

Datasheet: MCA1425F

Description:	MOUSE ANTI BOVINE CD11b:FITC
Specificity:	CD11b
Other names:	INTEGRIN ALPHA M CHAIN, MAC-1
Format:	FITC
Product Type:	Monoclonal Antibody
Clone:	CC126
Isotype:	IgG2b
Quantity:	0.1 mg

Product Details

Applications

This product has been reported to work in the following applications. This information is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for further information. For general protocol recommendations, please visit www.bio-rad-antibodies.com/protocols.

	Yes	No	Not Determined	Suggested Dilution
Flow Cytometry	▪			1/10

Where this antibody has not been tested for use in a particular technique this does not necessarily exclude its use in such procedures. Suggested working dilutions are given as a guide only. It is recommended that the user titrates the antibody for use in their own system using appropriate negative/positive controls.

Target Species

Bovine

Species Cross Reactivity

Reacts with: Goat, Sheep

N.B. Antibody reactivity and working conditions may vary between species. Cross reactivity is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for further information.

Product Form

Purified IgG conjugated to Fluorescein Isothiocyanate Isomer 1 (FITC) - liquid

Max Ex/Em

Fluorophore	Excitation Max (nm)	Emission Max (nm)
FITC	490	525

Preparation

Purified IgG prepared by affinity chromatography on Protein A from tissue culture supernatant

Buffer Solution

Phosphate buffered saline

Preservative Stabilisers	0.09% Sodium Azide 1% Bovine Serum Albumin
Approx. Protein Concentrations	IgG concentration 0.1 mg/ml
RRID	AB_2265131
Fusion Partners	Spleen cells from immunised BALB/c mice were fused with cells of the mouse NS1 myeloma cell line.
Specificity	Mouse anti Bovine CD11b antibody, clone CC126 recognizes the bovine CD11b cell surface antigen.
Flow Cytometry	Use 10ul of the suggested working dilution to label 10 ⁶ cells in 100ul.
References	<ol style="list-style-type: none"> Howard, C.J. & Naessens, J. (1993) Summary of workshop findings for cattle (tables 1 and 2). Vet Immunol Immunopathol. 39 (1-3): 25-47. Naessens, J. <i>et al.</i> (1993) Cross-reactivity of workshop antibodies with cells from domestic and wild ruminants. Vet Immunol Immunopathol. 39 (1-3): 283-90. Akesson, C.P. <i>et al.</i> (2008) Phenotypic characterisation of intestinal dendritic cells in sheep. Dev Comp Immunol. 32: 837-49. Lecchi, C. <i>et al.</i> (2008) Bovine alpha-1 acid glycoprotein can reduce the chemotaxis of bovine monocytes and modulate CD18 expression. Vet Res. 39: 50. Connelley, T. <i>et al.</i> (2011) NKp46 defines ovine cells that have characteristics corresponding to NK cells. Vet Res. 42: 37. Kallapur, S.G. <i>et al.</i> (2011) Pulmonary and systemic inflammatory responses to intra-amniotic IL-1α in fetal sheep. Am J Physiol Lung Cell Mol Physiol. 301 (3): L285-95. Albenzio, M. & Caroprese, M. (2011) Differential leucocyte count for ewe milk with low and high somatic cell count. J Dairy Res. 78 (1): 43-8. Graff, J.C. and Jutila, M.A. (2007) Differential regulation of CD11b on gammadelta T cells and monocytes in response to unripe apple polyphenols. J Leukoc Biol. 82: 603-7. Top, S. <i>et al.</i> (2011) Infection of nonhost species dendritic cells <i>in vitro</i> with an attenuated myxoma virus induces gene expression that predicts its efficacy as a vaccine vector. J Virol. 85: 12982-94. Albenzio, M. <i>et al.</i> (2012) Immune competence of the mammary gland as affected by somatic cell and pathogenic bacteria in ewes with subclinical mastitis. J Dairy Sci. 95: 3877-87. Muñoz-Caro, T. <i>et al.</i> (2015) <i>Eimeria bovis</i>-triggered neutrophil extracellular trap formation is CD11b-, ERK 1/2-, p38 MAP kinase- and SOCE-dependent. Vet Res. 46: 23. Dedieu, L. <i>et al.</i> (2005) <i>Mycoplasma mycoides</i> ssp. <i>mycoides</i> biotype small colony-secreted components induce apoptotic cell death in bovine leucocytes. Scand J Immunol. 62 (6): 528-38. Flameng, W. <i>et al.</i> (2014) Coating with fibronectin and stromal cell-derived factor-1α of decellularized homografts used for right ventricular outflow tract reconstruction eliminates immune response-related degeneration. J Thorac Cardiovasc Surg. 147 (4): 1398-1404.e2.

