

Datasheet: MCA1223F

Description:	MOUSE ANTI PIG wCD8 ALPHA:FITC		
Specificity:	CD8 ALPHA		
Other names:	CD8		
Format:	FITC		
Product Type:	Monoclonal Antibody		
Clone:	MIL12		
Isotype:	lgG2a		
Quantity:	0.1 mg		

Product Details

Applications

This product has been reported to work in the following applications. This information is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for further information. For general protocol recommendations, please visit www.bio-rad-antibodies.com/protocols.

	Yes	No	Not Determined	Suggested Dilution
Flow Cytometry				Neat

Where this product has not been tested for use in a particular technique this does not necessarily exclude its use in such procedures. Suggested working dilutions are given as a guide only. It is recommended that the user titrates the product for use in their own system using appropriate negative/positive controls.

Immunogen	Porcine mesenteric lymphocytes.
RRID	AB_323250
Fusion Partners	Spleen cells from immunized BALB/c mice were fused with cells of the P3 - X63 - Ag.653 myeloma cell line.
Specificity	Mouse anti Pig wCD8 alpha antibody, clone MIL12 recognizes an epitope on the alpha chain of porcine wCD8. Clone MIl12 was clustered at the Third International Swine CD Workshop (Haverson et al. 2001). Mouse anti Pig wCD8 alpha antibody, clone MIL12 was determined to bind to the CD8a epitope on the alpha chain based on its staining pattern on T lymphocytes and on its ability to block binding of the previously characterized CD8a antibody clone 76-2-11 to T lymphocytes (Saalmuller et al.2001).
Flow Cytometry	Use 10µl of the suggested working dilution to label 10 ⁶ cells in 100µl.
References	1. Goujon, J.M. <i>et al.</i> (2000) Influence of cold-storage conditions on renal function of autotransplanted large pig kidneys. <u>Kidney Int. 58: 838-50.</u>

- 2. Spreeuwenberg, M.A. *et al.* (2001) Small intestine epithelial barrier function is compromised in pigs with low feed intake at weaning. <u>J Nutr. 131: 1520-7.</u>
- 3. Carter, D.B. *et al.* (2002) Phenotyping of transgenic cloned piglets. <u>Cloning Stem Cells.</u> 4: 131-45.
- 4. Hauet, T. *et al.* (2002) Polyethylene glycol reduces the inflammatory injury due to cold ischemia/reperfusion in autotransplanted pig kidneys. <u>Kidney Int. 62: 654-67.</u>
- 5. Swamy, H.V. *et al.* (2003) Effects of feeding a blend of grains naturally contaminated with Fusarium mycotoxins on growth and immunological measurements of starter pigs, and the efficacy of a polymeric glucomannan mycotoxin adsorbent. <u>J Anim Sci. 81:</u> 2792-803.
- 6. Sarradell, J. *et al.* (2003) A morphologic and immunohistochemical study of the bronchus-associated lymphoid tissue of pigs naturally infected with *Mycoplasma hyopneumoniae*. <u>Vet Pathol. 40: 395-404.</u>
- 7. Ostrowska, E. *et al.* (2004) Effects of dietary conjugated linoleic acid on haematological and humoral responses in the grower pig <u>Austral J Agric Res. 55 (7): 711.</u>
- 8. Piva, A. *et al.* (2005) Activated carbon does not prevent the toxicity of culture material containing fumonisin B1 when fed to weanling piglets. <u>J Anim Sci. 83 (8): 1939-47.</u>
- 9. Clapperton, M. *et al.* (2005) Innate immune traits differ between Meishan and Large White pigs. Vet Immunol Immunopathol. 104: 131-44.
- 10. Shi, K. *et al.* (2008) Changes in peripheral blood leukocyte subpopulations in piglets co-infected experimentally with porcine reproductive and respiratory syndrome virus and porcine circovirus type 2. <u>Vet Microbiol. 129: 367-77.</u>
- 11. Clapperton, M. *et al.* (2008) Pig peripheral blood mononuclear leucocyte subsets are heritable and genetically correlated with performance. <u>Animal. 2: 1575-84.</u>
- 12. Kick, A.R. *et al.* (2011) Evaluation of peripheral lymphocytes after weaning and vaccination for *Mycoplasma hyopneumoniae*. Res Vet Sci. 91 (3): e68-72.
- 13. Monroy-Salazar, H.G. *et al.* (2012) Effects of a live yeast dietary supplement on fecal coliform counts and on peripheral blood CD4+ and CD8+ lymphocyte subpopulations in nursery pigs. <u>J Swine Health Prod. 20: 276-82.</u>
- 14. Lu, X. et al. (2012) Genome-wide association study for T lymphocyte subpopulations

