## Datasheet: MCA1396A

BATCH NUMBER 158897

| Description: | MOUSE ANTI HISTIDINE TAG:ALK.PHOS. |
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| Specificity: | HISTIDINE TAG |
| Format: | Alk. Phos. |
| Product Type: | Monoclonal Antibody |
| Clone: | AD1.1.10 |
| Isotype: | $\operatorname{lgG} 1$ |
| Quantity: | 0.1 mg |

## Product Details

Applications This product has been reported to work in the following applications. This information is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for further information. For general protocol recommendations, please visit www.bio-rad-antibodies.com/protocols.

|  | Yes | No | Not Determined | Suggested Dilution |
| :--- | :---: | :---: | :---: | :---: |
| Flow Cytometry |  | - |  |  |
| Immunohistology - Frozen |  | - |  |  |
| Immunohistology - Paraffin |  |  |  |  |
| ELISA | - |  | $1 / 20-1 / 1000$ |  |
| Immunoprecipitation |  | - |  |  |
| Western Blotting | - |  | $1 / 1000-1 / 5000$ |  |

Where this antibody has not been tested for use in a particular technique this does not necessarily exclude its use in such procedures. It is recommended that the user titrates the antibody for use in their own system using appropriate negative/positive controls.

| Target Species | Synthetic Peptide |
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| Product Form | Purified IgG conjugated to Alkaline Phosphatase - liquid |
| Preparation | Purified IgG prepared by affinity chromatography on Protein G from tissue culture <br> supernatant |
| Buffer Solution | Phosphate buffered saline |
| Preservative <br> Stabilisers | $0.09 \%$ Sodium Azide $\left(\mathrm{NaN}_{3}\right)$ |
| Approx. Protein <br> Concentrations | $\operatorname{IgG}$ concentration $0.5 \mathrm{mg} / \mathrm{ml}$ |


| Immunogen | PAX6 transcription factor linked to histidine tag. |
| :---: | :---: |
| RRID | AB_1605025 |
| Fusion Partners | Spleen cells from immunised Balb/c mice were fused with cells of the mouse NS1 myeloma cell line. |
| Specificity | Mouse anti Histidine tag antibody, clone AD1.1.10 recognizes proteins and peptides containing the motif $\mathrm{H}-\mathrm{H}-\mathrm{H}-\mathrm{H}-\mathrm{H}-\mathrm{H}$ and is therefore of value in detecting proteins containing histidine tags. Clone AD1.1.10 has been used to detect and purify histidinetagged proteins expressed in mammalian (Hoffmann et al. 2007) and Hwang et al. 2008) and non-mammalian (Zheng et al. 2007; Gunnarsen et al. 2010; and de Vooght et al. 2012) cell lines. |
|  | In Western blotting of bacterial extracts the antibody has been shown not to cross-react with any endogenous products, although some cross-reactivity may be seen with extracts of insect or mammalian cells. |
|  | This antibody is routinely tested in Western blotting on histidine tagged recombinant proteins and reacts against all histidine-tagged proteins so far tested. |
| References | 1. Els Conrath, K. et al. (2001) Camel single-domain antibodies as modular building units in bispecific and bivalent antibody constructs. J Biol Chem. 276 (10): 7346-50. <br> 2. Suen, J.L. et al. (2001) Characterization of self-T-cell response and antigenic determinant of U1A protein with bone marrow-derived dendritic cells in NZB $\times$ NZW $F_{1}$ mice. Immunol. 103: 301-309. |
|  | 3. Hoffmann, S.C. et al. (2007) Identification of CLEC12B, an inhibitory receptor on myeloid cells. J Biol Chem. 282 (31): 22370-5. |
|  | 4. Zheng, J. et al. (2007) Serum from mice immunized in the context of Treg inhibition identifies DEK as a neuroblastoma tumor antigen. BMC Immunol. 8: 4. <br> 5. Bahi, A. \& Dreyer, J.L. (2008) Overexpression of plasminogen activators in the nucleus accumbens enhances cocaine-, amphetamine- and morphine-induced reward and behavioral sensitization. Genes Brain Behav. 7 (2): 244-56. |
|  | 6. Wrighton, K.H. et al. (2009) Transforming Growth Factor \{beta\} Can Stimulate Smad1 Phosphorylation Independently of Bone Morphogenic Protein Receptors. J Biol Chem. 284 (15): 9755-63. |
|  | 7. Diefenbacher, M. et al. (2011) The Dsl1 Tethering Complex Actively Participates in Soluble NSF (N-Ethylmaleimide-sensitive Factor) Attachment Protein Receptor (SNARE) Complex Assembly at the Endoplasmic Reticulum in Saccharomyces cerevisiae. JBiol Chem. 286: 25027-38. |
|  | 8. Alvarez, M.M. et al. (2010) Specific recognition of influenza A/H1N1/2009 antibodies in human serum: a simple virus-free ELISA method. PLoS One. 5: e10176. |
|  | 9. Bahi, A. et al. (2008) The role of tissue-type plasminogen activator system in amphetamine-induced conditional place preference extinction and reinstatement. Neuropsychopharmacology. 33: 2726-34. |
|  | 10. Gunnarsen, K.S. et al. (2010) Periplasmic expression of soluble single chain T cell receptors is rescued by the chaperone FkpA. BMC Biotechnol. 10: 8. |

11. Hwang, H.Y. et al. (2008) Highly specific inhibition of C1q globular-head binding to human IgG: a novel approach to control and regulate the classical complement pathway using an engineered single chain antibody variable fragment. Mol Immunol. 45: 2570-80.
12. De Vooght, L. et al. (2012) Expression and extracellular release of a functional anti-trypanosome Nanobody ${ }^{\circledR}$ in Sodalis glossinidius, a bacterial symbiont of the tsetse fly. Microb Cell Fact. 11: 23.
13. Saerens, D. et al. (2004) Single domain antibodies derived from dromedary lymph node and peripheral blood lymphocytes sensing conformational variants of prostatespecific antigen. J Biol Chem. 279 (50): 51965-72.
14. Than, N.G. et al. (2014) Evolutionary origins of the placental expression of chromosome 19 cluster galectins and their complex dysregulation in preeclampsia.

