

## Datasheet: MCA1477T

**BATCH NUMBER 168619**

<b>Description:</b>	RAT ANTI HUMAN CD3
<b>Specificity:</b>	CD3
<b>Format:</b>	Purified
<b>Product Type:</b>	Monoclonal Antibody
<b>Clone:</b>	CD3-12
<b>Isotype:</b>	IgG1
<b>Quantity:</b>	25 µg

## Product Details

### Applications

This product has been reported to work in the following applications. This information is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for further information. For general protocol recommendations, please visit [www.bio-rad-antibodies.com/protocols](http://www.bio-rad-antibodies.com/protocols).

	Yes	No	Not Determined	Suggested Dilution
Flow Cytometry (1)	▪			1/50 - 1/100
Immunohistology - Frozen	▪			1/100
Immunohistology - Paraffin (2)	▪			1/100
Immunofluorescence	▪			

Where this product has not been tested for use in a particular technique this does not necessarily exclude its use in such procedures. Suggested working dilutions are given as a guide only. It is recommended that the user titrates the product for use in their own system using appropriate negative/positive controls.

**(1) Membrane permeabilization is required for this application. The use of Leucoperm (Product Code [BUF09](#)) is recommended for this purpose.**

**(2) This product requires antigen retrieval using heat treatment prior to staining of paraffin sections. Tris/EDTA buffer pH 9.0 is recommended for this purpose.**

<b>Target Species</b>	Human
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<b>Species Cross Reactivity</b>	<p>Reacts with: Bovine, Dog, Horse, Rhesus Monkey, Pig, Chicken, Mouse, Duck, Koala, Harbour Porpoise, Alpaca, Cynomolgus monkey, Spotted Hyena, Sea Lion, Cat, Amazon Parrot, Raccoon, Great horned owl (<i>Bubo virginianus</i>), Bullfrog, Xenopus, Rabbit, African green monkey</p> <p>Based on sequence similarity, is expected to react with: Mammals, Birds, Amphibia</p> <p><b>N.B.</b> Antibody reactivity and working conditions may vary between species. Cross reactivity is derived from testing within our laboratories, peer-reviewed publications or</p>
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personal communications from the originators. Please refer to references indicated for further information.

<b>Product Form</b>	Purified IgG - liquid
<b>Preparation</b>	Purified IgG prepared from tissue culture supernatant
<b>Buffer Solution</b>	Phosphate buffered saline
<b>Preservative Stabilisers</b>	0.09% sodium azide (NaN <sub>3</sub> )
<b>Carrier Free</b>	Yes
<b>Approx. Protein Concentrations</b>	IgG concentration 1.0 mg/ml
<b>Immunogen</b>	Synthetic peptide sequence derived from cytoplasmic epitope of CD3 (Glu-Arg-Pro-Pro-Pro-Val-Pro-Asn-Pro-Asp-Tyr-Glu-Pro-Cys) (ERPPPVPNPDYEPC )
<b>External Database Links</b>	<b>UniProt:</b> <a href="#">P07766</a> <a href="#">Related reagents</a>  <b>Entrez Gene:</b> <a href="#">916</a> CD3E <a href="#">Related reagents</a>
<b>Synonyms</b>	T3E
<b>RRID</b>	AB_10845948
<b>Specificity</b>	<p><b>Rat anti Human CD3 antibody, clone CD3-12</b> raised against a peptide representing an invariant cytoplasmic sequence within the CD3<math>\epsilon</math> chain recognizes human CD3<math>\epsilon</math>. CD3 is a multimeric protein complex composed of four distinct polypeptide chains (<math>\epsilon</math>, <math>\gamma</math>, <math>\delta</math>, <math>\zeta</math>) that assemble and function as three pairs of dimers (<math>\epsilon\gamma</math>, <math>\epsilon\delta</math>, <math>\zeta\zeta</math>). The CD3 complex serves as a T cell co-receptor that associates non-covalently with the T cell receptor (TCR) (<a href="#">Malissen 2008</a>; <a href="#">Guy and Vignali 2009</a>; <a href="#">Smith-Garvin et al. 2009</a>). CD3 is a defining feature of cells belonging to the T cell lineage and can therefore be used as T cell marker.</p> <p>As Rat anti Human CD3, clone CD3-12 has been raised against an epitope within the epsilon peptide chain, highly conserved among species clone CD3-12 has a very broad species crossreactivity for the CD3 marker. (<a href="#">Jones et al. 1993</a>; <a href="#">Kothlow et al. 2005</a>).</p>
<b>Histology Positive Control Tissue</b>	Human tonsil
<b>References</b>	<ol style="list-style-type: none"><li>1. Jones, M. <i>et al.</i> (1993) Detection of T and B cells in many animal species using cross-reactive anti-peptide antibodies. <a href="#">J Immunol. 150 (12): 5429-35.</a></li><li>2. Cornet, A. <i>et al.</i> (2001) Enterocolitis induced by autoimmune targeting of enteric glial</li></ol>

