonal antibodies. J Histochem Cytochem. 43 (3): 313-20.</li> <li>Muller, D.N. <i>et al.</i> (2002) Immunosuppressive treatment protects against angiotensin II-induced renal damage. Am J Pathol. 161: 1679-93.</li> <li>Polfliet, M.M.J. <i>et al.</i> (2002) Identification of the rat mature macrophage antigen ED2 as CD163: Regulation by glucocorticoids and role in the production of proinflammatory</li> </ol>

- mediators. PhD Thesis. Vrije University, Amsterdam.
- 6. Banerjee, S. *et al.* (2003) Development of organised conjunctival leucocyte aggregates after corneal transplantation in rats. Br J Ophthalmol. 87: 1515-22.
- 7. Moghaddami, M. *et al.* (2005) MHC class II compartment, endocytosis and phagocytic activity of macrophages and putative dendritic cells isolated from normal tissues rich in synovium. Int Immunol. 17: 1117-30.
- 8. Ghiringhelli, F. *et al.* (2005) Tumor cells convert immature myeloid dendritic cells into TGF-beta-secreting cells inducing CD4+CD25+ regulatory T cell proliferation. <u>J Exp Med.</u> 202: 919-29.
- 9. Deng, X. *et al.* (2005) Chronic alcohol consumption accelerates fibrosis in response to cerulein-induced pancreatitis in rats. Am J Pathol. 166 (1): 93-106.
- 10. Fabriek, B.O. *et al.* (2007) The macrophage CD163 surface glycoprotein is an erythroblast adhesion receptor. <u>Blood. 109 (12): 5223-9.</u>
- 11. Wehner, S. *et al.* (2007) Inhibition of macrophage function prevents intestinal inflammation and postoperative ileus in rodents. <u>Gut. 56: 176-85.</u>
- 12. Hamada, T. *et al.* (2007) Oncostatin M gene therapy attenuates liver damage induced by dimethylnitrosamine in rats. <u>Am J Pathol. 171: 872-81.</u>
- 13. Keitel, V. *et al.* (2008) Expression and function of the bile acid receptor TGR5 in Kupffer cells. <u>Biochem Biophys Res Commun.</u> 372: 78-84.
- 14. Starke-Buzetti, W.A. *et al.* (2008) Increased glial-derived neurotrophic factor in the small intestine of rats infected with the tapeworm, *Hymenolepis diminuta*. <u>Int J Exp Pathol.</u> 89: 458-65.
- 15. Jensen, K.T. *et al.* (2008) Early inflammatory response of knee ligaments to prolotherapy in a rat model. J Orthop Res. 26: 816-23.
- 16. Rosas-Ballina, M. *et al.* (2008) Splenic nerve is required for cholinergic antiinflammatory pathway control of TNF in endotoxemia. <u>Proc Natl Acad Sci U S A. 105:</u> 11008-13.
- 17. Santos, M. *et al.* (2009) An unbiased stereological study on subpopulations of rat liver macrophages and on their numerical relation with the hepatocytes and stellate cells. <u>J Anat. 214: 744-51.</u>
- 18. Yamato, M. *et al.* (2009) PET and macro- and microautoradiographic studies combined with immunohistochemistry for monitoring rat intestinal ulceration and healing processes. <u>J Nucl Med. 50: 266-73.</u>
- 19. Yu, Y. *et al.* (2010) Brain perivascular macrophages and the sympathetic response to inflammation in rats after myocardial infarction. <u>Hypertension</u>. <u>55</u>: 652-9.
- 20. Loureiro, J. *et al.* (2010) BMP-7 blocks mesenchymal conversion of mesothelial cells and prevents peritoneal damage induced by dialysis fluid exposure. <u>Nephrol Dial Transplant. 25: 1098-108.</u>
- 21. Bedi, A. *et al.* (2010) Effect of early and delayed mechanical loading on tendon-to-bone healing after anterior cruciate ligament reconstruction. <u>J Bone Joint Surg Am. 92: 2387-401.</u>
- 22. Harty, M.W. *et al.* (2010) Neutrophil depletion blocks early collagen degradation in repairing cholestatic rat livers. <u>Am J Pathol. 176: 1271-81.</u>
- 23. Fujita, E. *et al.* (2010) Statin attenuates experimental anti-glomerular basement membrane glomerulonephritis together with the augmentation of alternatively activated macrophages. <u>Am J Pathol. 177 (3): 1143-54.</u>
- 24. Zeng, Z. et al. (2010) Heme oxygenase-1 protects donor livers from