## Placenta. 35: 855-65.

15. Elders RC et al. (2014) Recombinant canine IgE Fc and an IgE Fc-TRAIL fusion protein bind to neoplastic canine mast cells. Vet Immunol Immunopathol. 159 (1-2): 29-40. 16. Chin, S.E. et al. (2015) Isolation of high-affinity, neutralizing anti-idiotype antibodies by phage and ribosome display for application in immunogenicity and pharmacokinetic analyses. J Immunol Methods. 416: 49-58.
16. Peyrassol, X. et al. (2016) Development by Genetic Immunization of Monovalent Antibodies (Nanobodies) Behaving as Antagonists of the Human ChemR23 Receptor. J Immunol. 196 (6): 2893-901.
17. Kim H \& Loparo JJ (2016) Multistep assembly of DNA condensation clusters by SMC. Nat Commun. 7: 10200.
18. Borg M et al. (2014) A novel interaction between Rab7b and actomyosin reveals a dual role in intracellular transport and cell migration. J Cell Sci. 127 (Pt 22): 4927-39. 20. De Meyer, T. et al. (2015) Comparison of VHH-Fc antibody production in Arabidopsis thaliana, Nicotiana benthamiana and Pichia pastoris. Plant Biotechnol J. 13 (7): 938-47. 21. Siddiqui AA et al. (2015) Humoral immune responses to a recombinant Plasmodium vivax tryptophan-rich antigen among Plasmodium vivax-infected patients and its localization in the parasite. Appl Biochem Biotechnol. 175 (4): 2166-77.
19. Warnecke, A. et al. (2017) Nitration of MOG diminishes its encephalitogenicity depending on MHC haplotype. J Neuroimmunol. 303: 1-12.
20. Bertucci, A. et al. (2011) A new coral carbonic anhydrase in Stylophora pistillata. Mar Biotechnol (NY). 13 (5): 992-1002.
21. Liu, C.C. et al. (2016) The Fimbrial Protein is a Virulence Factor and Potential Vaccine Antigen of Avibacterium paragallinarum. Avian Dis. 60 (3): 649-55.
22. Boujon, C.L. et al. (2017) Development and validation of an immunohistochemistry procedure for the detection of a neurotropic bovine astrovirus. J Virol Methods. 239: 26-33.
23. Cartwright, S.P. et al. (2017) Rapid expression and purification of the hepatitis delta virus antigen using the methylotropic yeast Pichia pastoris. BMC Res Notes. 10 (1): 340. 27. Thanongsaksrikul, J. et al. (2018) Identification and production of mouse scFv to specific epitope of enterovirus-71 virion protein-2 (VP2). Arch Virol. 163 (5): 1141-1152. 28. Gunnarsen, K.S. et al. (2018) Soluble T-cell receptor design influences functional yield in an E. coli chaperone-assisted expression system. PLoS One. 13 (4): e0195868. 29. Ascione, A. et al. (2019) Development of a novel human phage display-derived anti-LAG3 scFv antibody targeting CD8 ${ }^{+}$T lymphocyte exhaustion. BMC Biotechnol. 19 (1): 67.
24. Zoccola, D. et al. (2017) Structural and functional analysis of coral Hypoxia Inducible Factor. PLoS One. 12 (11): e0186262.
25. Kimura, K. et al. (2021) Overexpression of human BAG3 ${ }^{\text {P209L }}$ in mice causes restrictive cardiomyopathy. Nat Commun. 12 (1): 3575.
26. Dongdem, J.T. et al. (2021) Modification of small ubiquitin-related modifier 2 (SUMO2) by phosphoubiquitin in HEK293T cells. Proteomics. 21 (15): e2000234.
27. Chuang, H.C. et al. (2021) Effect of cell-permeable grouper Manganese Superoxide Dismutase on environmental stress in fish. Protein Expr Purif. 187: 105951. 34. Cheng, C.M. et al. (2021) Heterologous expression of bacterial CotA-laccase, characterization and its application for biodegradation of malachite green. Bioresour Technol. 340: 125708.
28. De Vooght, L. et al. (2022) Targeting the tsetse-trypanosome interplay using genetically engineered Sodalis glossinidius.. PLoS Pathog. 18 (3): e1010376.
29. Minami, S.A. et al. (2022) Production of novel SARS-CoV-2 Spike truncations in Chinese hamster ovary cells leads to high expression and binding to antibodies Biotechnol J. Jun 3 [Epub ahead of print]
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\begin{array}{lll}\hline \text { Storage } & \begin{array}{l}\text { Store at }+4^{\circ} \mathrm{C} \text {. DO NOT FREEZE. } \\
\text { This product should be stored undiluted. Should this product contain a precipitate we } \\
\text { recommend microcentrifugation before use. }\end{array}
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\hline Guarantee \& 12 months from date of despatch\end{array}\right]\)|  |  |
| :--- | :--- | :--- |
| Acknowledgements | His-tag is a registered trademark of EMD Biosciences. |

To find a batch/lot specific datasheet for this product, please use our online search tool at: bio-rad-antibodies.com/datasheets 'M365245:200529'

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