- cells: a possible mechanism in Crohn's disease? [Proc Natl Acad Sci U S A. 98: 13306-11.](#)
3. Trebst, C. *et al.* (2003) CC chemokine receptor 8 in the central nervous system is associated with phagocytic macrophages. [Am J Pathol. 162: 427-38.](#)
  4. Kapturczak, M.H. *et al.* (2004) Heme oxygenase-1 modulates early inflammatory responses: evidence from the heme oxygenase-1-deficient mouse. [Am J Pathol. 165 \(3\): 1045-53.](#)
  5. Shulga-Morskaya, S. *et al.* (2004) B cell-activating factor belonging to the TNF family acts through separate receptors to support B cell survival and T cell-independent antibody formation. [J Immunol. 173 \(4\): 2331-41.](#)
  6. Pusterla, N. *et al.* (2006) Multicentric T-cell lymphosarcoma in an alpaca. [Vet J. 171: 181-5.](#)
  7. Merkler, D. *et al.* (2006) "Viral déjà vu" elicits organ-specific immune disease independent of reactivity to self. [J Clin Invest. 116: 1254-63.](#)
  8. Herrmann, I. *et al.* (2006) *Streptococcus pneumoniae* Infection aggravates experimental autoimmune encephalomyelitis via Toll-like receptor 2. [Infect Immun. 74: 4841-8.](#)
  9. Hudson, K.J. and Bouton, A.H. (2006) *Yersinia pseudotuberculosis* adhesins regulate tissue-specific colonization and immune cell localization in a mouse model of systemic infection. [Infect Immun. 74: 6487-90.](#)
  10. Patole, P.S. *et al.* (2006) Expression and regulation of Toll-like receptors in lupus-like immune complex glomerulonephritis of MRL-Fas(lpr) mice. [Nephrol Dial Transplant 21 \(11\): 3062-73.](#)
  11. Beineke, A. *et al.* (2007) Phenotypical characterization of changes in thymus and spleen associated with lymphoid depletion in free-ranging harbor porpoises (*Phocoena phocoena*). [Vet Immunol Immunopathol. 117: 254-65.](#)
  12. Singleton, C.L. *et al.* (2007) Diagnosis and treatment of chronic T-lymphocytic leukemia in a spotted hyena (*Crocuta crocuta*). [J Zoo Wildl Med. 38: 488-91.](#)
  13. Malka, S. *et al.* (2008) Disseminated lymphoma of presumptive T-cell origin in a great horned owl (*Bubo virginianus*). [J Avian Med Surg. 22: 226-33.](#)
  14. Steinberg, J.D. and Keating, J.H. (2008) What is your diagnosis? Cervical mass in a cat. [Vet Clin Pathol. 37: 323-7.](#)
  15. Tzartos, J.S. *et al.* (2008) Interleukin-17 production in central nervous system-infiltrating T cells and glial cells is associated with active disease in multiple sclerosis. [Am J Pathol. 172: 146-55.](#)
  16. Muljono, A. *et al.* (2009) Primary cutaneous lymphoblastic lymphoma in children: series of eight cases with review of the literature. [Pathology. 41 \(3\): 223-8.](#)
  17. Foryst-Ludwig, A. *et al.* (2010) PPARgamma activation attenuates T-lymphocyte-dependent inflammation of adipose tissue and development of insulin resistance in obese mice. [Cardiovasc Diabetol. 9: 64.](#)
  18. Erdmann, N. *et al.* (2010) Evaluation of the soft tissue biocompatibility of MgCa0.8 and surgical steel 316L in vivo: a comparative study in rabbits. [Biomed Eng Online. 9: 63.](#)
  19. Gendronneau, G. *et al.* (2010) Influence of Hoxa5 on p53 tumorigenic outcome in mice. [Am J Pathol. 176: 995-1005.](#)
  20. Kleiter, I. *et al.* (2010) Smad7 in T cells drives T helper 1 responses in multiple sclerosis and experimental autoimmune encephalomyelitis. [Brain. 2010 Apr;133\(Pt 4\):1067-81.](#)
  21. Redon, C.E. *et al.* (2010) Tumors induce complex DNA damage in distant proliferative tissues *in vivo*. [Proc Natl Acad Sci U S A. 107: 17992-7.](#)

