

Datasheet: MCA1971PE

BATCH NUMBER 165674

Description:	MOUSE ANTI PIG CD16:RPE
Specificity:	CD16
Other names:	FcRIII
Format:	RPE
Product Type:	Monoclonal Antibody
Clone:	G7
Isotype:	IgG1
Quantity:	100 TESTS

Product Details

Applications

This product has been reported to work in the following applications. This information is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for further information. For general protocol recommendations, please visit www.bio-rad-antibodies.com/protocols.

	Yes	No	Not Determined	Suggested Dilution
Flow Cytometry	▪			Neat

Where this product has not been tested for use in a particular technique this does not necessarily exclude its use in such procedures. Suggested working dilutions are given as a guide only. It is recommended that the user titrates the product for use in their own system using appropriate negative/positive controls.

Target Species	Pig		
Product Form	Purified IgG conjugated to R. Phycoerythrin (RPE) - lyophilized		
Reconstitution	Reconstitute with 1.0 ml distilled water		
Max Ex/Em	Fluorophore	Excitation Max (nm)	Emission Max (nm)
	RPE 488nm laser	496	578
Preparation	Purified IgG prepared by affinity chromatography on Protein A from tissue culture supernatant		
Buffer Solution	Phosphate buffered saline		
Preservative	0.09% sodium azide (NaN ₃)		
Stabilisers	1% bovine serum albumin		

5% sucrose

Immunogen Porcine peripheral blood leucocytes

External Database Links

UniProt:

[Q28942](#) [Related reagents](#)

Entrez Gene:

[397684](#) FCGR3B [Related reagents](#)

RRID AB_2262739

Fusion Partners Spleen cells from immunised Balb/c mice were fused with cells of the mouse P3-X63-Ag8.653 myeloma cell line

Specificity **Mouse anti Pig CD16, clone G7** recognizes porcine CD16 also known as Fc-gamma RIII or the low affinity IgG (Fc) receptor III. Clone G7 was clustered as CD16 at the Second International Workshop to Define Swine Cluster of Differentiation (CD) Antigens ([Saalmuller et al. 1998](#)).

Mouse anti pig CD16 immunoprecipitates a protein of ~40 kDa from porcine neutrophils and NK cells ([Wierda et al. 1993](#)). Subsequent cloning and characterization of the G7 molecule indicated that G7 was the porcine homologue of Human CD16 ([Halloran et al. 1994](#)).

Flow Cytometry Use 10µl of the suggested working dilution to label 10⁶ cells in 100µl

References

1. Wierda, W.G. *et al.* (1993) Two distinct porcine natural killer lytic trigger molecules as PNK-E/G7 molecular complex. [Cell Immunol. 146 \(2\): 270-83.](#)
2. Halloran, P.J. *et al.* (1994) Biochemical characterization of the porcine Fc gamma RIII alpha homologue G7. [Cell Immunol. 158 \(2\): 400-13.](#)
3. Sánchez, C. *et al.* (1999) The porcine 2A10 antigen is homologous to human CD163 and related to macrophage differentiation. [J Immunol. 162 \(9\): 5230-7.](#)
4. Terzic, S. *et al.* (2002) Immunophenotyping of leukocyte subsets in peripheral blood and palatine tonsils of prefattening pigs. [Vet Res Commun. 26: 273 - 83.](#)
5. Vincent, I.E. *et al.* (2003) Dendritic cells harbor infectious porcine circovirus type 2 in the absence of apparent cell modulation or replication of the virus. [J Virol. 77: 13288 - 300.](#)
6. Summerfield, A. *et al.* (2003) Porcine peripheral blood dendritic cells and natural interferon-producing cells. [Immunology 110: 440-9.](#)
7. Inman, C.F. *et al.* (2010) Rearing environment affects development of the immune system in neonates. [Clin Exp Immunol. 160 \(3\): 431-9.](#)
8. Inman, C.F. *et al.* (2010) Dendritic cells interact with CD4 T cells in intestinal mucosa. [J Leukoc Biol. 88 \(3\): 571-8.](#)
9. Devriendt, B. *et al.* (2010) Targeting of *Escherichia coli* F4 fimbriae to Fcγ receptors enhances the maturation of porcine dendritic cells. [Vet Immunol Immunopathol. 135 \(3-4\): 188-98.](#)

