## Datasheet: MCA2389B BATCH NUMBER 165770

Description:	RAT ANTI MOUSE Ly-6C:Biotin
Specificity:	Ly-6C
Other names:	Lymphocyte antigen 6C2
Format:	Biotin
Product Type:	Monoclonal Antibody
Clone:	ER-MP20
lsotype:	lgG2a
Quantity:	0.1 mg

## **Product Details**

Applications	This product has been re derived from testing with communications from the information. For general rad-antibodies.com/proto	in our labo e originato protocol re	oratories, rs. Please	peer-reviewed public refer to references	cations or personal indicated for further
		Yes	No	Not Determined	Suggested Dilution
	Flow Cytometry	-			Neat - 1/50
	Where this product has r necessarily exclude its u a guide only. It is recomr system using appropriate	se in such nended th	procedur at the use	es. Suggested work titrates the produc	ing dilutions are given as
Target Species	Mouse				
Product Form	Purified IgG conjugated	to biotin - I	iquid		
Preparation	Purified IgG prepared by supernatant	affinity ch	romatogr	aphy on Protein G fr	om tissue culture
Buffer Solution	Phosphate buffered salir	ie			
Preservative Stabilisers	0.09% sodium azide (Na	•,			
StabiliserS	1% bovine serum album	IN			
Approx. Protein Concentrations	IgG concentration 0.1 m	g/ml			
Immunogen	Balb/c macrophage prec	ursor cell l	nybrids.		

External Database Links	UniProt: <u>P0CW03</u> <u>Related reagents</u>
RRID	AB_844550
Fusion Partners	Spleen cells from immunized rats were fused with cells of the Y3-Ag1.2.3 myeloma cell line.
Specificity	<b>Rat anti Mouse Ly-6C antibody, clone ER-MP20</b> recognizes murine Ly-6C, a 131 amino acid ~14 kDa differentiation antigen, expressed on macrophage/dendritic cell precursors in mid-stage development (late CFU-M, monoblasts and immature monocytes), granulocytes, and on a wide range of endothelial cells and subpopulations of B- and T-lymphocytes.
	Rat anti Mouse Ly-6C antibody, clone ER-MP20 is able to distinguish multiple mouse blood monocyte subsets: immature Ly-6C <sup>hi</sup> monocytes are recruited to acute peripheral inflammation and develop into Ly-6C <sup>+</sup> exudate macrophages, whereas more mature Ly-6C <sup>-/lo</sup> monocytes are precursors for tissue macrophages and dendritic cells in steady state.
	Rat anti Mouse Ly-6C, clone ER-MP20 can be used in conjunction with clone <u>ER-MP12</u> in two colour flow cytometric analysis, to identify different stages of myeloid progenitor cells in mouse bone marrow ( <u>Leenen <i>et al.</i> 1990</u> ).
	Rat anti Mouse Ly-6C was originally described as recognizing a protein encoded by the LY6C gene. It has subsequently become apparent that the LY6C locus demonstrates polymorphism and the LY6C gene has been re-designated LY6C2. The LY6C1 gene encodes a similar protein with ~95% sequence homology to LY6C2.
Flow Cytometry	Use 10µl of the suggested working dilution to label 10 <sup>6</sup> cells in 100µl. The Fc region of monoclonal antibodies may bind to cells expressing low affinity Fc receptors. This may be reduced by using SeroBlock FcR ( <u>BUF041A/BUF041B</u> ).
References	<ol> <li>Zhang, Y. &amp; Bliska, J.B. (2010) YopJ-promoted cytotoxicity and systemic colonization are associated with high levels of murine interleukin-18, gamma interferon, and neutrophils in a live vaccine model of <i>Yersinia pseudotuberculosis</i> infection. Infect Immun 78: 2329-41.</li> <li>Leenen, P.J. <i>et al.</i> (1990) Murine macrophage precursor characterization. II. Monoclonal antibodies against macrophage precursor antigens. <u>Eur J Immunol. 20 (1): 27-34.</u></li> <li>de Bruijn, M.F. <i>et al.</i> (1998) Bone marrow cellular composition in Listeria monocytogenes infected mice detected using ER-MP12 and ER-MP20 antibodies: a flow cytometric alternative to differential counting. <u>J Immunol Methods. 217 (1-2): 27-39.</u></li> <li>Schatteman, G.C. <i>et al.</i> (2010) Lin- Cells Mediate Tissue Repair by Regulating MCP-1/CCL-2. <u>Am J Pathol. 177: 2002-10.</u></li> <li>Baumeister, T. <i>et al.</i> (2003) Interleukin-3Ralpha+ myeloid dendritic cells and mast cells develop simultaneously from different bone marrow precursors in cultures with interleukin-3. <u>J Invest Dermatol. 121: 280-8.</u></li> </ol>