22. Colegrove, K.M. *et al.* (2010) Polyomavirus infection in a free-ranging California sea lion (*Zalophus californianus*) with intestinal T-cell lymphoma. [J Vet Diagn Invest. 22: 628-32.](#)
23. Bartlett SL *et al.* (2010) Intestinal lymphoma of granular lymphocytes in a fisher (*Martes pennanti*) and a Eurasian otter (*Lutra lutra*). [J Zoo Wildl Med. 41 \(2\): 309-15.](#)
24. Dewals B.G., *et al.* (2011) Malignant catarrhal fever induced by Alcelaphine herpesvirus 1 is characterized by an expansion of activated CD3+CD8+CD4- T cells expressing a cytotoxic phenotype in both lymphoid and non-lymphoid tissues [Vet Res. 42:95](#)
25. Osofsky, A. *et al.* (2011) T-cell chronic lymphocytic leukemia in a double yellow-headed Amazon parrot (*Amazona ochrocephala oratrix*). [J Avian Med Surg. 25: 286-94.](#)
26. Tosiek, M.J. *et al.* (2011) CD4+CD25+Foxp3+ regulatory T cells are dispensable for controlling CD8+ T cell-mediated lung inflammation. [J Immunol. 186: 6106-18.](#)
27. Osorio, Y. *et al.* (2011) Identification of small molecule lead compounds for visceral leishmaniasis using a novel *ex vivo* splenic explant model system. [PLoS Negl Trop Dis. 5:e962.](#)
28. Wiessner, C. *et al.* (2011) The Second-Generation Active A{beta} Immunotherapy CAD106 Reduces Amyloid Accumulation in APP Transgenic Mice While Minimizing Potential Side Effects. [J Neurosci. 31: 9323-31.](#)
29. Flatz, L. *et al.* (2011) T cell-dependence of Lassa fever pathogenesis. [PLoS Pathog. 6: e1000836.](#)
30. Lau, Q. *et al.* (2012) Expression and *in vitro* upregulation of MHCII in koala lymphocytes. [Vet Immunol Immunopathol. 147: 35-43.](#)
31. Ruf, M.T. *et al.* (2012) Chemotherapy-Associated Changes of Histopathological Features of *Mycobacterium ulcerans* Lesions in a Buruli Ulcer Mouse Model. [Antimicrob Agents Chemother. 56: 687-96.](#)
32. Campuzano, O. *et al.* (2012) Arrhythmogenic right ventricular cardiomyopathy: severe structural alterations are associated with inflammation. [J Clin Pathol. 65 \(12\): 1077-83.](#)
33. Bricker, N.K. *et al.* (2012) Cytochemical and immunocytochemical characterization of blood cells and immunohistochemical analysis of spleen cells from 2 species of frog, *Rana (Aquarana) catesbeiana* and *Xenopus laevis*. [Vet Clin Pathol. 41: 353-61.](#)
34. Roy, M. *et al.* (2012) CXCL1 can be regulated by IL-6 and promotes granulocyte adhesion to brain capillaries during bacterial toxin exposure and encephalomyelitis. [J Neuroinflammation. 9: 18.](#)
35. Giannitti, F. *et al.* (2014) Temporal and geographic clustering of polyomavirus-associated olfactory tumors in 10 free-ranging raccoons (*Procyon lotor*). [Vet Pathol. 51 \(4\): 832-45.](#)
36. Wen, J. *et al.* (2015) TNF-like weak inducer of apoptosis promotes blood brain barrier disruption and increases neuronal cell death in MRL/lpr mice. [J Autoimmun. 60: 40-50.](#)
37. Ito, D. *et al.* (2015) A double blinded, placebo-controlled pilot study to examine reduction of CD34<sup>+</sup>/CD117<sup>+</sup>/CD133<sup>+</sup> lymphoma progenitor cells and duration of remission induced by neoadjuvant valspodar in dogs with large B-cell lymphoma. [F1000Res. 4: 42.](#)
38. Zhang, M.Z. *et al.* (2015) Inhibition of cyclooxygenase-2 in hematopoietic cells results in salt-sensitive hypertension. [J Clin Invest. 125 \(11\): 4281-94.](#)
39. de Winde, C.M. *et al.* (2015) Multispectral imaging reveals the tissue distribution of tetraspanins in human lymphoid organs. [Histochem Cell Biol. 144 \(2\): 133-46.](#)