- ischemia/reperfusion injury: the role of Kupffer cells. World J Gastroenterol. 16: 1285-92.
- 25. Dessem, D. *et al.* (2010) Eccentric muscle contraction and stretching evoke mechanical hyperalgesia and modulate CGRP and P2X(3) expression in a functionally relevant manner. Pain. 149: 284-95.
- 26. Schwartzkopff, J. *et al.* (2010) NK cell depletion delays corneal allograft rejection in baby rats. Mol Vis. 16: 1928-35.
- 27. Teng, B.T. *et al.* (2011) Protective effect of caspase inhibition on compression-induced muscle damage. <u>J Physiol</u>. 589: 3349-69.
- 28. Kajita, M. *et al.* (2011) iNOS expression in vascular resident macrophages contributes to circulatory dysfunction of splanchnic vascular smooth muscle contractions in portal hypertensive rats. <u>Am J Physiol Heart Circ Physiol. 300: H1021-31.</u>
- 29. Baker, S.C. *et al.* (2011) Cellular integration and vascularisation promoted by a resorbable, particulate-leached, cross-linked poly(ε-caprolactone) scaffold. <u>Macromol Biosci. 11 (5): 618-27.</u>
- 30. Kawakami, A.P. *et al.* (2011) Inflammatory Process Modulation by Homeopathic Arnica montana 6CH: The Role of Individual Variation. <u>Evid Based Complement Alternat Med.</u> 2011: 917541.
- 31. Wojcik, M. *et al.* (2012) Immunodetection of cyclooxygenase-2 (COX-2) is restricted to tissue macrophages in normal rat liver and to recruited mononuclear phagocytes in liver injury and cholangiocarcinoma. <u>Histochem Cell Biol. 137: 217-33.</u>
- 32. Jiao, K. *et al.* (2013) The Identification of CD163 Expressing Phagocytic Chondrocytes in Joint Cartilage and Its Novel Scavenger Role in Cartilage Degradation. <u>PLoS One.</u> 8(1):e53312.
- 33. Duchesne, E. *et al.* (2013) Mast cells can regulate skeletal muscle cell proliferation by multiple mechanisms. Muscle Nerve. 48 (3): 403-14.
- 34. Fujii, Y. *et al.* (2013) Effect of enzymatically modified isoquercitrin on preneoplastic liver cell lesions induced by thioacetamide promotion in a two-stage hepatocarcinogenesis model using rats. <u>Toxicology. 305: 30-40.</u>
- 35. Lobato-Pascual, A. *et al.* (2013) Rat macrophage C-type lectin is an activating receptor expressed by phagocytic cells. <u>PLoS One. 8: e57406.</u>
- 36. Park, E.S. *et al.* (2014) Establishment of a rat model for canine necrotizing meningoencephalitis (NME). <u>Vet Pathol. 51 (6): 1151-64.</u>
- 37. Han, T.T. *et al.* (2015) Adipose-derived stromal cells mediate in vivo adipogenesis, angiogenesis and inflammation in decellularized adipose tissue bioscaffolds. <u>Biomaterials.</u> 72: 125-37.
- 38. Stavenuiter, A.W. *et al.* (2015) Protective Effects of Paricalcitol on Peritoneal Remodeling during Peritoneal Dialysis. <u>Biomed Res Int. 2015</u>: 468574.
- 39. Zakrzewicz, A. *et al.* (2015) Monocytic Tissue Transglutaminase in a Rat Model for Reversible Acute Rejection and Chronic Renal Allograft Injury. <u>Mediators Inflamm. 2015:</u> 429653.
- 40. Fernandez-Bustamante, A. *et al.* (2015) Brief Glutamine Pretreatment Increases Alveolar Macrophage CD163/Heme Oxygenase-1/p38-MAPK Dephosphorylation Pathway and Decreases Capillary Damage but Not Neutrophil Recruitment in IL-1/LPS-Insufflated Rats. <u>PLoS One. 10 (7): e0130764.</u>
- 41. Adamo, H.H. *et al.* (2015) Adaptive (TINT) Changes in the Tumor Bearing Organ Are Related to Prostate Tumor Size and Aggressiveness. PLoS One. 10 (11): e0141601.
- 42. Tentillier N et al. (2016) Anti-Inflammatory Modulation of Microglia via CD163-Targeted