Devey, L. *et al.* (2009) Tissue-resident macrophages protect the liver from ischemia reperfusion injury via a heme oxygenase-1-dependent mechanism. <u>Mol Ther. 17: 65-72.</u>
 Nikolic, T. *et al.* (2003) Developmental stages of myeloid dendritic cells in mouse bone marrow. <u>Int Immunol. 15: 515-24.</u>

8. Wynn, A.A. *et al.* (2001) Role of granulocyte/macrophage colony-stimulating factor in zymocel-induced hepatic granuloma formation. <u>Am J Pathol. 158 (1): 131-45.</u>

9. Lesokhin, A.M. *et al.* (2012) Monocytic CCR2+ Myeloid-Derived Suppressor Cells Promote Immune Escape by Limiting Activated CD8 T-cell Infiltration into the Tumor Microenvironment. <u>Cancer Res. 72: 876-86.</u>

10. Chan, J. *et al.* (1998) Macrophage lineage cells in inflammation: characterization by colony-stimulating factor-1 (CSF-1) receptor (c-Fms), ER-MP58, and ER-MP20 (Ly-6C) expression. <u>Blood. 92: 1423-31.</u>

11. van Rijt, L.S. *et al.* (2002) Allergen-induced accumulation of airway dendritic cells is supported by an increase in CD31(hi)Ly-6C(neg) bone marrow precursors in a mouse model of asthma. <u>Blood. 100: 3663-71.</u>

12. Arnardottir, H.H.*et al.* (2012) Dietary Fish Oil Decreases the Proportion of Classical Monocytes in Blood in Healthy Mice but Increases Their Proportion upon Induction of Inflammation. <u>J Nutr. 142: 803-8.</u>

13. Henkel, G. *et al.* (1999) Commitment to the monocytic lineage occurs in the absence of the transcription factor PU.1. <u>Blood. 93:2849-58.</u>

14. Bossaller, L. *et al.* (2013) Overexpression of membrane-bound fas ligand (CD95L) exacerbates autoimmune disease and renal pathology in pristane-induced lupus. J Immunol. 191: 2104-14.

15. Garcia, J.A. *et al.* (2013) Regulation of adaptive immunity by the fractalkine receptor during autoimmune inflammation. J Immunol. 191: 1063-72.

16. Stijlemans, B. *et al.* (2015) Murine Liver Myeloid Cell Isolation Protocol BIO-PROTOCOL. 5 (10).

17. Damya, L. *et al.* (2014) Purification of Tumor-Associated Macrophages (TAM) and Tumor-Associated Dendritic Cells (TADC) <u>BIO-PROTOCOL. 4 (22).</u>

18. Morganti, J.M. *et al.* (2016) Age exacerbates the CCR2/5-mediated neuroinflammatory response to traumatic brain injury. <u>J Neuroinflammation. 13 (1): 80.</u>

19. Mooney, J.E. *et al.* (2010) Cellular plasticity of inflammatory myeloid cells in the peritoneal foreign body response. <u>Am J Pathol. 176 (1): 369-80.</u>

20. Iwasaki, Y. *et al.* (2011) *In situ* proliferation and differentiation of macrophages in dental pulp. <u>Cell Tissue Res. 346 (1): 99-109.</u>

21. Movahedi, K. *et al.* (2012) Nanobody-based targeting of the macrophage mannose receptor for effective in vivo imaging of tumor-associated macrophages. <u>Cancer Res. 72</u> (<u>16</u>): <u>4165-77</u>.

22. Ribechini, E. *et al.* (2009) Gr-1 antibody induces STAT signaling, macrophage marker expression and abrogation of myeloid-derived suppressor cell activity in BM cells. <u>Eur J</u> <u>Immunol. 39 (12): 3538-51.</u>

23. Bossaller, L. *et al.* (2016) TLR9 Deficiency Leads to Accelerated Renal Disease and Myeloid Lineage Abnormalities in Pristane-Induced Murine Lupus. J Immunol. 197 (4): 1044-53.

24. Barnes, M.A. *et al.* (2015) Macrophage migration inhibitory factor is required for recruitment of scar-associated macrophages during liver fibrosis. <u>J Leukoc Biol. 97 (1):</u> <u>161-9.</u>

25. Ohnishi, K. *et al.* (2012) Immunohistochemical detection of possible cellular origin of hepatic histiocytic sarcoma in mice. J Clin Exp Hematop. 52 (3): 171-7.