40. Velu, V. *et al.* (2016) Induction of Th1-Biased T Follicular Helper (Tfh) Cells in Lymphoid Tissues during Chronic Simian Immunodeficiency Virus Infection Defines Functionally Distinct Germinal Center Tfh Cells. [J Immunol. 197 \(5\): 1832-42.](#)
41. DaSilva, A.V.A. *et al.* (2018) Morphophysiological changes in the splenic extracellular matrix of *Leishmania infantum*-naturally infected dogs is associated with alterations in lymphoid niches and the CD4+ T cell frequency in spleens. [PLoS Negl Trop Dis. 12 \(4\): e0006445.](#)
42. Bonnefont-Rebeix, C. *et al.* (2016) Characterization of a novel canine T-cell line established from a spontaneously occurring aggressive T-cell lymphoma with large granular cell morphology. [Immunobiology. 221 \(1\): 12-22.](#)
43. Withers, S.S. *et al.* (2018) Multi-color flow cytometry for evaluating age-related changes in memory lymphocyte subsets in dogs. [Dev Comp Immunol. 87: 64-74.](#)
44. Sommer, A. *et al.* (2016) Infiltrating T lymphocytes reduce myeloid phagocytosis activity in synucleinopathy model. [J Neuroinflammation 13 \(1\): 174.](#)
45. Houser, K.V. *et al.* (2017) Enhanced inflammation in New Zealand white rabbits when MERS-CoV reinfection occurs in the absence of neutralizing antibody. [PLoS Pathog. 13 \(8\): e1006565.](#)
46. Montes-Cobos, E. *et al.* (2017) Targeted delivery of glucocorticoids to macrophages in a mouse model of multiple sclerosis using inorganic-organic hybrid nanoparticles. [J Control Release. 245: 157-169.](#)
47. Sample, S.J. *et al.* (2017) Radiographic and magnetic resonance imaging predicts severity of cruciate ligament fiber damage and synovitis in dogs with cranial cruciate ligament rupture. [PLoS One. 12 \(6\): e0178086.](#)
48. Kallikourdis, M. *et al.* (2017) T cell costimulation blockade blunts pressure overload-induced heart failure. [Nat Commun. 8: 14680.](#)
49. Sparger, E.E. *et al.* (2018) Investigation of immune cell markers in feline oral squamous cell carcinoma. [Vet Immunol Immunopathol. 202: 52-62.](#)
50. Palomo, J. *et al.* (2018) The severity of imiquimod-induced mouse skin inflammation is independent of endogenous IL-38 expression. [PLoS One. 13 \(3\): e0194667.](#)
51. Declue, A.E. *et al.* (2018) Identification of immunologic and clinical characteristics that predict inflammatory response to C. Novyi-NT bacteriolytic immunotherapy. [BMC Vet Res. 14 \(1\): 119.](#)
52. Pellegrini, S. *et al.* (2019) Selective local irradiation improves islet engraftment and survival in intra-bone marrow islet transplantation. [Cytotherapy. 21 \(10\): 1025-32.](#)
53. Basu, A. *et al.* (2019) Association of PD-L1, PD-L2, and Immune Response Markers in Matched Renal Clear Cell Carcinoma Primary and Metastatic Tissue Specimens. [Am J Clin Pathol. 151 \(2\): 217-25.](#)
54. Gasparitsch, M. *et al.* (2019) Tyrphostin AG490 reduces inflammation and fibrosis in neonatal obstructive nephropathy. [PLoS One. 14 \(12\): e0226675.](#)
55. Thiele, L.S.N. *et al.* (2020) Functional relevance of the multi-drug transporter abcg2 on teriflunomide therapy in an animal model of multiple sclerosis. [J Neuroinflammation. 17 \(1\): 9.](#)
56. Portillo, S. *et al.* (2019) A prophylactic  $\alpha$ -Gal-based glycovaccine effectively protects against murine acute Chagas disease. [NPJ Vaccines. 4: 13.](#)
57. Mejido, D.C.P. *et al.* (2019) Evidences of HEV genotype 3 persistence and reactivity in liver parenchyma from experimentally infected cynomolgus monkeys (*Macaca fascicularis*). [PLoS One. 14 \(6\): e0218472.](#)

