

Datasheet: MCA1971A647

Description:	MOUSE ANTI PIG CD16:Alexa Fluor® 647		
Specificity:	CD16		
Other names:	FcRIII		
Format:	ALEXA FLUOR® 647		
Product Type:	Monoclonal Antibody		
Clone:	G7		
Isotype:	IgG1		
Quantity:	100 TESTS/1ml		

Product Details

Applications

This product has been reported to work in the following applications. This information is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for further information. For general protocol recommendations, please visit www.bio-rad-antibodies.com/protocols.

	Yes	No	Not Determined	Suggested Dilution
Flow Cytometry				Neat - 1/10

Where this product has not been tested for use in a particular technique this does not necessarily exclude its use in such procedures. Suggested working dilutions are given as a guide only. It is recommended that the user titrates the product for use in their own system using appropriate negative/positive controls.

Target Species	Pig		
Product Form	Purified IgG conjugate	ed to Alexa Fluor® 64	7 - liquid
Max Ex/Em	Fluorophore	Excitation Max (nm)	Emission Max (nm
	Alexa Fluor®647	650	665
Preparation	Purified IgG prepared supernatant	by affinity chromatog	raphy on Protein A
Buffer Solution	Phosphate buffered sa	aline	
Preservative	0.09% sodium azide (I	NaN ₃)	
Stabilisers	1% bovine serum albu	ımin	
Approx. Protein			

Immunogen	lm	m	un	OQ	en
------------------	----	---	----	----	----

Porcine peripheral blood leucocytes

External Database Links

UniProt:

Q28942 Related reagents

Entrez Gene:

397684 FCGR3B Related reagents

Fusion Partners

Spleen cells from immunized Balb/c mice were fused with cells of the mouse P3-X63-Ag8.653 myeloma cell line

Specificity

Mouse anti Pig CD16, clone G7 recognizes porcine CD16 also known as Fc-gamma RIII or the low affinity IgG (Fc) receptor III. Clone G7 was clustered as CD16 at the Second International Workshop to Define Swine Cluster of Differentiation (CD) Antigens (Saalmuller et al. 1998).

Mouse anti pig CD16 immunoprecipitates a protein of ~40 kDa from porcine neutrophils and NK cells (<u>Wierda et al. 1993</u>). Subsequent cloning and characterization of the G7 molecule indicated that G7 was the porcine homologue of Human CD16 (<u>Halloran et al. 1994</u>).

Flow Cytometry

Use 10µl of the suggested working dilution to label 10⁶ cells in 100µl

References

- 1. Wierda, W.G. *et al.* (1993) Two distinct porcine natural killer lytic trigger molecules as PNK-E/G7 molecular complex. <u>Cell Immunol. 146 (2): 270-83.</u>
- 2. Halloran, P.J. *et al.* (1994) Biochemical characterization of the porcine Fc gamma RIII alpha homologue G7. Cell Immunol. 158 (2): 400-13.
- 3. Sánchez, C. *et al.* (1999) The porcine 2A10 antigen is homologous to human CD163 and related to macrophage differentiation. <u>J Immunol</u>. 162 (9): 5230-7.
- 4. Terzic, S. *et al.* (2002) Immunophenotyping of leukocyte subsets in peripheral blood and palatine tonsils of prefattening pigs. Vet Res Commun. 26: 273 83.
- 5. Vincent, I.E. *et al.* (2003) Dendritic cells harbor infectious porcine circovirus type 2 in the absence of apparent cell modulation or replication of the virus. <u>J Virol. 77: 13288 300</u>.
- 6. Summerfield, A. *et al.* (2003) Porcine peripheral blood dendritic cells and natural interferon-producing cells. <u>Immunology 110: 440-9.</u>
- 7. Inman, C.F. *et al.* (2010) Rearing environment affects development of the immune system in neonates. <u>Clin Exp Immunol. 160 (3): 431-9.</u>
- 8. Inman, C.F. *et al.* (2010) Dendritic cells interact with CD4 T cells in intestinal mucosa. <u>J</u> Leukoc Biol. 88 (3): 571-8.
- 9. Devriendt, B. *et al.* (2010) Targeting of *Escherichia coli* F4 fimbriae to Fcgamma receptors enhances the maturation of porcine dendritic cells. <u>Vet Immunol Immunopathol.</u> 135 (3-4): 188-98.
- 10. Gimeno, M. *et al.* (2011) Cytokine profiles and phenotype regulation of antigen presenting cells by genotype-I porcine reproductive and respiratory syndrome virus isolates. Vet Res. 42: 9.
- 11. Mussá, T. et al. (2011) Interaction of porcine conventional dendritic cells with swine

