

Datasheet: MCA2261F BATCH NUMBER 1707

| Description: | MOUSE ANTI PIG SLA CLASS I:FITC |
|---------------|---------------------------------|
| Specificity: | SLA CLASS I |
| Format: | FITC |
| Product Type: | Monoclonal Antibody |
| Clone: | JM1E3 |
| Isotype: | lgG1 |
| Quantity: | 0.1 mg |

Product Details

| Applications | This product has been reported to work in the following applications. This information is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for further | | | | | | |
|---|---|----------------------|--------|--------------------------|--------------------|--|--|
| | information. For general protocol recommendations, please visit <u>www.bio-</u> | | | | | | |
| | rad-antibodies.com/protocols. | | | | | | |
| | | Yes | No | Not Determined | Suggested Dilution | | |
| | Flow Cytometry | - | | | Neat | | |
| Where this antibody has not been tested for use in a particular technique this do necessarily exclude its use in such procedures. Suggested working dilutions are a guide only. It is recommended that the user titrates the antibody for use in the system using the appropriate negative/positive controls. | | | | | | | |
| Target Species | Pig | | | | | | |
| Species Cross Reactivity | Reacts with: Human N.B. Antibody reactivity and working conditions may vary between species. Cross reactivity is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for further information. | | | | | | |
| Product Form | Purified IgG conjugated to Fluorescein Isothiocyanate Isomer 1 (FITC) - liquid | | | | | | |
| Max Ex/Em | Fluorophore FITC | Excitation Ma 490 | x (nm) | Emission Max (nm) 525 | | | |
| Preparation | Purified IgG prepared by affinity chromatography on Protein A from tissue culture supernatant | | | | | | |
| Buffer Solution | Phosphate buffered sa | line | | | | | |

| Preservative Stabilisers | 0.09% Sodium Azide 1% Bovine Serum Albumin |
|-----------------------------------|--|
| Approx. Protein Concentrations | IgG concentration 0.1 mg/ml |
| Immunogen | Porcine peripheral blood mononuclear cells. |
| RRID | AB_324826 |
| Fusion Partners | Spleen cells from immunised BALB/c mice were fused with cells of the mouse SP2/0 - Ag14 myeloma cell line. |
| Specificity | Mouse anti Pig SLA Class I antibody, clone JM1E3 recognizes a monomorphic epitope expressed by porcine MHC class I molecules (SLA - 1). SLA - 1 is expressed by all nucleated porcine cells, but not on erythrocytes. This antibody has also been shown to cross-react with human MHC Class I, including HLA-E. (Galiani <i>et al.</i> 2002) The major histocompatibility complex (MHC) is a cluster of genes that are important in the immune response to infections. In pigs, this is referred to as the swine leukocyte antigen (SLA) region. Mouse anti pig SLA class I, clone JM1E3 has been reported to block the interaction of MHC Class I antigens with inhibitory NK cell receptors (Galiani <i>et al.</i> 2002). |
| Flow Cytometry | Use 10 ul of the suggested working dilution to label 10 ⁶ cells in 100ul. |
| References | Galiani, D. <i>et al.</i> (2002) A new monoclonal antibody (JM1E3) specific for porcine SLA Class I antigen recognises HLA Class I antigens and interferes with HLA recognition by human NK inhibitory receptors. In Leucocyte Typing VII. Edited by Mason. D. <i>et al.</i> Oxford University Press pp 437-39. Jeong, H.J. <i>et al.</i> (2010) Comparative measurement of cell-mediated immune responses of swine to the M and N proteins of porcine reproductive and respiratory syndrome virus. <u>Clin Vaccine Immunol. 17: 503-12.</u> Hurtado, C. <i>et al.</i> (2011) The African swine fever virus lectin EP153R modulates the surface membrane expression of MHC class I antigens. <u>Arch Virol. 156: 219-34.</u> Ding, G. <i>et al.</i> (2010) Suppression of T cell proliferation by root apical papilla stem cells in vitro. <u>Cells Tissues Organs. 191: 357-64.</u> Park, J.Y. <i>et al.</i> (2008) Characterization of interaction between porcine reproductive and respiratory syndrome virus and porcine dendritic cells. <u>J Microbiol Biotechnol. 18:</u> <u>1709-16.</u> Van Parys, A. <i>et al.</i> (2012) Salmonella Typhimurium induces SPI-1 and SPI-2 regulated and strain dependent downregulation of MHC II expression on porcine alveolar macrophages. <u>Vet Res. 43: 52.</u> Löndt, B.Z. <i>et al.</i> (2013) Enhanced infectivity of H5N1 highly pathogenic avian influenza (HPAI) virus in pig <i>ex vivo</i> respiratory tract organ cultures following adaptation by <i>in vitro</i> passage. <u>Virus Res. 178(2):383-91.</u> Blázquez, R. <i>et al.</i> (2015) Intrapericardial administration of mesenchymal stem cells in a large animal model: a bio-distribution analysis. <u>PLoS One. 10 (3): e0122377.</u> |