58. Choi, S.C. *et al.* (2020) Gut microbiota dysbiosis and altered tryptophan catabolism contribute to autoimmunity in lupus-susceptible mice. [Sci Transl Med. 12 \(551\): eaax2220.](#)
59. Khodadoust, M.S. *et al.* (2020) Pembrolizumab in Relapsed and Refractory Mycosis Fungoides and Sézary Syndrome: A Multicenter Phase II Study. [J Clin Oncol. 38 \(1\): 20-8.](#)
60. Ricat, C.M. *et al.* (2020) Immunohistochemical Findings in Idiopathic Inflammatory Bowel Disease in Nine Cats [BioMed Res Int. 2020: 1-6.](#)
61. Datta, M. *et al.* (2020) Microglial Expression of Hdac1 and Hdac2 is Dispensable for Experimental Autoimmune Encephalomyelitis (EAE) Progression [J. 3 \(4\): 358-65.](#)
62. Bagnoud, M. *et al.* (2020) c-Jun N-Terminal Kinase as a Therapeutic Target in Experimental Autoimmune Encephalomyelitis. [Cells. 9\(10\): 2154.](#)
63. Ishida, Y. *et al.* (2020) Prevention of CaCl<sub>2</sub>-induced aortic inflammation and subsequent aneurysm formation by the CCL3-CCR5 axis. [Nat Commun. 11 \(1\): 5994.](#)
64. Nishri, Y. *et al.* (2020) Continuous Immune-Modulatory Effects of Human Olig2+ Precursor Cells Attenuating a Chronic-Active Model of Multiple Sclerosis. [Mol Neurobiol. 57 \(2\): 1021-34.](#)
65. Ricart, C.M. *et al.* (2020) Immunohistochemical Findings in Idiopathic Inflammatory Bowel Disease in Nine Cats [BioMed Research International. 2020: 1-6.](#)
66. Berghoff, S.A. *et al.* (2021) Microglia facilitate repair of demyelinated lesions via post-squalene sterol synthesis. [Nat Neurosci. 24 \(1\): 47-60.](#)
67. Phillips, D. *et al.* (2021) Immune cell topography predicts response to PD-1 blockade in cutaneous T cell lymphoma. [Nat Commun. 12 \(1\): 6726.](#)
68. Sahin, M. *et al.* (2021) The Janus Kinase Inhibitor Ruxolitinib Prevents Terminal Shock in a Mouse Model of Arenavirus Hemorrhagic Fever. [Microorganisms. 9\(3\):564.](#)
69. Jala, V.R. *et al.* (2021) Absence of CCR2 reduces spontaneous intestinal tumorigenesis in the Apc(Min) (+) mouse model. [Int J Cancer. Jan 26 \[Epub ahead of print\].](#)
70. Tigano, M. *et al.* (2021) *In Vivo* Analysis of mtDNA Replication at the Single Molecule Level and with High Resolution. [Methods Mol Biol. 2192: 21-34.](#)
71. Srivastava, S. *et al.* (2021) Immunogenic Chemotherapy Enhances Recruitment of CAR-T Cells to Lung Tumors and Improves Antitumor Efficacy when Combined with Checkpoint Blockade. [Cancer Cell. 39 \(2\): 193-208.e10.](#)
72. Rajendran, R. *et al.* (2021) Oligodendrocyte-Specific Deletion of *FGFR1* Reduces Cerebellar Inflammation and Neurodegeneration in MOG<sub>35-55</sub>-Induced EAE. [Int J Mol Sci. 22 \(17\): 9495.](#)
73. Häusler, D. *et al.* (2021) CNS inflammation after natalizumab therapy for multiple sclerosis: A retrospective histopathological and CSF cohort study. [Brain Pathol. 31 \(6\): e12969.](#)
74. Datta, M. & Staszewski, O. (2021) Hdac1 and Hdac2 are essential for physiological maturation of a Cx3cr1 expressing subset of T-lymphocytes. [BMC Res Notes. 14 \(1\): 135.](#)
75. Sahin, M. *et al.* (2021) The Janus Kinase Inhibitor Ruxolitinib Prevents Terminal Shock in a Mouse Model of Arenavirus Hemorrhagic Fever. [Microorganisms. 9 \(3\): 564.](#)
76. Winkler, A. *et al.* (2021) Blood-brain barrier resealing in neuromyelitis optica occurs independently of astrocyte regeneration. [J Clin Invest. 131 \(5\): e141694.](#)
77. Cohen, M. *et al.* (2021) Meningeal lymphoid structures are activated under acute and chronic spinal cord pathologies. [Life Sci Alliance. 4 \(1\): e202000907.](#)
78. Huot, N. *et al.* (2021) SIV-induced terminally differentiated adaptive NK cells in lymph nodes associated with enhanced MHC-E restricted activity. [Nat Commun. 12 \(1\): 1282.](#)



