

Datasheet: MCA1044F

Description:	RAT ANTI DOG MHC CLASS II MONOMORPHIC:FITC		
Specificity:	MHC CLASS II MONOMORPHIC		
Format:	FITC		
Product Type:	Monoclonal Antibody		
Clone:	YKIX334.2		
Isotype:	IgG2a		
Quantity:	100 TESTS		

## **Product Details**

# **Applications**

This product has been reported to work in the following applications. This information is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for further information. For general protocol recommendations, please visit <a href="www.bio-rad-antibodies.com/protocols">www.bio-rad-antibodies.com/protocols</a>.

	Yes	No	Not Determined	Suggested Dilution
Flow Cytometry				Neat

Where this antibody has not been tested for use in a particular technique this does not necessarily exclude its use in such procedures. Suggested working dilutions are given as a guide only. It is recommended that the user titrates the antibody for use in their own system using appropriate negative/positive controls.

arget Species	Dog				
species Cross Reactivity	Does not react with:Hooded Seal				
roduct Form	Purified IgG conjuga	ted to Fluorescein Isoth	niocyanate Isomer 1		
lax Ex/Em	Fluorophore	Excitation Max (nm)	Emission Max (nm		
	FITC	490	525		
Preparation	Purified IgG prepared supernatant	d by affinity chromatog	raphy on Protein G		
uffer Solution	Phosphate buffered	saline			
reservative	0.09% sodium azide	(NaN <sub>3</sub> )			
Stabilisers	1% bovine serum alb	oumin			
Approx. Protein	IgG concentration 0.	1 mg/ml			

#### Concentrations

Immunogen	Con A activated canine peripheral blood cells.	
RRID	AB_322642	
Fusion Partners	Spleen cells from immunized DA rats were fused with cells of the rat Y3/Ag1.2.3 myeloma cell line.	
Specificity	Rat anti Dog MHC Class II Monomorphic antibody, clone YKIX334.2 recognizes a monomorphic epitope on canine MHC Class II and was classified at the First Canine Leucocyte Antigen Workshop (Cobbold et al. 1994). The major histocompatibility complex (MHC) is a cluster of genes that are important in the immune response to infections. In dogs, this is referred to as the dog leukocyte antigen (DLA) region. Rat anti Dog MHC Class II immunoprecipitates an antigen of ~32/34 kDa and blocks the proliferation of MHC Class II dependent responses <i>in vitro</i> . In dogs, MHC Class II is expressed by all peripheral blood mononuclear cells.	
Flow Cytometry	Use 10µl of the suggested working dilution to label 10 <sup>6</sup> cells in 100µl	
References	<ol> <li>Cobbold, S. &amp; Metcalfe, S. (1994) Monoclonal antibodies that define canine homologues of human CD antigens: summary of the First International Canine Leul Antigen Workshop (CLAW). Tissue Antigens. 43 (3): 137-54.</li> <li>Watson, C.J. et al. (1994) Immunosuppression of canine renal allograft recipients CD4 and CD8 monoclonal antibodies. Tissue Antigens. 43 (3): 155-62.</li> <li>Liu, Y. et al. (2000) Immunosuppressant-free allotransplantation of the tracheaTh antigenicity of tracheal grafts can be reduced by removing the epithelium and mixed glands from the graft by detergent treatment. J Thorac Cardiovasc Surg. 120: 108-4.</li> <li>Sanchez, M.A. et al. (2004) Organ-specific immunity in canine visceral leishmania analysis of symptomatic and asymptomatic dogs naturally infected with Leishmania chagasi. Am J Trop Med Hyg. 70: 618-24.</li> <li>Reis, A.B. et al. (2006) Phenotypic features of circulating leucocytes as immunol markers for clinical status and bone marrow parasite density in dogs naturally infecteishmania chagasi. Clin Exp Immunol.146: 303-11.</li> <li>Bonnefont-Rebeix, C. et al. (2007) Toll-like receptor 3 (TLR3): a new marker of comonocytes-derived dendritic cells (cMo-DC). Vet Immunol Immunopathol. 2007 Jul. 15:118(1-2):134-9.</li> <li>Schütze, N. et al. (2009) Inactivated parapoxvirus ovis activates canine blood phagocytes and T lymphocytes. Vet Microbiol. 137: 260-7.</li> <li>Bund, D. et al. (2010) Canine-DCs using different serum-free methods as an app to provide an animal-model for immunotherapeutic strategies. Cell Immunol. 263: 8.</li> <li>Mito, K. et al. (2010) IFNy markedly cooperates with intratumoral dendritic cell vain dog tumor models. Cancer Res. 70: 7093-101.</li> <li>Araújo, M.S. et al. (2011) Immunological changes in canine peripheral blood leukocytes triggered by immunization with first or second generation vaccines again canine visceral leishmaniasis. Vet Immunol Immunopathol. 141: 64-75.</li> </ol>	

11. Larsen, A.K. *et al.* (2013) Entry and elimination of marine mammal Brucella spp. by hooded seal (Cystophora cristata) alveolar macrophages *in vitro*. <u>PLoS One. 8: e70186.</u>