- influenza virus. Virology 420: 125-34.
- 12. Lecours, M.P. *et al.* (2011) Characterization of porcine dendritic cell response to *Streptococcus suis*. <u>Vet Res. 42: 72.</u>
- 13. Mair, K.H. *et al.* (2012) NKp46 expression discriminates porcine NK cells with different functional properties. <u>Eur J Immunol. 42: 1261-71.</u>
- 14. Hester, S.N. *et al.* (2012) Intestinal and systemic immune development and response to vaccination are unaffected by dietary (1,3/1,6)-β-D-glucan supplementation in neonatal piglets. Clin Vaccine Immunol. 19 (9): 1499-508.
- 15. Thierry, A. *et al.* (2012) Identification of invariant natural killer T cells in porcine peripheral blood. <u>Vet Immunol Immunopathol.</u> 149 (3-4): 272-9.
- 16. Inman, C.F. *et al.* (2012) Neonatal colonisation expands a specific intestinal antigen-presenting cell subset prior to CD4 T-cell expansion, without altering T-cell repertoire. PLoS One 7: e33707.
- 17. Masure, D. *et al.* (2013) A Role for Eosinophils in the Intestinal Immunity against Infective *Ascaris suum* Larvae. <u>PLoS Negl Trop Dis. 7: e2138.</u>
- 18. Mair, K.H. *et al.* (2013) Porcine CD8αdim/-NKp46high NK cells are in a highly activated state. <u>Vet Res. 44: 13.</u>
- 19. Kyrova, K. *et al.* (2014) The response of porcine monocyte derived macrophages and dendritic cells to *Salmonella typhimurium* and lipopolysaccharide. <u>BMC Vet Res. 10: 244.</u>
- 20. Waide, E.H. *et al.* (2015) Not All SCID Pigs Are Created Equally: Two Independent Mutations in the Artemis Gene Cause SCID in Pigs. <u>J Immunol</u>. 195 (7): 3171-9.
- 21. Auray, G. *et al.* (2016) Characterization and Transcriptomic Analysis of Porcine Blood Conventional and Plasmacytoid Dendritic Cells Reveals Striking Species-Specific Differences. J Immunol. 197 (12): 4791-806.
- 22. Suzuki, S. *et al.* (2016) Generation and characterization of RAG2 knockout pigs as animal model for severe combined immunodeficiency. <u>Vet Immunol Immunopathol. 178:</u> 37-49.
- 23. LeLuduec, J.B. *et al.* (2016) Intradermal vaccination with un-adjuvanted sub-unit vaccines triggers skin innate immunity and confers protective respiratory immunity in domestic swine. <u>Vaccine</u>. 34 (7): 914-22.
- 24. Waddell, L.A. *et al.* (2018) ADGRE1 (EMR1, F4/80) Is a Rapidly-Evolving Gene Expressed in Mammalian Monocyte-Macrophages. Front Immunol. 9: 2246.
- 25. Fernández-Caballero, T. *et al.* (2018) Phenotypic and functional characterization of porcine bone marrow monocyte subsets. <u>Dev Comp Immunol. 81: 95-104.</u>
- 26. Loss, H. *et al.* (2018) Effects of a pathogenic ETEC strain and a probiotic Enterococcus faecium strain on the inflammasome response in porcine dendritic cells. <u>Vet Immunol Immunopathol. 203: 78-87.</u>
- 27. Skovdal, S.M. *et al.* (2019) Inhaled nebulized glatiramer acetate against Gram-negative bacteria is not associated with adverse pulmonary reactions in healthy, young adult female pigs. <u>PLoS One. 14 (10): e0223647.</u>
- 28. Ferret-Bernard, S. *et al.* (2020) Maternal Supplementation of Food Ingredient (Prebiotic) or Food Contaminant (Mycotoxin) Influences Mucosal Immune System in Piglets. Nutrients. 12 (7): 2115.
- 29. Boettcher, A.N. *et al.* (2020) CD3ε(+) Cells in Pigs With Severe Combined Immunodeficiency Due to Defects in ARTEMIS. Front Immunol. 11: 510.
- 30. Van der Weken, H. *et al.* (2021) Antibody-Mediated Targeting of Antigens to Intestinal Aminopeptidase N Elicits Gut IgA Responses in Pigs. <u>Front Immunol. 12: 753371.</u>