79. Arad, T. *et al.* (2021) CD200 -dependent and -independent immune-modulatory functions of neural stem cells. [Stem Cell Res. 56: 102559.](#)
80. Bianchi, A. *et al.* (2021) Moderate Exercise Inhibits Age-Related Inflammation, Liver Steatosis, Senescence, and Tumorigenesis. [J Immunol. ji2001022.](#)
81. Watts, D. *et al.* (2021) Transient Depletion of Foxp3(+) Regulatory T Cells Selectively Promotes Aggressive  $\beta$  Cell Autoimmunity in Genetically Susceptible DEREG Mice. [Front Immunol. 12: 720133.](#)
82. Spitzel, M. *et al.* (2022) Dysregulation of Immune Response Mediators and Pain-Related Ion Channels Is Associated with Pain-like Behavior in the GLA KO Mouse Model of Fabry Disease. [Cells. 11 \(11\): 1730.](#)
83. Martínez-Vallespín, B. *et al.* (2022) Evaluation of High Doses of Phytase in a Low-Phosphorus Diet in Comparison to a Phytate-Free Diet on Performance, Apparent Ileal Digestibility of Nutrients, Bone Mineralization, Intestinal Morphology, and Immune Traits in 21-Day-Old Broiler Chickens. [Animals \(Basel\). 12 \(15\): 1955.](#)
84. Monguió-Tortajada, M. *et al.* (2022) Acellular cardiac scaffolds enriched with MSC-derived extracellular vesicles limit ventricular remodelling and exert local and systemic immunomodulation in a myocardial infarction porcine model. [Theranostics. 12 \(10\): 4656-70.](#)
85. Pepple, K.L. *et al.* (2022) Systemic prime exacerbates the ocular immune response to heat-killed *Mycobacterium tuberculosis*. [Exp Eye Res. 223: 109198.](#)
86. Cequier, A. *et al.* (2022) Equine Mesenchymal Stem Cells Influence the Proliferative Response of Lymphocytes: Effect of Inflammation, Differentiation and MHC-Compatibility. [Animals \(Basel\). 12 \(8\): 984.](#)
87. Maximova, O.A. *et al.* (2023) Pathogenesis and outcome of VA1 astrovirus infection in the human brain are defined by disruption of neural functions and imbalanced host immune responses. [PLoS Pathog. 19 \(8\): e1011544.](#)
88. Tian, Z. *et al.* (2023) Preclinical development of 1B7/CD3, a novel anti-TSLPR bispecific antibody that targets CRLF2-rearranged Ph-like B-ALL. [Leukemia. 37 \(10\): 2006-16.](#)
89. Gogulamudi, V.R. *et al.* (2023) Heterozygosity for ADP-ribosylation factor 6 suppresses the burden and severity of atherosclerosis. [PLoS One. 18 \(5\): e0285253.](#)
90. Wyczanska, M. *et al.* (2023) TLR2 mediates renal apoptosis in neonatal mice subjected experimentally to obstructive nephropathy. [PLoS One. 18 \(11\): e0294142.](#)
91. Enz, L.S. *et al.* (2023) An Animal Model for Chronic Meningeal Inflammation and Inflammatory Demyelination of the Cerebral Cortex. [Int J Mol Sci. 24 \(18\):13893.](#)
92. Ishida, Y. *et al.* (2023) Essential Involvement of Neutrophil Elastase in Acute Acetaminophen Hepatotoxicity Using BALB/c Mice. [Int J Mol Sci. 24 \(9\):7845.](#)
93. Migalska, M. *et al.* (2023) Cross-reactivity of T cell-specific antibodies in the bank vole (*Myodes glareolus*). [J Immunol Methods. 520: 113524.](#)
94. Martini, V. *et al.* (2018) A retrospective study of flow cytometric characterization of suspected extranodal lymphomas in dogs. [J Vet Diagn Invest. 30 \(6\): 830-6.](#)
95. DeClue, A.E. *et al.* (2020) Transportation and Routine Veterinary Interventions Alter Immune Function in the Dog. [Top Companion Anim Med. 39: 100408.](#)
96. Rütgen, B.C. *et al.* (2022) Composition of lymphocyte subpopulations in normal and mildly reactive peripheral lymph nodes in cats. [J Feline Med Surg. 24 \(2\): 77-90.](#)
97. Cha, S. *et al.* (2023) Non-B, Non-T Acute Lymphoblastic Leukemia in a Cat [Journal of Veterinary Clinics. 40 \(4\): 298-302.](#)