| | 9. Rayat, G.R. <i>et al.</i> (2016) First update of the International Xenotransplantation Association consensus statement on conditions for undertaking clinical trials of porcine islet products in type 1 diabetes - Chapter 3: Porcine islet product manufacturing and release testing criteria. <u>Xenotransplantation. 23 (1): 38-45.</u> 10. Suarez-Pinzon, W. <i>et al.</i> (2015) A Novel Protocol for Culturing Adult Porcine Islets for Transplantation in Type 1 Diabetic Patients <u>Minn Acad Sci J Student Res.3: 1-11.</u> 11. Richmond, O. <i>et al.</i> (2015) PD-L1 expression is increased in monocyte derived dendritic cells in response to porcine circovirus type 2 and porcine reproductive and respiratory syndrome virus infections. <u>Vet Immunol Immunopathol. 168 (1-2): 24-9.</u> 12. Iwase H <i>et al.</i> (2015) Initial <i>in vivo</i> experience of pig artery patch transplantation in baboons using mutant MHC (CIITA-DN) pigs. <u>Transpl Immunol. 32 (2): 99-108.</u> 13. Park, K.M. <i>et al.</i> (2013) Generation of porcine induced pluripotent stem cells and evaluation of their major histocompatibility complex protein expression <i>in vitro</i>. <u>Vet Res Commun. 37 (4): 293-301.</u> 14. Le, T.M. <i>et al.</i> (2017) β2-microglobulin gene duplication in cetartiodactyla remains intact only in pigs and possibly confers selective advantage to the species. <u>PLoS One. 12</u> |
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| | (8): e0182322. 15. Linard, C. <i>et al.</i> (2018) Autologous Bone Marrow Mesenchymal Stem Cells Improve the Quality and Stability of Vascularized Flap Surgery of Irradiated Skin in Pigs. <u>Stem</u> <u>Cells Transl Med. 7 (8): 569-582.</u> |
| Further Reading | 1. Piriou-Guzylack, L. (2008) Membrane markers of the immune cells in swine: an update. <u>Vet Res. 39: 54.</u> |
| Storage | Store at +4°C or at -20°C if preferred. |
| | This product should be stored undiluted. |
| | Storage in frost-free freezers is not recommended. This product is photosensitive and should be protected from light. |
| | Avoid repeated freezing and thawing as this may denature the antibody. Should this product contain a precipitate we recommend microcentrifugation before use. |
| Guarantee | 12 months from date of despatch |
| Health And Safety Information | Material Safety Datasheet documentation #10041 available at: https://www.bio-rad-antibodies.com/SDS/MCA2261F 10041 |
| Regulatory | For research purposes only |
| | |

Related Products

Recommended Negative Controls

MOUSE IgG1 NEGATIVE CONTROL: FITC (MCA928F)

To find a batch/lot specific datasheet for this product, please use our online search tool at: bio-rad-antibodies.com/datasheets

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Printed on 18 Jan 2024

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