

## Datasheet: MCA1218PE

**BATCH NUMBER 165446**

<b>Description:</b>	MOUSE ANTI PIG CD14:RPE
<b>Specificity:</b>	CD14
<b>Format:</b>	RPE
<b>Product Type:</b>	Monoclonal Antibody
<b>Clone:</b>	MIL2
<b>Isotype:</b>	IgG2b
<b>Quantity:</b>	100 TESTS

### Product Details

#### Applications

This product has been reported to work in the following applications. This information is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for further information. For general protocol recommendations, please visit [www.bio-rad-antibodies.com/protocols](http://www.bio-rad-antibodies.com/protocols).

	Yes	No	Not Determined	Suggested Dilution
Flow Cytometry	▪			Neat - 1/2

Where this product has not been tested for use in a particular technique this does not necessarily exclude its use in such procedures. Suggested working dilutions are given as a guide only. It is recommended that the user titrates the product for use in their own system using appropriate negative/positive controls.

<b>Target Species</b>	Pig		
<b>Species Cross Reactivity</b>	Reacts with: Human <b>N.B.</b> Antibody reactivity and working conditions may vary between species. Cross reactivity is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for further information.		
<b>Product Form</b>	Purified IgG conjugated to R. Phycoerythrin (RPE) - lyophilized		
<b>Reconstitution</b>	Reconstitute with 1 ml distilled water		
<b>Max Ex/Em</b>	<b>Fluorophore</b>	<b>Excitation Max (nm)</b>	<b>Emission Max (nm)</b>
	RPE 488nm laser	496	578
	RPE 561nm laser	546	578

**Preparation** Purified IgG prepared by affinity chromatography on Protein A from tissue culture

supernatant

---

**Buffer Solution** Phosphate buffered saline

---

**Preservative** 0.09% Sodium Azide (NaN<sub>3</sub>)  
**Stabilisers** 1% Bovine Serum Albumin  
5% Sucrose

---

**Immunogen** Porcine peripheral blood lymphocytes.

---

**External Database Links** **UniProt:**  
[A2SW51](#) [Related reagents](#)

---

**Fusion Partners** Spleen cells from immunized Balb/c mice were fused with cells from the P2-X63-Ag.653 mouse myeloma.

---

**Specificity** **Mouse anti Pig CD14, clone MIL2** recognizes porcine CD14. Clone MIL2 was clustered as porcine CD14 at the Third International Workshop on Swine Leukocyte Differentiation Antigens ([Haverson et al. 2001](#)). Mouse anti Pig CD14, clone MIL2 immunoprecipitates a protein of ~50 kDa consistent with the expected apparent molecular weight of porcine CD14, and demonstrates the expected CD14 profile by dual labelling and competition assays. Further, pre-incubation of peripheral blood monocytes with MIL2 inhibits the binding of FITC labelled LPS, consistent with masking the CD14 LPS binding site ([Thacker et al. 2001](#)).

Mouse anti pig CD14, clone MIL2 demonstrates staining of both monocytes and neutrophils in peripheral blood by flow cytometry with a similar expression pattern to the anti human CD14 clone Tük4, lymphocytes and eosinophils are negative for MIL2 staining ([Zelnickova et al. 2007](#)). Cloning and characterization of porcine CD14 indicates a high degree of both functional and structural conservation when compared to CD14 from other mammalian species, the gene maps to chromosome 2 and is expressed on a wide range of tissues in a manner consistent with expression on myeloid cells. ([Petersen et al. 2007](#), [Sanz et al. 2007](#)).

---

**Flow Cytometry** Use 10ul of the suggested working dilution to label 1x10<sup>6</sup> cells in 100ul

---

**References**

1. Hauet, T. *et al.* (2000) Trimetazidine reduces renal dysfunction by limiting the cold ischemia/reperfusion injury in autotransplanted pig kidneys. [J Am Soc Nephrol. 11: 138-48.](#)
2. Thacker, E. *et al.* (2001) Summary of workshop findings for porcine myelomonocytic markers. [Vet Immunol Immunopathol. 80 \(1-2\): 93-109.](#)
3. Thorgersen, E.B. *et al.* (2010) CD14 inhibition efficiently attenuates early inflammatory and hemostatic responses in *Escherichia coli* sepsis in pigs. [FASEB J. 24: 712-22.](#)
4. Goujon, J.M. *et al.* (2000) Influence of cold-storage conditions on renal function of autotransplanted large pig kidneys. [Kidney Int. 58: 838-50.](#)
5. Li, Y. *et al.* (2014) Identification of apoptotic cells in the thymus of piglets infected with highly pathogenic porcine reproductive and respiratory syndrome virus. [Virus Res. 189:](#)