26. Van den Bossche. J. *et al.* (2012) Claudin-1, claudin-2 and claudin-11 genes differentially associate with distinct types of anti-inflammatory macrophages *in vitro* and with parasite- and tumour-elicited macrophages *in vivo*. <u>Scand J Immunol. 75 (6): 588-98.</u>
27. Houthuys, E. *et al.* (2010) A method for the isolation and purification of mouse peripheral blood monocytes. J Immunol Methods. 359 (1-2): 1-10.

28. Greifenberg, V. *et al.* (2009) Myeloid-derived suppressor cell activation by combined LPS and IFN-gamma treatment impairs DC development. <u>Eur J Immunol. 39 (10):</u> 2865-76.

29. Cardona, S.M. *et al.* (2015) Disruption of Fractalkine Signaling Leads to Microglial Activation and Neuronal Damage in the Diabetic Retina. <u>ASN Neuro. 7</u> (5):1759091415608204.

30. Waddell, A. *et al.* (2011) Colonic eosinophilic inflammation in experimental colitis is mediated by Ly6C(high) CCR2(+) inflammatory monocyte/macrophage-derived CCL11. J

Immunol. 186 (10): 5993-6003.

31. Robbie, S.J. *et al.* (2016) Enhanced Ccl2-Ccr2 signaling drives more severe choroidal neovascularization with aging. <u>Neurobiol Aging. 40: 110-9.</u>

32. Cao, Y. *et al.* (2016) IL-1 $\beta$  differently stimulates proliferation and multinucleation of distinct mouse bone marrow osteoclast precursor subsets. <u>J Leukoc Biol. 100 (3): 513-23.</u> 33. Cao, Y. *et al.* (2017) TNF- $\alpha$  has both stimulatory and inhibitory effects on mouse monocyte-derived osteoclastogenesis. <u>J Cell Physiol. 232 (12): 3273-85.</u>

34. Khedoe, P.P.S.J. *et al.* (2017) Acute and chronic effects of treatment with mesenchymal stromal cells on LPS-induced pulmonary inflammation, emphysema and atherosclerosis development. <u>PLoS One. 12 (9): e0183741.</u>

35. Koohy, H. *et al.* (2018) Genome organization and chromatin analysis identify transcriptional downregulation of insulin-like growth factor signaling as a hallmark of aging in developing B cells. <u>Genome Biol. 19 (1): 126.</u>

36. Pluijmert, N.J. *et al.* (2020) Effects on cardiac function, remodeling and inflammation following myocardial ischemia-reperfusion injury or unreperfused myocardial infarction in hypercholesterolemic APOE\*3-Leiden mice. <u>Sci Rep. 10 (1): 16601.</u>

37. Ascone, G. *et al.* (2020) Increase in the Number of Bone Marrow Osteoclast Precursors at Different Skeletal Sites, Particularly in Long Bone and Jaw Marrow in Mice Lacking IL-1RA. Int J Mol Sci. 21 (11): 3774.

38. Pluijmert, N.J. *et al.* (2021) Phosphorylcholine antibodies restrict infarct size and left ventricular remodelling by attenuating the unreperfused post-ischaemic inflammatory response. J Cell Mol Med. 25 (16): 7772-82.

39. Njock, M-K. (2022) Endothelial extracellular vesicles promote tumour growth by tumour-associated macrophage reprogramming <u>J Extracell Vesicles 2022</u> Jun;11(6):e12228.

40. Vainchtein, I.D. *et al.* (2023) Characterizing microglial gene expression in a model of secondary progressive multiple sclerosis. <u>Glia. 71 (3): 588-601.</u>

41. Mielczarek, O. *et al.* (2023) Intra- and interchromosomal contact mapping reveals the lgh locus has extensive conformational heterogeneity and interacts with B-lineage genes. <u>Cell Rep 42 (9):113074.</u>

42. Thiem, K. *et al.* (2019) Deletion of hematopoietic Dectin-2 or CARD9 does not protect against atherosclerotic plaque formation in hyperlipidemic mice. <u>Sci Rep. 9 (1): 4337.</u>

	<ul> <li>43. Heissig, B. <i>et al.</i> (2022) siRNA against CD40 delivered via a fungal recognition receptor ameliorates murine acute graft-versus-host disease. <u>EJHaem. 3 (3): 849-61.</u></li> <li>44. Cardona, S.M. <i>et al.</i> (2018) Role of the Fractalkine Receptor in CNS Autoimmune Inflammation: New Approach Utilizing a Mouse Model Expressing the Human CX3CR1(I249/M280) Variant. <u>Front Cell Neurosci. 12: 365.</u></li> </ul>					
	This product is shipped at ambient temperature. It is recommended to aliquot and store a -20°C on receipt. When thawed, aliquot the sample as needed. Keep aliquots at 2-8°C fo short term use (up to 4 weeks) and store the remaining aliquots at -20°C.					
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