98. Yang, L. *et al.* (2018) Association of the expression of Th cytokines with peripheral CD4 and CD8 lymphocyte subsets after vaccination with FMD vaccine in Holstein young sires. [Res Vet Sci. 119: 79-84.](#)
99. Sandersfeld, M. *et al.* (2024) Macrophage subpopulations in pediatric patients with lupus nephritis and other inflammatory diseases affecting the kidney. [Arthritis Res Ther. 26 \(1\): 46.](#)
100. Vos, W.G. *et al.* (2024) T cell specific deletion of Casitas B lineage lymphoma-b reduces atherosclerosis, but increases plaque T cell infiltration and systemic T cell activation [Frontiers in Immunology. 15 04 Mar \[Epub ahead of print\].](#)
101. Wyczanska, M. *et al.* (2024) Interleukin-10 enhances recruitment of immune cells in the neonatal mouse model of obstructive nephropathy. [Sci Rep. 14 \(1\): 5495.](#)
102. Rütgen, B.C. *et al.* (2024) Flowcytometric data of intermediate-large cell gastrointestinal lymphoma presenting a gross mass in 32 cats - "let them glow in the flow". [Front Vet Sci. 11: 1378826.](#)
103. Yang, S.J. *et al.* (2022) Pancreatic islet-specific engineered T(regs) exhibit robust antigen-specific and bystander immune suppression in type 1 diabetes models. [Sci Transl Med. 14 \(665\): eabn1716.](#)
104. Wolfesberger, B. *et al.* (2024) Immunophenotype investigation in feline intestinal non-B-cell lymphoma. [J Comp Pathol. 212: 20-26.](#)
105. White, M.G. *et al.* (2021) Short-term treatment with multi-drug regimens combining BRAF/MEK-targeted therapy and immunotherapy results in durable responses in Braf-mutated melanoma. [Oncoimmunology. 10 \(1\): 1992880.](#)
106. Bilkei-Gorzo, O. *et al.* (2022) The E3 ubiquitin ligase RNF115 regulates phagosome maturation and host response to bacterial infection. [EMBO J. 41 \(23\): e108970.](#)
107. Yilmaz, E.N. *et al.* (2023) Influx of T cells into corpus callosum increases axonal injury, but does not change the course of remyelination in toxic demyelination. [Glia. 71 \(4\): 991-1001.](#)
108. Snyder, J.M. *et al.* (2020) Knockout of Cyp26a1 and Cyp26b1 during postnatal life causes reduced lifespan, dermatitis, splenomegaly, and systemic inflammation in mice. [FASEB J. 34 \(12\): 15788-15804.](#)
109. Frost, S.H.L. *et al.* (2024) <sup>211</sup>At-Labeled Anti-CD45 Antibody as a Nonmyeloablative Conditioning for Canine DLA-Haploidentical Stem Cell Transplantation [Journal of Nuclear Medicine. 65 \(9\): 1443-9.](#)