[29-33.](#)

6. Summerfield, A. *et al.* (2003) Porcine peripheral blood dendritic cells and natural interferon-producing cells. [Immunology. 110: 440-9.](#)
7. Vanderheijden, N. *et al.* (2003) Involvement of sialoadhesin in entry of porcine reproductive and respiratory syndrome virus into porcine alveolar macrophages. [J Virol. 77: 8207-15.](#)
8. Barratt-Due, A. *et al.* (2011) *Ornithodoros moubata* Complement Inhibitor Is an Equally Effective C5 Inhibitor in Pigs and Humans. [J Immunol. 187: 4913-9.](#)
9. Hauet, T. *et al.* (2002) Polyethylene glycol reduces the inflammatory injury due to cold ischemia/reperfusion in autotransplanted pig kidneys. [Kidney Int. 62: 654-67.](#)
10. Kapetanovic, R. *et al.* (2012) Pig bone marrow-derived macrophages resemble human macrophages in their response to bacterial lipopolysaccharide. [J Immunol. 188: 3382-94.](#)
11. Thorgersen, E.B. *et al.* (2009) Inhibition of complement and CD14 attenuates the *Escherichia coli*-induced inflammatory response in porcine whole blood. [Infect Immun. 77: 725-32.](#)
12. Zelnickova, P. *et al.* (2007) Intracellular cytokine detection by flow cytometry in pigs: fixation, permeabilization and cell surface staining. [J Immunol Methods. 327: 18-29.](#)
13. Facci, M.R. *et al.* (2011) Stability of expression of reference genes in porcine peripheral blood mononuclear and dendritic cells. [Vet Immunol Immunopathol. 141: 11-5.](#)
14. Koutná, I. *et al.* (2012) Flow Cytometry Analysis of Intracellular Protein In: [Flow Cytometry - Recent Perspectives, Schmid, I. \(Ed.\), ISBN: 978-953-51.](#)
15. Facci, M.R. *et al.* (2010) A comparison between isolated blood dendritic cells and monocyte-derived dendritic cells in pigs. [Immunology. 129: 396-405.](#)
16. Schierack, P. *et al.* (2009) Effects of *Bacillus cereus* var. *toyoi* on immune parameters of pregnant sows. [Vet Immunol Immunopathol. 127: 26-37.](#)
17. Lundeland, B. *et al.* (2011) Severe gunshot injuries in a porcine model: impact on central markers of innate immunity. [Acta Anaesthesiol Scand. 55: 28-34.](#)
18. Thorgersen, E.B. *et al.* (2008) Cyanobacterial LPS antagonist (CyP)-a novel and efficient inhibitor of *Escherichia coli* LPS-induced cytokine response in the pig. [Mol Immunol. 45: 3553-7.](#)
19. Schierack, P. *et al.* (2007) *Bacillus cereus* var. *toyoi* enhanced systemic immune response in piglets. [Vet Immunol Immunopathol. 118: 1-11.](#)
20. Ondrackova, P. *et al.* (2012) Interaction of porcine neutrophils with different strains of enterotoxigenic *Escherichia coli*. [Vet Microbiol. 160: 108-16.](#)
21. Ondrackova, P. *et al.* (2013) Phenotypic characterisation of the monocyte subpopulations in healthy adult pigs and *Salmonella*-infected piglets by seven-colour flow cytometry. [Res Vet Sci. 94 \(2\): 240-5.](#)
22. Vicenova, M. *et al.* (2014) Evaluation of *in vitro* and *in vivo* anti-inflammatory activity of biologically active phospholipids with anti-neoplastic potential in porcine model. [BMC Complement Altern Med. 14: 339.](#)
23. Alvarez, B. *et al.* (2015) Phenotypic and functional heterogeneity of CD169<sup>+</sup> and CD163<sup>+</sup> macrophages from porcine lymph nodes and spleen. [Dev Comp Immunol. 44: 44-9.](#)
24. Moffat, L. *et al.* (2014) Development and characterisation of monoclonal antibodies reactive with porcine CSF1R (CD115). [Dev Comp Immunol. 47 \(1\): 123-8.](#)
25. Kyrova K *et al.* (2014) The response of porcine monocyte derived macrophages and dendritic cells to *Salmonella typhimurium* and lipopolysaccharide. [BMC Vet Res. 10: 244.](#)