- in swine. BMC Genomics. 13: 488.
- 15. Leifer, I. *et al.* (2012) Characterization of C-strain "Riems" TAV-epitope escape variants obtained through selective antibody pressure in cell culture. <u>Vet Res. 43: 33.</u>
- 16. Tuchscherer, M. *et al.* (2012) Effects of inadequate maternal dietary protein:carbohydrate ratios during pregnancy on offspring immunity in pigs. <u>BMC Vet Res.</u> 8: 232.
- 17. Kick, A.R. *et al.* (2012) Effects of stress associated with weaning on the adaptive immune system in pigs. <u>J Anim Sci. 90</u>: 649-56.
- 18. Tambuyzer, B.R. *et al.* (2012) Osteopontin alters the functional profile of porcine microglia *in vitro*. Cell Biol Int. 36 (12): 1233-8.
- 19. Stenfeldt, C. *et al.* (2014) Morphologic and phenotypic characteristics of myocarditis in two pigs infected by foot-and mouth disease virus strains of serotypes O or A. <u>Acta Vet Scand</u>. 56: 42.
- 20. Alex, P.J. (2014) Grouping Pig-Specific Responses to Mitogen with Similar Responder Animals may Facilitate the Interpretation of Results Obtained in an Out-Bred Animal Model <u>J Vaccines & Vaccination</u>. 05 (05)
- 21. Zeigler, B.M. *et al.* (2015) The development and validation of methods for evaluating the immune system in preweaning piglets. <u>Food Chem Toxicol. 84: 197-207.</u>
- 22. Liermann, W. *et al.* (2017) Effects of two commercial diets and technical feed treatment on stomach lesions and immune system of fattening pigs. <u>J Anim Physiol Anim Nutr</u> (Berl). 101 (5): e414-e426.
- 23. Hemmink, J.D. *et al.* (2016) Distinct immune responses and virus shedding in pigs following aerosol, intra-nasal and contact infection with pandemic swine influenza A virus, A(H1N1)09. <u>Vet Res. 47 (1): 103.</u>
- 24. López, E. *et al.* (2019) Identification of very early inflammatory markers in a porcine myocardial infarction model. <u>BMC Vet Res. 15 (1): 91.</u>
- 25. Hu, Z. *et al.* (2019) Genomic variant in porcine TNFRSF1A gene and its effects on TNF signaling pathway *in vitro*. Gene. 700: 105-9.
- 26. Fogle, J.E. *et al.* (2019) Antibiotic Therapy Does Not Alter the Humoral Response to Vaccination for Porcine Circovirus 2 in Weaned Pigs. Vet Sci. 6 (2): 51.
- 27. Nielsen, O.L. *et al.* (2022) A porcine model of subcutaneous *Staphylococcus aureus* infection: a pilot study. <u>APMIS. 130 (7): 359-70.</u>
- 28. Maciag, S.S. *et al.* (2022) On the influence of the source of porcine colostrum in the development of early immune ontogeny in piglets. <u>Sci Rep. 12 (1): 15630.</u>
- 29. Wu, M.C. *et al.* (2023) A protein-based subunit vaccine with biological adjuvants provides effective protection against *Pasteurella multocida* in pigs. <u>Vet Res. 54 (1): 17.</u>
- 30. Bujňák, L. *et al.* (2023) The Effect of Dietary Humic Substances on Cellular Immunity and Blood Characteristics in Piglets <u>Agriculture</u>. 13 (3): 636.
- 31. Haach, V. *et al.* (2023) A polyvalent virosomal influenza vaccine induces broad cellular and humoral immunity in pigs. <u>Virol J. 20 (1): 181.</u>
- 32. Maciag, S. *et al.* (2022) Effects of freezing storage on the stability of maternal cellular and humoral immune components in porcine colostrum. <u>Vet Immunol Immunopathol. 254:</u> 110520.
- 33. Forner, R. *et al.* (2021) Distribution difference of colostrum-derived B and T cells subsets in gilts and sows. PLoS One. 16 (5): e0249366.
- 34. Novotný, J. *et al.* (2024) The Use of Flow Cytometry in the Analysis of Sows' Colostrum and Milk Folia Veterinaria. 68 (3): 44-51.

	35. Zhao, W. <i>et al.</i> (2025) Impact of Foot-and-Mouth Disease Virus on Memory T and B Cell Populations in Swine <u>Vet Microbiol. 110406. 5 Jan [Preprint]</u>
Further Reading	1. Piriou-Guzylack, L. (2008) Membrane markers of the immune cells in swine: an update. Vet Res. 39: 54.
Storage	This product is shipped at ambient temperature. It is recommended to aliquot and store at -20°C on receipt. When thawed, aliquot the sample as needed. Keep aliquots at 2-8°C for short term use (up to 4 weeks) and store the remaining aliquots at -20°C.
	Avoid repeated freezing and thawing as this may denature the antibody. Storage in frost-free freezers is not recommended. This product is photosensitive and should be protected from light.
Guarantee	12 months from date of despatch
Health And Safety Information	Material Safety Datasheet documentation #10041 available at: https://www.bio-rad-antibodies.com/SDS/MCA1223F 10041
Regulatory	For research purposes only

Related Products

North & South Tel: +1 800 265 7376

Recommended Negative Controls

MOUSE IgG2a NEGATIVE CONTROL:FITC (MCA929F)

America Fax: +1 919 878 3751

Worldwide Fax: +44 (0)1865 852 739

Tel: +44 (0)1865 852 700

Europe

Tel: +49 (0) 89 8090 95 21 Fax: +49 (0) 89 8090 95 50

Email: antibody_sales_us@bio-rad.com

Email: antibody_sales_uk@bio-rad.com

Email: antibody_sales_de@bio-rad.com

To find a batch/lot specific datasheet for this product, please use our online search tool at: bio-rad-antibodies.com/datasheets 'M408578:221013'

Printed on 24 May 2025

© 2025 Bio-Rad Laboratories Inc | Legal | Imprint