- 31. Teuben, M. *et al.* (2021) Instant intra-operative neutropenia despite the emergence of banded (CD16^{dim}/CD62L^{bright}) neutrophils in peripheral blood An observational study during extensive trauma-surgery in pigs. <u>Injury. 52 (3): 426-33.</u>
- 32. Teuben, M.P.J. *et al.* (2022) Standardized porcine unilateral femoral nailing is associated with changes in PMN activation status, rather than aberrant systemic PMN prevalence. <u>Eur J Trauma Emerg Surg. 48 (3): 1601-11.</u>
- 33. Zhao, H. *et al.* (2022) Development of *RAG2* -/- *IL2Ry* -/Y immune deficient FAH-knockout miniature pig. <u>Front Immunol. 13: 950194.</u>
- 34. Štěpánová, H. *et al.* (2022) Characterization of Porcine Monocyte-Derived Macrophages Cultured in Serum-Reduced Medium. Biology (Basel). 11(10):1457.
- 35. Monguió-Tortajada, M. *et al.* (2022) Acellular cardiac scaffolds enriched with MSC-derived extracellular vesicles limit ventricular remodelling and exert local and systemic immunomodulation in a myocardial infarction porcine model. <u>Theranostics. 12</u> (10): 4656-70.
- 36. Haach, V. *et al.* (2023) A polyvalent virosomal influenza vaccine induces broad cellular and humoral immunity in pigs. <u>Virol J. 20 (1): 181.</u>
- 37. Álvarez, B. *et al.* (2023) Porcine Macrophage Markers and Populations: An Update. Cells. 12 (16): 2103.
- 38. Maciag, S. *et al.* (2022) Effects of freezing storage on the stability of maternal cellular and humoral immune components in porcine colostrum. <u>Vet Immunol Immunopathol. 254:</u> 110520.
- 39. Forner, R. *et al.* (2021) Distribution difference of colostrum-derived B and T cells subsets in gilts and sows. <u>PLoS One. 16 (5): e0249366.</u>
- 40. Muir, A. *et al.* (2024) Single-cell analysis reveals lasting immunological consequences of influenza infection and respiratory immunization in the pig lung. <u>PLoS Pathog. 20 (7):</u> e1011910.

Further Reading

- 1. Piriou-Guzylack, L. (2008) Membrane markers of the immune cells in swine: an update. Vet Res. 39: 54.
- 2. Gerner W *et al.* (2015) Phenotypic and functional differentiation of porcine $\alpha\beta$ T cells: current knowledge and available tools. Mol Immunol. 66 (1): 3-13.

Storage

This product is shipped at ambient temperature. It is recommended to aliquot and store at -20°C on receipt. When thawed, aliquot the sample as needed. Keep aliquots at 2-8°C for short term use (up to 4 weeks) and store the remaining aliquots at -20°C.

Avoid repeated freezing and thawing as this may denature the antibody. Storage in frost-free freezers is not recommended.

Guarantee

12 months from date of despatch

Acknowledgements

This product is provided under an intellectual property licence from Life Technologies Corporation. The transfer of this product is contingent on the buyer using the purchased product solely in research, excluding contract research or any fee for service research, and the buyer must not sell or otherwise transfer this product or its components for (a) diagnostic, therapeutic or prophylactic purposes; (b) testing, analysis or screening services, or information in return for compensation on a per-test basis; (c) manufacturing or quality assurance or quality control, or (d) resale, whether or not resold for use in

research. For information on purchasing a license to this product for purposes other than as described above, contact Life Technologies Corporation, 5791 Van Allen Way, Carlsbad

CA 92008 USA or outlicensing@thermofisher.com

Health And Safety Information

Material Safety Datasheet documentation #10041 available at:

https://www.bio-rad-antibodies.com/SDS/MCA1971A647

10041

Regulatory For research purposes only

Related Products

Recommended Negative Controls

MOUSE IgG1 NEGATIVE CONTROL: Alexa Fluor® 647 (MCA928A647)

 North & South
 Tel: +1 800 265 7376
 Worldwide
 Tel: +44 (0)1865 852 700
 Europe
 Tel: +49 (0) 89 8090 95 21

 America
 Fax: +1 919 878 3751
 Fax: +44 (0)1865 852 739
 Fax: +49 (0) 89 8090 95 50

To find a batch/lot specific datasheet for this product, please use our online search tool at: bio-rad-antibodies.com/datasheets 'M412397:221111'

Printed on 29 Apr 2025

© 2025 Bio-Rad Laboratories Inc | Legal | Imprint