- Glucocorticoids Protects Dopaminergic Neurons in the 6-OHDA Parkinson's Disease Model. J Neurosci. 36 (36): 9375-90.
- 43. Pannell, M. *et al.* (2016) Adoptive transfer of M2 macrophages reduces neuropathic pain via opioid peptides. <u>J Neuroinflammation</u>. 13 (1): 262.
- 44. Almahrog, A.J. *et al.* (2016) *In vivo*. association of immunophenotyped macrophages expressing CD163 with PDGF-β in gingival overgrowth-induced by three different categories of medications. <u>J Oral Biol Craniofac Res. 6 (1): 10-7.</u>
- 45. Ibarra, V. *et al.* (2016) Evaluation of the tissue response to alginate encapsulated islets in an omentum pouch model. <u>J Biomed Mater Res A. 104 (7): 1581-90.</u>
- 46. Wang, M. *et al.* (2017) Characterization of the Micro-Environment of the Testis that Shapes the Phenotype and Function of Testicular Macrophages. <u>J Immunol. 198 (11):</u> 4327-40.
- 47. Hawkins, K.E. *et al.* (2017) Targeting resolution of neuroinflammation after ischemic stroke with a lipoxin A<sub>4</sub> analog: Protective mechanisms and long-term effects on neurological recovery. <u>Brain Behav. 7 (5): e00688.</u>
- 48. Chang, J.C. *et al.* (2019) Early Immune Response to Acute Gastric Fluid Aspiration in a Rat Model of Lung Transplantation. Exp Clin Transplant. 17 (1): 84-92.
- 49. Azam, M. *et al.* (2019) Addition of 2-deoxy-d-ribose to clinically used alginate dressings stimulates angiogenesis and accelerates wound healing in diabetic rats. <u>J</u> Biomater Appl. 34 (4): 463-75.
- 50. Bhandari, S. *et al.* (2020) Transcriptome and proteome profiling reveal complementary scavenger and immune features of rat liver sinusoidal endothelial cells and liver macrophages. <u>BMC Mol Cell Biol. 21 (1): 85.</u>
- 51. Nishida, Y. *et al.* (2020) Intra-Articular Injection of Stromal Cell-Derived Factor 1α Promotes Meniscal Healing via Macrophage and Mesenchymal Stem Cell Accumulation in a Rat Meniscal Defect Model. Int J Mol Sci. 21(15):5454.
- 52. Solár, P. *et al.* (2020) Subarachnoid Hemorrhage Increases Level of Heme Oxygenase-1 and Biliverdin Reductase in the Choroid Plexus. <u>Front Cell Neurosci. 14:</u> 593305.
- 53. Li, R. *et al.* (2020) Bone marrow mesenchymal stem cell-derived exosomal microRNA-124-3p attenuates neurological damage in spinal cord ischemia-reperfusion injury by downregulating Ern1 and promoting M2 macrophage polarization. <u>Arthritis Res Ther. 22</u> (1): 75.
- 54. Bloomer, S.A. *et al.* (2020) Aging results in accumulation of M1 and M2 hepatic macrophages and a differential response to gadolinium chloride. <u>Histochem Cell Biol. 153</u> (1): 37-48.
- 55. Broekaart, D.W.M. *et al.* (2020) Increased matrix metalloproteinases expression in tuberous sclerosis complex: modulation by microRNA 146a and 147b *in vitro*.. Neuropathol Appl Neurobiol. 46 (2): 142-159.
- 56. Kuramochi, M. *et al.* (2021) Involvement of neutrophils in rat livers by low-dose thioacetamide administration. <u>J Vet Med Sci. 83 (3): 390-396.</u>
- 57. Matsumoto, K. *et al.* (2021) Role of transient receptor potential vanilloid subtype 2 in lower oesophageal sphincter in rat acid reflux oesophagitis. <u>J Pharmacol Sci. 146 (3): 125-35.</u>
- 58. Koppe, C. *et al.* (2021) Local Inflammatory Response after Intramuscularly Implantation of Anti-Adhesive Plasma-Fluorocarbon-Polymer Coated Ti6Al4V Discs in Rats. <u>Polymers (Basel)</u>. 13(16):2684.