- 12. Lin, S-C. *et al.* (2014) Immune Characterization of Peripheral Blood Mononuclear cells of the Dogs Restored from Innoculation of Canine Transmissible Venereal Tumor Cells. Tai Vet J. 40 (04): 181-90.
- 13. Constantinoiu, C.C. *et al.* (2015) Mucosal tolerance of the hookworm Ancylostoma caninum in the gut of naturally infected wild dogs. <u>Parasite Immunol. 37 (10): 510-20.</u>
- 14. Bonnefont-Rebeix, C. *et al.* (2016) Characterization of a novel canine T-cell line established from a spontaneously occurring aggressive T-cell lymphoma with large granular cell morphology. <a href="https://linearchy.com/
- 15. Lu, T. *et al.* (2017) Effects of cryopreservation on tracheal allograft antigenicity in dogs. <u>J Thorac Dis. 9 (7): 2038-2047.</u>
- 16. Reineking, W. *et al.* (2018) Canine primary jejunal and colonic epithelial cells predominantly express TLR5 and TLR9 but do not change TLR expression pattern after stimulation with certain Toll-like receptor ligands. Vet Immunol Immunopathol. 206: 16-24.
- 17. Martini, V. *et al.* (2019) Prognostic role of non-neoplastic lymphocytes in lymph node aspirates from dogs with diffuse large B-cell lymphoma treated with chemo-immunotherapy. Res Vet Sci. 125: 130-5.
- 18. Grudzien, M. *et al.* (2021) A newly established canine NK-type cell line and its cytotoxic properties. <u>Vet Comp Oncol. 19 (3): 567-77.</u>
- 19. Bragato, J.P. *et al.* (2022) miRNA-21 regulates CD69 and IL-10 expression in canine leishmaniasis. PLoS One. 17 (3): e0265192.
- 20. Riccardo, F. *et al.* (2022) Antigen mimicry as an effective strategy to induce CSPG4-targeted immunity in dogs with oral melanoma: a veterinary trial. <u>J Immunother Cancer. 10</u> (5): e004007.
- 21. Jaensch, S.M. *et al.* (2022) Clinicopathologic and immunophenotypic features in dogs with presumptive large granular lymphocyte leukaemia. Aust Vet J. 100 (11): 527-32.
- 22. Yang, V.K. *et al.* (2021) Intravenous administration of allogeneic Wharton jelly-derived mesenchymal stem cells for treatment of dogs with congestive heart failure secondary to myxomatous mitral valve disease. Am J Vet Res. 82 (6): 487-93.
- 23. Crain, S.K. *et al.* (2019) Extracellular Vesicles from Wharton's Jelly Mesenchymal Stem Cells Suppress CD4 Expressing T Cells Through Transforming Growth Factor Beta and Adenosine Signaling in a Canine Model. Stem Cells Dev. 28 (3): 212-26.
- 24. Martini, V. *et al.* (2018) A retrospective study of flow cytometric characterization of suspected extranodal lymphomas in dogs. <u>J Vet Diagn Invest. 30 (6): 830-6.</u>
- 25. Sainz, Á. *et al.* (2021) Effect of chemically modified tetracycline-8 (CMT-8) on hematology, blood chemistry, cytokines and peripheral blood lymphocyte subsets of healthy dogs. Res Vet Sci. 136: 200-8.
- 26. Milhau, N. *et al.* (2020) *In vitro* evaluations on canine monocyte-derived dendritic cells of a nanoparticles delivery system for vaccine antigen against *Echinococcus granulosus*. PLoS One. 15 (2): e0229121.
- 27. Sheng, R. *et al.* (2023) Prognostic significance of CD25 expression in dogs with a noninvasive diagnosis of B-cell lymphoma treated with CHOP chemotherapy. <u>Vet Comp Oncol. 21 (1): 28-35.</u>
- 28. Benavides, F.P. *et al.* (2021) Intrathecal Transplantation of Autologous and Allogeneic Bone Marrow-Derived Mesenchymal Stem Cells in Dogs. <u>Cell Transplant. 30:</u> 9636897211034464.
- 29. Stokol, T. *et al.* (2024) Flow cytometric-based detection of CD80 is a useful diagnostic marker of acute myeloid leukemia in dogs. <u>Front Vet Sci. 11: 1405297.</u>

30. Rogato, F. et al. (2024) Leukemia cutis as a prominent clinical sign in a dog with acute myeloid leukemia. Vet Clin Pathol. 53 (4): 448-457. This product is shipped at ambient temperature. It is recommended to aliquot and store at -20°C on receipt. When thawed, aliquot the sample as needed. Keep aliquots at 2-8°C for short term use (up to 4 weeks) and store the remaining aliquots at -20°C.

Avoid repeated freezing and thawing as this may denature the antibody. Storage in frost-free freezers is not recommended. This product is photosensitive and should be

protected from light.

Guarantee 12 months from date of despatch **Health And Safety** Material Safety Datasheet documentation #10041 available at: Information https://www.bio-rad-antibodies.com/SDS/MCA1044F 10041 Regulatory For research purposes only

# Related Products

## **Recommended Negative Controls**

RAT IgG2a NEGATIVE CONTROL:FITC (MCA6005F) RAT IgG2a NEGATIVE CONTROL:FITC (MCA1212F)

America

**Storage** 

North & South Tel: +1 800 265 7376 Fax: +1 919 878 3751 Worldwide

Tel: +44 (0)1865 852 700 Fax: +44 (0)1865 852 739 Europe

Tel: +49 (0) 89 8090 95 21 Fax: +49 (0) 89 8090 95 50

Email: antibody\_sales\_us@bio-rad.com

Email: antibody\_sales\_uk@bio-rad.com

Email: antibody\_sales\_de@bio-rad.com

To find a batch/lot specific datasheet for this product, please use our online search tool at: bio-rad-antibodies.com/datasheets 'M407953:221010'

#### Printed on 28 May 2025

© 2025 Bio-Rad Laboratories Inc | Legal | Imprint