14. Edwards, J.C. *et al.* (2010) PrP(Sc) is associated with B cells in the blood of scrapie-infected sheep. [Virology. 405 \(1\): 110-9.](#)
15. McClenahan, D.J. *et al.* (2000) Association among filamentous actin content, CD11b expression, and membrane deformability in stimulated and unstimulated bovine neutrophils. [Am J Vet Res. 61 \(4\): 380-6.](#)
16. Foulon, E. & Foucras, G. (2008) Two populations of ovine bone marrow-derived dendritic cells can be generated with recombinant GM-CSF and separated on CD11b expression. [J Immunol Methods. 339 \(1\): 1-10.](#)
17. Lin, Y. *et al.* (2013) Spatial distribution of polychlorinated naphthalenes in the atmosphere across North China based on gridded field observations. [Environ Pollut. 180: 27-33.](#)
18. Lamote, I. *et al.* (2006) Influence of sex steroids on the viability and CD11b, CD18 and CD47 expression of blood neutrophils from dairy cows in the last month of gestation. [Vet Res. 37 \(1\): 61-74.](#)
19. Weber, B. *et al.* (2016) *In vitro* fabrication of autologous living tissue-engineered vascular grafts based on prenatally harvested ovine amniotic fluid-derived stem cells. [J Tissue Eng Regen Med. 10 \(1\): 52-70.](#)
20. Reber, A.J. *et al.* (2008) Transfer of maternal colostrum leukocytes promotes development of the neonatal immune system I. Effects on monocyte lineage cells. [Vet Immunol Immunopathol. 123 \(3-4\): 186-96.](#)
21. Ozawa, T. *et al.* (2012) Effect of intramammary infusion of rbGM-CSF on SCC and expression of polymorphonuclear neutrophil adhesion molecules in subclinical mastitis cows. [Vet Res Commun. 36 \(1\): 21-7.](#)
22. Zoldan, K. *et al.* (2014) Increase of CD25 expression on bovine neutrophils correlates with disease severity in post-partum and early lactating dairy cows. [Dev Comp Immunol. 47 \(2\): 254-63.](#)
23. Fach, S.J. *et al.* (2006) Pulmonary dendritic cells isolated from neonatal and adult ovine lung tissue. [Vet Immunol Immunopathol. 112 \(3-4\): 171-82.](#)
24. Godoy, R.F. *et al.* (2014) Do progenitor cells from different tissue have the same phenotype? [Res Vet Sci. 96 \(3\): 454-9.](#)
25. Fach, S.J. *et al.* (2007) Neonatal ovine pulmonary dendritic cells support bovine respiratory syncytial virus replication with enhanced interleukin (IL)-4 and IL-10 gene transcripts. [Viral Immunol. 20 \(1\): 119-30.](#)
26. Sassa, Y. *et al.* (2010) Bovine macrophage degradation of scrapie and BSE PrPSc. [Vet Immunol Immunopathol. 133 \(1\): 33-9.](#)
27. Summers, C. *et al.* (2005) An influx of macrophages is the predominant local immune response in ovine pulmonary adenocarcinoma. [Vet Immunol Immunopathol. 106 \(3-4\): 285-94.](#)
28. Pilla, R. *et al.* (2013) Differential cell count as an alternative method to diagnose dairy cow mastitis. [J Dairy Sci. 96 \(3\): 1653-60.](#)
29. Conejeros, I. *et al.* (2015) Effect of the synthetic Toll-like receptor ligands LPS, Pam3CSK4, HKLM and FSL-1 in the function of bovine polymorphonuclear neutrophils. [Dev Comp Immunol. 52 \(2\): 215-25.](#)
30. Merriman, K.E. *et al.* (2018) Intramammary 25-hydroxyvitamin D₃ treatment modulates innate immune responses to endotoxin-induced mastitis. [J Dairy Sci. 101 \(8\): 7593-607.](#)
31. Brodzki, P. *et al.* (2019) Selected leukocyte subpopulations in peripheral blood and uterine washings in cows before and after intrauterine administration of cefapirin and

methisoprinol. [Anim Sci J. Nov 06 \[Epub ahead of print\]](#).

32. Poindexter, M.B. *et al.* (2020) Feeding supplemental 25-hydroxyvitamin D₃ increases serum mineral concentrations and alters mammary immunity of lactating dairy cows. [J Dairy Sci. 103 \(1\): 805-22](#).

Storage

This product is shipped at ambient temperature. It is recommended to aliquot and store at -20°C on receipt. When thawed, aliquot the sample as needed. Keep aliquots at 2-8°C for short term use (up to 4 weeks) and store the remaining aliquots at -20°C.

Avoid repeated freezing and thawing as this may denature the antibody. Storage in frost-free freezers is not recommended. This product is photosensitive and should be protected from light.

Guarantee

12 months from date of despatch

Health And Safety Information

Material Safety Datasheet documentation #10041 available at: 10041: <https://www.bio-rad-antibodies.com/uploads/MSDS/10041.pdf>

Regulatory

For research purposes only

Related Products

Recommended Negative Controls

[MOUSE IgG2b NEGATIVE CONTROL:FITC \(MCA691F\)](#)

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To find a batch/lot specific datasheet for this product, please use our online search tool at: [bio-rad-antibodies.com/datasheets](https://www.bio-rad-antibodies.com/datasheets)

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