- 59. Matsuyama, S. *et al.* (2021) Properties of macrophages and lymphocytes appearing in rat renal fibrosis followed by repeated injection of cisplatin. <u>J Vet Med Sci. 83 (9): 1435-42.</u> 60. Kim, S. & Son, Y. (2021) Astrocytes Stimulate Microglial Proliferation and M2 Polarization *In Vitro.* through Crosstalk between Astrocytes and Microglia. <u>Int J Mol Sci. 22</u> (16): 8800.
- 61. Zakerkish, F. *et al.* (2021) Differential effects of the immunosuppressive calcineurin inhibitors cyclosporine-A and tacrolimus on ovulation in a murine model. <u>Hum Reprod</u> Open. 2021 (2): hoab012.
- 62. Anderson, L.E. *et al.* (2021) Injection of Micronized Human Amnion/Chorion Membrane Results in Increased Early Supraspinatus Muscle Regeneration in a Chronic Model of Rotator Cuff Tear. <u>Ann Biomed Eng. 49 (12): 3698-710.</u>
- 63. Hou, Y. *et al.* (2021) Pseudoginsenoside-F11 promotes functional recovery after transient cerebral ischemia by regulating the microglia/macrophage polarization in rats. <u>Int Immunopharmacol. 99: 107896.</u>
- 64. Garcia, G.E. *et al.* (2021) A Novel Treatment for Glomerular Disease: Targeting the Activated Macrophage Folate Receptor with a Trojan Horse Therapy in Rats. <u>Cells. 10(8)</u>: 2113.
- 65. Pervin, M. *et al.* (2022) Possible Cytoprotection of Low Dose Lipopolysaccharide in Rat Thioacetamide-Induced Liver Lesions, Focusing on the Analyses of Hepatic Macrophages and Autophagy. <u>Toxicol Pathol.</u>: 1926233221076758.
- 66. Itoh, M. *et al.* (2022) Time-series biological responses toward decellularized bovine tendon graft and autograft for 52 consecutive weeks after rat anterior cruciate ligament reconstruction. Sci Rep. 12 (1): 6751.
- 67. Elahi, E. *et al.* (2022) Immune Modifying Effect of Drug Free Biodegradable Nanoparticles on Disease Course of Experimental Autoimmune Neuritis. <u>Pharmaceutics</u>. 14 (11): 2410.
- 68. Hoff, U. *et al.* (2022) The mTOR inhibitor Rapamycin protects from premature cellular senescence early after experimental kidney transplantation. <u>PLoS One. 17 (4): e0266319.</u> 69. Wu, C.C. *et al.* (2022) Preventive Intrathecal Injection of Bupivacaine Alleviated Microglia Activation and Neuropathic Pain in a Rat Model of Chronic Constriction Injury. <u>Int J Mol Sci. 23 (13): 7197.</u>
- 70. Köhler, R. *et al.* (2022) Association of systemic antibody response against polyethylene terephthalate with inflammatory serum cytokine profile following implantation of differently coated vascular prostheses in a rat animal model. <u>J Biomed Mater Res A.</u> 110 (1): 52-63.
- 71. Takei, R. *et al.* (2022) Dynamic switch of immunity and antitumor effects of metformin in rat spontaneous esophageal carcinogenesis. <u>Cancer Immunol Immunother. 71 (4):</u> 777-89.
- 72. Yu, M. *et al.* (2022) Inhibition of Bruton's Tyrosine Kinase Alleviates Monocrotaline-Induced Pulmonary Arterial Hypertension by Modulating Macrophage Polarization. <u>Oxid Med Cell Longev.</u> 2022: 6526036.
- 73. Takami, Y. *et al.* (2023) The effect of lipopolysaccharide on liver homeostasis and diseases based on the mutual interaction of macrophages, autophagy, and damage-associated molecular patterns in male F344/DuCrlCrlj rats. <u>Vet Pathol. 60 (4): 461-472.</u>
  74. Anderson, L.E. *et al.* (2024) Bone Marrow Mobilization and Local Stromal Cell-Derived Factor-1α Delivery Enhances Nascent Supraspinatus Muscle Fiber Growth. <u>Tissue Eng Part A. 30 (1-2): 45-60.</u>