26. Nguyen, D.N. *et al.* (2016) Oral antibiotics increase blood neutrophil maturation and reduce bacteremia and necrotizing enterocolitis in the immediate postnatal period of preterm pigs. [Innate Immun. 22 \(1\): 51-62.](#)
27. Egge, K.H. *et al.* (2015) Organ inflammation in porcine *Escherichia coli* sepsis is markedly attenuated by combined inhibition of C5 and CD14. [Immunobiology. 220 \(8\): 999-1005.](#)
28. Liu J *et al.* (2016) The Role of Porcine Monocyte Derived Dendritic Cells (MoDC) in the Inflammation Storm Caused by *Streptococcus suis* Serotype 2 Infection. [PLoS One. 11 \(3\): e0151256.](#)
29. Singleton, H. *et al.* (2016) Establishing Porcine Monocyte-Derived Macrophage and Dendritic Cell Systems for Studying the Interaction with PRRSV-1. [Front Microbiol. 7: 832.](#)
30. Zemankova, N. *et al.* (2016) Bovine lactoferrin free of lipopolysaccharide can induce a proinflammatory response of macrophages. [BMC Vet Res. 12 \(1\): 251.](#)
31. Auray, G. *et al.* (2016) Characterization and Transcriptomic Analysis of Porcine Blood Conventional and Plasmacytoid Dendritic Cells Reveals Striking Species-Specific Differences. [J Immunol. 197 \(12\): 4791-806.](#)
32. Kavanová L *et al.* (2017) Concurrent infection with porcine reproductive and respiratory syndrome virus and *Haemophilus parasuis* in two types of porcine macrophages: apoptosis, production of ROS and formation of multinucleated giant cells. [Vet Res. 48 \(1\): 28.](#)
33. Bacou, E. *et al.* (2017)  $\beta$ 2-adrenoreceptor stimulation dampens the LPS-induced M1 polarization in pig macrophages. [Dev Comp Immunol. 76: 169-76.](#)
34. Yang, G. *et al.* (2017) Characterizing porcine invariant natural killer T cells: A comparative study with NK cells and T cells. [Dev Comp Immunol. 76: 343-351.](#)
35. Uitterdijk, A. *et al.* (2017) Time course of VCAM-1 expression in reperfused myocardial infarction in swine and its relation to retention of intracoronary administered bone marrow-derived mononuclear cells. [PLoS One. 12 \(6\): e0178779.](#)
36. Sánchez, E.G. *et al.* (2017) Phenotyping and susceptibility of established porcine cells lines to African Swine Fever Virus infection and viral production. [Sci Rep. 7 \(1\): 10369.](#)
37. Fernández-Caballero, T. *et al.* (2018) Phenotypic and functional characterization of porcine bone marrow monocyte subsets. [Dev Comp Immunol. 81: 95-104.](#)
38. Sautter, C.A. *et al.* (2018) Phenotypic and functional modulations of porcine macrophages by interferons and interleukin-4. [Dev Comp Immunol. 84: 181-92.](#)
39. López, E. *et al.* (2019) Identification of very early inflammatory markers in a porcine myocardial infarction model. [BMC Vet Res. 15 \(1\): 91.](#)
40. Forner, R. *et al.* (2021) Distribution difference of colostrum-derived B and T cells subsets in gilts and sows. [PLoS One. 16 \(5\): e0249366.](#)
41. Skovdal, S.M. *et al.* (2019) Inhaled nebulized glatiramer acetate against Gram-negative bacteria is not associated with adverse pulmonary reactions in healthy, young adult female pigs. [PLoS One. 14 \(10\): e0223647.](#)
42. Vreman, S. *et al.* (2018) Neonatal porcine blood derived dendritic cell subsets show activation after TLR2 or TLR9 stimulation. [Dev Comp Immunol. 84: 361-70.](#)
43. Lau, C. *et al.* (2020) NHDL, a recombinant V<sub>L</sub>/V<sub>H</sub> hybrid antibody control for IgG2/4 antibodies. [MAbs. 12 \(1\): 1686319.](#)
44. Nielsen, O.L. *et al.* (2022) A porcine model of subcutaneous *Staphylococcus aureus* infection: a pilot study. [APMIS. 130 \(7\): 359-70.](#)
45. Melgoza-González, A.E. *et al.* (2022) Antigen Targeting of Porcine Skin DEC205+