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**Further Reading**

1. Alterio de Goss, M. *et al.* (1998) Control of cytomegalovirus in bone marrow transplantation chimeras lacking the prevailing antigen-presenting molecule in recipient tissues rests primarily on recipient-derived CD8 T cells. [J Virol. 72 \(10\): 7733-44.](#)
2. Burudi, E.M. *et al.* (2002) Regulation of indoleamine 2,3-dioxygenase expression in simian immunodeficiency virus-infected monkey brains. [J Virol. 76 \(23\): 12233-41.](#)
3. Piriou-Guzylack, L. (2008) Membrane markers of the immune cells in swine: an update. [Vet Res. 39: 54.](#)

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**Storage**

This product is shipped at ambient temperature. It is recommended to aliquot and store at -20°C on receipt. When thawed, aliquot the sample as needed. Keep aliquots at 2-8°C for short term use (up to 4 weeks) and store the remaining aliquots at -20°C.

Avoid repeated freezing and thawing as this may denature the antibody. Storage in



frost-free freezers is not recommended.

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<b>Guarantee</b>	12 months from date of despatch
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<b>Health And Safety Information</b>	Material Safety Datasheet documentation #10040 available at: <a href="https://www.bio-rad-antibodies.com/SDS/MCA1477T">https://www.bio-rad-antibodies.com/SDS/MCA1477T</a> 10040
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<b>Regulatory</b>	For research purposes only
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## Related Products

### Recommended Secondary Antibodies

Rabbit Anti Rat IgG (STAR16...)	<a href="#">DyLight®800</a>
Rabbit Anti Rat IgG (STAR17...)	<a href="#">FITC</a>
Goat Anti Rat IgG (STAR69...)	<a href="#">FITC</a>
Goat Anti Rat IgG (STAR73...)	<a href="#">RPE</a>
Rabbit Anti Rat IgG (STAR21...)	<a href="#">HRP</a>
Goat Anti Rat IgG (MOUSE ADSORBED) (STAR71...)	<a href="#">DyLight®550</a> , <a href="#">DyLight®650</a> , <a href="#">DyLight®800</a>
Goat Anti Rat IgG (STAR131...)	<a href="#">Alk. Phos.</a> , <a href="#">Biotin</a>
Goat Anti Rat IgG (STAR72...)	<a href="#">HRP</a>

### Recommended Negative Controls

[RAT IgG1 NEGATIVE CONTROL \(MCA6004GA\)](#)

<b>North &amp; South America</b>	Tel: +1 800 265 7376 Fax: +1 919 878 3751 Email: <a href="mailto:antibody_sales_us@bio-rad.com">antibody_sales_us@bio-rad.com</a>	<b>Worldwide</b>	Tel: +44 (0)1865 852 700 Fax: +44 (0)1865 852 739 Email: <a href="mailto:antibody_sales_uk@bio-rad.com">antibody_sales_uk@bio-rad.com</a>	<b>Europe</b>	Tel: +49 (0) 89 8090 95 21 Fax: +49 (0) 89 8090 95 50 Email: <a href="mailto:antibody_sales_de@bio-rad.com">antibody_sales_de@bio-rad.com</a>
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To find a batch/lot specific datasheet for this product, please use our online search tool at: [bio-rad-antibodies.com/datasheets](https://bio-rad-antibodies.com/datasheets)  
'M409811:221021'

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