

## Datasheet: MCA2171GA

<b>Description:</b>	MOUSE ANTI CHICKEN MHC CLASS II MONOMORPHIC
<b>Specificity:</b>	MHC CLASS II MONOMORPHIC
<b>Format:</b>	Purified
<b>Product Type:</b>	Monoclonal Antibody
<b>Clone:</b>	21-1A6
<b>Isotype:</b>	IgG1
<b>Quantity:</b>	0.1 mg

## Product Details

### Applications

This product has been reported to work in the following applications. This information is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for further information. For general protocol recommendations, please visit [www.bio-rad-antibodies.com/protocols](http://www.bio-rad-antibodies.com/protocols).

	Yes	No	Not Determined	Suggested Dilution
Flow Cytometry	▪			1/50 - 1/100
Immunohistology - Frozen	▪			
Immunohistology - Paraffin			▪	
ELISA			▪	
Immunoprecipitation	▪			
Western Blotting			▪	

Where this product has not been tested for use in a particular technique this does not necessarily exclude its use in such procedures. Suggested working dilutions are given as a guide only. It is recommended that the user titrates the product for use in their own system using appropriate negative/positive controls.

<b>Target Species</b>	Chicken
<b>Product Form</b>	Purified IgG - liquid
<b>Preparation</b>	Purified IgG prepared by affinity chromatography on Protein A from tissue culture supernatant
<b>Buffer Solution</b>	Phosphate buffered saline
<b>Preservative Stabilisers</b>	0.09% sodium azide (NaN <sub>3</sub> )
<b>Carrier Free</b>	Yes

Approx. Protein Concentrations	IgG concentration 1.0 mg/ml
Immunogen	Chicken bursa cells
Fusion Partners	Spleen cells from immunized Balb/c mice were fused with cells of the mouse NS-1 myeloma cell line
Specificity	<p><b>Mouse anti Chicken MHC Class II (monomorphic) antibody, clone 21-1A6</b>, recognizes a monomorphic determinant on the chicken B-L molecule, the chicken class II major histocompatibility complex (MHC).</p> <p>The level of B-L expression is reported to increase during the bursal phase of B cell differentiation (<a href="#">Veromaa et al. 1988</a>).</p>
Flow Cytometry	Use 10µl of the suggested working dilution to label 10 <sup>6</sup> cells in 100µl
References	<ol style="list-style-type: none"> <li>1. Veromaa, T. <i>et al.</i> (1988) Expression of B-L and Bu-1 antigens in chickens bursectomized at 60 h of incubation. <a href="#">Eur J Immunol. 18 (2): 225-30.</a></li> <li>2. Vainio, O. <i>et al.</i> (1988) Antigen-presenting cell-T cell interaction in the chicken is MHC class II antigen restricted. <a href="#">J Immunol. 140 (9): 2864-8.</a></li> <li>3. Silva, A.B. <i>et al.</i> (2008) Functional analysis of neuropeptides in avian thymocyte development. <a href="#">Dev Comp Immunol. 32 (4): 410-20.</a></li> <li>4. Watrang, E. (2009) Phosphorothioate oligodeoxyribonucleotides induce in vitro proliferation of chicken B-cells. <a href="#">Vet Immunol Immunopathol. 131 (3-4): 218-28.</a></li> <li>5. Petkov, D.I. <i>et al.</i> (2009) Identification and characterization of two distinct bursal B-cell subpopulations following infectious bursal disease virus infection of White Leghorn chickens. <a href="#">Avian Dis. 53 (3): 347-55.</a></li> <li>6. Pavlova, S.P. <i>et al.</i> (2010) <i>In vitro</i> and <i>in vivo</i> characterization of glycoprotein C-deleted infectious laryngotracheitis virus. <a href="#">J Gen Virol. 91 (Pt 4): 847-57.</a></li> <li>7. Kamble, N.M. <i>et al.</i> (2016) Interaction of a live attenuated <i>Salmonella gallinarum</i> vaccine candidate with chicken bone marrow-derived dendritic cells. <a href="#">Avian Pathol. 45 (2): 235-43.</a></li> <li>8. Kamble, N.M. <i>et al.</i> (2016) Activation of chicken bone marrow-derived dendritic cells induced by a <i>Salmonella Enteritidis</i> ghost vaccine candidate. <a href="#">Poult Sci. 95 (10): 2274-80.</a></li> <li>9. Eren, U. <i>et al.</i> (2016) The several elements of intestinal innate immune system at the beginning of the life of broiler chicks. <a href="#">Microsc Res Tech. 79 (7): 604-14.</a></li> <li>10. Jarosz, Ł. <i>et al. et al.</i> (2016) Effects of feed supplementation with glycine chelate and iron sulfate on selected parameters of cell-mediated immune response in broiler chickens. <a href="#">Res Vet Sci. 107: 68-74.</a></li> <li>11. Jarosz, Ł.S. <i>et al.</i> (2018) The effect of feed supplementation with a copper-glycine chelate and copper sulphate on selected humoral and cell-mediated immune parameters, plasma superoxide dismutase activity, ceruloplasmin and cytokine concentration in broiler chickens. <a href="#">J Anim Physiol Anim Nutr (Berl). 102 (1): e326-e336.</a></li> <li>12. Shojadoost, B. <i>et al.</i> (2019) Interactions between lactobacilli and chicken macrophages induce antiviral responses against avian influenza virus. <a href="#">Res Vet Sci. 125: 441-50.</a></li> <li>13. Garrido, D. <i>et al.</i> (2018) The role of type I interferons (IFNs) in the regulation of</li> </ol>