10. Gimeno, M. *et al.* (2011) Cytokine profiles and phenotype regulation of antigen presenting cells by genotype-I porcine reproductive and respiratory syndrome virus isolates. [Vet Res. 42: 9.](#)
11. Mussá, T. *et al.* (2011) Interaction of porcine conventional dendritic cells with swine influenza virus. [Virology 420: 125-34.](#)
12. Lecours, M.P. *et al.* (2011) Characterization of porcine dendritic cell response to *Streptococcus suis*. [Vet Res. 42: 72.](#)
13. Mair, K.H. *et al.* (2012) NKp46 expression discriminates porcine NK cells with different functional properties. [Eur J Immunol. 42: 1261-71.](#)
14. Hester, S.N. *et al.* (2012) Intestinal and systemic immune development and response to vaccination are unaffected by dietary (1,3/1,6)- β -D-glucan supplementation in neonatal piglets. [Clin Vaccine Immunol. 19 \(9\): 1499-508.](#)
15. Inman, C.F. *et al.* (2012) Neonatal colonisation expands a specific intestinal antigen-presenting cell subset prior to CD4 T-cell expansion, without altering T-cell repertoire. [PLoS One 7: e33707.](#)
16. Mair, K.H. *et al.* (2013) Porcine CD8 α dim⁻/NKp46^{high} NK cells are in a highly activated state. [Vet Res. 44: 13.](#)
17. Masure, D. *et al.* (2013) A Role for Eosinophils in the Intestinal Immunity against Infective *Ascaris suum* Larvae. [PLoS Negl Trop Dis. 7: e2138.](#)
18. Kyrova, K. *et al.* (2014) The response of porcine monocyte derived macrophages and dendritic cells to *Salmonella typhimurium* and lipopolysaccharide. [BMC Vet Res. 10: 244.](#)
19. Waide, E.H. *et al.* (2015) Not All SCID Pigs Are Created Equally: Two Independent Mutations in the Artemis Gene Cause SCID in Pigs. [J Immunol. 195 \(7\): 3171-9.](#)
20. Auray, G. *et al.* (2016) Characterization and Transcriptomic Analysis of Porcine Blood Conventional and Plasmacytoid Dendritic Cells Reveals Striking Species-Specific Differences. [J Immunol. 197 \(12\): 4791-806.](#)
21. Suzuki, S. *et al.* (2016) Generation and characterization of RAG2 knockout pigs as animal model for severe combined immunodeficiency. [Vet Immunol Immunopathol. 178: 37-49.](#)
22. LeLuduec, J.B. *et al.* (2016) Intradermal vaccination with un-adjuvanted sub-unit vaccines triggers skin innate immunity and confers protective respiratory immunity in domestic swine. [Vaccine. 34 \(7\): 914-22.](#)
23. Loss, H. *et al.* (2018) Effects of a pathogenic ETEC strain and a probiotic *Enterococcus faecium* strain on the inflammasome response in porcine dendritic cells. [Vet Immunol Immunopathol. 203: 78-87.](#)
24. Fernández-Caballero, T. *et al.* (2018) Phenotypic and functional characterization of porcine bone marrow monocyte subsets. [Dev Comp Immunol. 81: 95-104.](#)
25. Skovdal, S.M. *et al.* (2019) Inhaled nebulized glatiramer acetate against Gram-negative bacteria is not associated with adverse pulmonary reactions in healthy, young adult female pigs. [PLoS One. 14 \(10\): e0223647.](#)
26. Ferret-Bernard, S. *et al.* (2020) Maternal Supplementation of Food Ingredient (Prebiotic) or Food Contaminant (Mycotoxin) Influences Mucosal Immune System in Piglets. [Nutrients. 12 \(7\): 2115.](#)
27. Boettcher, A.N. *et al.* (2020) CD3 ϵ (+) Cells in Pigs With Severe Combined Immunodeficiency Due to Defects in ARTEMIS. [Front Immunol. 11: 510.](#)
28. Teuben, M.P.J. *et al.* (2022) Standardized porcine unilateral femoral nailing is associated with changes in PMN activation status, rather than aberrant systemic PMN

prevalence. [Eur J Trauma Emerg Surg. 48 \(3\): 1601-11.](#)

29. Teuben, M. *et al.* (2021) Instant intra-operative neutropenia despite the emergence of banded (CD16^{dim}/CD62L^{bright}) neutrophils in peripheral blood - An observational study during extensive trauma-surgery in pigs. [Injury. 52 \(3\): 426-33.](#)

30. Van der Weken, H. *et al.* (2021) Antibody-Mediated Targeting of Antigens to Intestinal Aminopeptidase N Elicits Gut IgA Responses in Pigs. [Front Immunol. 12: 753371.](#)

31. Zhao, H. *et al.* (2022) Development of *RAG2*^{-/-} *IL2Rγ*^{-/-} immune deficient FAH-knockout miniature pig. [Front Immunol. 13: 950194.](#)

32. Štěpánová, H. *et al.* (2022) Characterization of Porcine Monocyte-Derived Macrophages Cultured in Serum-Reduced Medium. [Biology \(Basel\). 11\(10\):1457.](#)

33. Monguió-Tortajada, M. *et al.* (2022) Acellular cardiac scaffolds enriched with MSC-derived extracellular vesicles limit ventricular remodelling and exert local and systemic immunomodulation in a myocardial infarction porcine model. [Theranostics. 12 \(10\): 4656-70.](#)

Further Reading	1. Piriou-Guzylack, L. (2008) Membrane markers of the immune cells in swine: an update. Vet Res. 39: 54. 2. Gerner W <i>et al.</i> (2015) Phenotypic and functional differentiation of porcine αβ T cells: current knowledge and available tools. Mol Immunol. 66 (1): 3-13.
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Storage	Store at +4°C. DO NOT FREEZE. This product should be stored undiluted. This product is photosensitive and should be protected from light. Should this product contain a precipitate we recommend microcentrifugation before use.
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Guarantee	12 months from date of despatch
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Health And Safety Information	Material Safety Datasheet documentation #20487 available at: https://www.bio-rad-antibodies.com/SDS/MCA1971PE 20487
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Regulatory	For research purposes only
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Related Products

Recommended Negative Controls

[MOUSE IgG1 NEGATIVE CONTROL:RPE \(MCA928PE\)](#)

North & South America	Tel: +1 800 265 7376 Fax: +1 919 878 3751 Email: antibody_sales_us@bio-rad.com	Worldwide	Tel: +44 (0)1865 852 700 Fax: +44 (0)1865 852 739 Email: antibody_sales_uk@bio-rad.com	Europe	Tel: +49 (0) 89 8090 95 21 Fax: +49 (0) 89 8090 95 50 Email: antibody_sales_de@bio-rad.com
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To find a batch/lot specific datasheet for this product, please use our online search tool at: [bio-rad-antibodies.com/datasheets](https://www.bio-rad-antibodies.com/datasheets)

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