75. Xu, Z. *et al.* (2024) Schwann Cells Do Not Promote Myogenic Differentiation in the EPI Loop Model. Tissue Eng Part A. 30 (5-6): 244-56.

76. Bodnar, T.S. *et al.* (2022) Modulatory role of prenatal alcohol exposure and adolescent stress on the response to arthritis challenge in adult female rats. <u>EBioMedicine. 77:</u> 103876.

77. Lourenssen, S.R. & Blennerhassett, M.G. (2020) M2 Macrophages and Phenotypic Modulation of Intestinal Smooth Muscle Cells Characterize Inflammatory Stricture Formation in Rats. Am J Pathol. 190 (9): 1843-58.

78. Ismail, T. *et al.* (2020) Platelet-rich plasma and stromal vascular fraction cells for the engineering of axially vascularized osteogenic grafts. <u>J Tissue Eng Regen Med. 14 (12):</u> 1908-17.

79. Suzuki S. *et al.* (2024) Metabolism and effects of acetoaceto-*o*-toluidine in the urinary bladder of humanized-liver mice <u>Journal of Toxicologic Pathology</u>. 18 Sept [Epub ahead of <u>print]</u>.

#### Storage

This product is shipped at ambient temperature. It is recommended to aliquot and store at -20°C on receipt. When thawed, aliquot the sample as needed. Keep aliquots at 2-8°C for short term use (up to 4 weeks) and store the remaining aliquots at -20°C.

Avoid repeated freezing and thawing as this may denature the antibody. Storage in frost-free freezers is not recommended.

Guarantee	12 months from date of despatch
Health And Safety Information	Material Safety Datasheet documentation #10040 available at: <a href="https://www.bio-rad-antibodies.com/SDS/MCA342R">https://www.bio-rad-antibodies.com/SDS/MCA342R</a> 10040
Regulatory	For research purposes only

#### **Related Products**

#### **Recommended Secondary Antibodies**

Rabbit Anti Mouse IgG (STAR12...)

Goat Anti Mouse IgG IgA IgM (STAR87...)

RPE

Goat Anti Mouse IgG (STAR76...)

RPE

Goat Anti Mouse IgG (STAR70...) FITC

Goat Anti Mouse IgG (H/L) (STAR117...) Alk. Phos., DyLight®488, DyLight®550,

<u>DyLight®650</u>, <u>DyLight®680</u>, <u>DyLight®800</u>,

FITC, HRP

Rabbit Anti Mouse IgG (STAR13...) HRP

Goat Anti Mouse IgG (Fc) (STAR120...) FITC, HRP

Rabbit Anti Mouse IgG (STAR9...) <u>FITC</u>
Goat Anti Mouse IgG (STAR77...) <u>HRP</u>

#### **Recommended Negative Controls**

MOUSE IgG1 NEGATIVE CONTROL (MCA1209)

North & South Tel: +1 800 265 7376 Worldwide Tel: +44 (0)1865 852 700 Europe Tel: +49 (0) 89 8090 95 21 America

Fax: +1 919 878 3751 Fax: +44 (0)1865 852 739 Fax: +49 (0) 89 8090 95 50 Email: antibody\_sales\_de@bio-rad.comd a Email: antibody\_sales\_us@bio-rad.com Email: antibody\_sales\_uk@bio-rad.com

То

batch/lot specific datasheet for this product, please use our online search tool at: bio-rad-antibodies.com/datasheets 'M384042:210513'

#### Printed on 19 Sep 2024

© 2024 Bio-Rad Laboratories Inc | Legal | Imprint