chicken macrophage inflammatory response to bacterial challenge. [Dev Comp Immunol. 86: 156-70.](#)

14. Yildiz, M. *et al.* (2019) Histological and immunohistochemical studies of the proximal caecum and caecal tonsils of quail (*Coturnix coturnix japonica*). [Anat Histol Embryol. 48 \(5\): 476-85.](#)

15. Eren, U. *et al.* (2022) TLR2 and TLR4 molecules and antigen-presenting cell compositions in cecal tonsils of broiler chicks (*Gallus gallus domesticus*.) in the first two weeks of the post-hatch period. [Anat Histol Embryol. 51 \(1\): 125-135.](#)

16. Saint-Martin, V. *et al.* (2024) The gut microbiota and its metabolite butyrate shape metabolism and antiviral immunity along the gut-lung axis in the chicken. [Commun Biol. 7 \(1\): 1185.](#)

17. Saxena, H.M. and Kaur, P. (2020) Insights into the Identity of the Putative Molecular Target of Infectious Bursal Disease Virus on Chicken Bursal Cells. [Acta Scientific Microbiology 3.11 \(2020\): 61-73.](#)

<b>Storage</b>	This product is shipped at ambient temperature. It is recommended to aliquot and store at -20°C on receipt. When thawed, aliquot the sample as needed. Keep aliquots at 2-8°C for short term use (up to 4 weeks) and store the remaining aliquots at -20°C.  Avoid repeated freezing and thawing as this may denature the antibody. Storage in frost-free freezers is not recommended.
<b>Guarantee</b>	12 months from date of despatch
<b>Health And Safety Information</b>	Material Safety Datasheet documentation #10040 available at: <a href="https://www.bio-rad-antibodies.com/SDS/MCA2171GA">https://www.bio-rad-antibodies.com/SDS/MCA2171GA</a> 10040
<b>Regulatory</b>	For research purposes only

## Related Products

### Recommended Secondary Antibodies

Rabbit Anti Mouse IgG (STAR12...)	<a href="#">RPE</a>
Goat Anti Mouse IgG IgA IgM (STAR87...)	<a href="#">HRP</a>
Goat Anti Mouse IgG (STAR76...)	<a href="#">RPE</a>
Goat Anti Mouse IgG (STAR70...)	<a href="#">FITC</a>
Rabbit Anti Mouse IgG (STAR13...)	<a href="#">HRP</a>
Goat Anti Mouse IgG (Fc) (STAR120...)	<a href="#">FITC</a> , <a href="#">HRP</a>
Rabbit Anti Mouse IgG (STAR9...)	<a href="#">FITC</a>
Goat Anti Mouse IgG (STAR77...)	<a href="#">HRP</a>
Goat Anti Mouse IgG (H/L) (STAR117...)	<a href="#">Alk. Phos.</a> , <a href="#">DyLight®488</a> , <a href="#">DyLight®550</a> , <a href="#">DyLight®650</a> , <a href="#">DyLight®680</a> , <a href="#">DyLight®800</a> , <a href="#">FITC</a> , <a href="#">HRP</a>

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'M433949:250116'

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