- Dendritic Cells [Vaccines. 10 \(5\): 684.](#)
46. Štěpánová, H. *et al.* (2022) Characterization of Porcine Monocyte-Derived Macrophages Cultured in Serum-Reduced Medium. [Biology \(Basel\). 11\(10\):1457.](#)
47. Monguió-Tortajada, M. *et al.* (2022) Acellular cardiac scaffolds enriched with MSC-derived extracellular vesicles limit ventricular remodelling and exert local and systemic immunomodulation in a myocardial infarction porcine model. [Theranostics. 12 \(10\): 4656-70.](#)
48. Bettin, L. *et al.* (2023) Co-stimulation by TLR7/8 ligand R848 modulates IFN- $\gamma$  production of porcine  $\gamma\delta$  T cells in a microenvironment-dependent manner. [Dev Comp Immunol. 138: 104543.](#)
49. Haach, V. *et al.* (2023) A polyvalent virosomal influenza vaccine induces broad cellular and humoral immunity in pigs. [Virol J. 20 \(1\): 181.](#)
50. Li, J. *et al.* (2024) Single-cell transcriptomic analysis reveals transcriptional and cell subpopulation differences between human and pig immune cells. [Genes Genomics. 46 \(3\): 303-22.](#)
51. Álvarez, B. *et al.* (2023) Porcine Macrophage Markers and Populations: An Update. [Cells. 12 \(16\): 2103.](#)
52. Lawrence, J. *et al.* (2024) Porcine Monocyte DNA Traps Formed during Infection with Pathogenic *Clostridioides difficile* Strains. [Pathogens. 13 \(3\): 228.](#)
53. Auray, G. *et al.* (2020) High-Resolution Profiling of Innate Immune Responses by Porcine Dendritic Cell Subsets *in vitro* and *in vivo*. [Front Immunol. 11: 1429.](#)
54. Nieto-Pelegrín, E. *et al.* (2020) Porcine CLEC12B is expressed on alveolar macrophages and blood dendritic cells. [Dev Comp Immunol. 111: 103767.](#)
55. Maciag, S. *et al.* (2022) Effects of freezing storage on the stability of maternal cellular and humoral immune components in porcine colostrum. [Vet Immunol Immunopathol. 254: 110520.](#)
56. Jarosova, R. *et al.* (2022) Cytokine expression by CD163+ monocytes in healthy and *Actinobacillus pleuropneumoniae*-infected pigs. [Res Vet Sci. 152: 1-9.](#)
57. Waddell, L.A. *et al.* (2018) ADGRE1 (EMR1, F4/80) Is a Rapidly-Evolving Gene Expressed in Mammalian Monocyte-Macrophages. [Front Immunol. 9: 2246.](#)
58. Boschetto, F. *et al.* (2024) Protocol for extracting and isolating porcine bone-marrow-derived macrophages from ribs. [STAR Protoc. 5 \(2\): 103085.](#)

---

**Further Reading**

1. Piriou-Guzylack, L. (2008) Membrane markers of the immune cells in swine: an update. [Vet Res. 39: 54.](#)
2. Petersen, C.B. *et al.* (2007) Cloning, characterization and mapping of porcine CD14 reveals a high conservation of mammalian CD14 structure, expression and locus organization. [Dev Comp Immunol. 31: 729-37.](#)
3. Sanz, G. *et al.* (2007) Molecular cloning, chromosomal location, and expression analysis of porcine CD14. [Dev Comp Immunol. 31\(7\):738-47.](#)

---

**Storage**

Prior to reconstitution store at +4°C.  
After reconstitution store at +4°C.  
DO NOT FREEZE. This product should be stored undiluted. This product is photosensitive and should be protected from light. Should this product contain a precipitate we recommend microcentrifugation before use.

---

**Guarantee**

12 months from date of despatch

---

**Health And Safety Information**      Material Safety Datasheet documentation #20487 available at:  
<https://www.bio-rad-antibodies.com/SDS/MCA1218PE>

---

**Regulatory**                      For research purposes only

---

## Related Products

### Recommended Negative Controls

[MOUSE IgG2b NEGATIVE CONTROL:RPE \(MCA691PE\)](#)

**Product inquiries:** [www.bio-rad-antibodies.com/technical-support](http://www.bio-rad-antibodies.com/technical-support)

To find a batch/lot specific datasheet for this product, please use our online search tool at: [bio-rad-antibodies.com/datasheets](http://bio-rad-antibodies.com/datasheets)  
'M426570:240206'

**Printed on 13 Mar 2026**

---

© 2026 Bio-Rad Laboratories Inc | [Legal](#) | [Imprint](#)