

Datasheet: AAI41F

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| Description: | GOAT ANTI PIG IgG (Fc):FITC |
| Specificity: | IgG (Fc) |
| Format: | FITC |
| Product Type: | Polyclonal Antibody |
| Isotype: | Polyclonal IgG |
| Quantity: | 1 mg |

Product Details

Applications

This product has been reported to work in the following applications. This information is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for further information. For general protocol recommendations, please visit www.bio-rad-antibodies.com/protocols.

| | Yes | No | Not Determined | Suggested Dilution |
|----------------------------|-----|----|----------------|--------------------|
| Flow Cytometry | ▪ | | | 1/20 - 1/100 |
| Immunohistology - Frozen | ▪ | | | 1/20 - 1/200 |
| Immunohistology - Paraffin | | | ▪ | |

Where this antibody has not been tested for use in a particular technique this does not necessarily exclude its use in such procedures. Suggested working dilutions are given as a guide only. It is recommended that the user titrates the antibody for use in their own system using the appropriate negative/positive controls.

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|-----------------------|--|----------------------------|--------------------------|
| Target Species | Pig | | |
| Product Form | Purified IgG conjugated to Fluorescein Isothiocyanate Isomer 1 (FITC) - liquid | | |
| Max Ex/Em | Fluorophore | Excitation Max (nm) | Emission Max (nm) |
| | FITC | 490 | 525 |

Antiserum Preparation Antisera to porcine IgG were raised by repeated immunisation of goat with highly purified antigen. Purified IgG prepared by affinity chromatography.

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|---------------------------------------|-----------------------------|
| Buffer Solution | Phosphate buffered saline |
| Preservative | 0.09% Sodium Azide |
| Stabilisers | 0.2% Bovine Serum Albumin |
| Approx. Protein Concentrations | IgG concentration 1.0 mg/ml |

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|--------------------|---|
| Immunogen | Purified porcine IgG. |
| RRID | AB_323037 |
| Specificity | <p>Goat anti Pig IgG (Fc) antibody recognizes the Fc region of the porcine IgG heavy chains and shows no cross - reactivity with other porcine immunoglobulin classes as evaluated by immunoelectrophoresis. Goat anti Pig IgG (Fc) has not been species cross adsorbed and may react with the Fc region of IgG from other species.</p> <p>Goat anti Pig IgG (Fc) antibody has been used extensively as a detection reagent for porcine IgG in ELISA, for example monitoring of the IgG response in influenza infected pigs (Crisci et al. 2013).</p> |
| References | <ol style="list-style-type: none"> Scharek, L. et al. (2005) Influence of a probiotic <i>Enterococcus faecium</i> strain on development of the immune system of sows and piglets. Vet Immunol Immunopathol. 105: 151-61. Scharek, L. et al. (2007) Impact of the probiotic bacteria <i>Enterococcus faecium</i> NCIMB 10415 (SF68) and <i>Bacillus cereus</i> var. <i>toyoi</i> NCIMB 40112 on the development of serum IgG and faecal IgA of sows and their piglets. Arch Anim Nutr. 61: 223-34. Kang, M.L. et al. (2008) Chitosan microspheres containing <i>Bordetella bronchiseptica</i> antigens as novel vaccine against atrophic rhinitis in pigs. J Microbiol Biotechnol. 18: 1179-85. Kim, T. et al. (2009) <i>Bordetella bronchiseptica</i> aroA mutant as a live vaccine vehicle for heterologous porcine circovirus type 2 major capsid protein expression. Vet Microbiol. 138: 318-24. Tsai, Y.C. et al. (2010) Porcine circovirus type 2 (PCV2) induces cell proliferation, fusion, and chemokine expression in swine monocytic cells <i>in vitro</i>. Vet Res. 41: 60. Assana, E. et al. (2010) Antibody responses to the host-protective <i>Taenia solium</i> oncosphere protein TSOL18 in pigs are directed against conformational epitopes. Parasite Immunol. 32: 399-405. Pyo, H. et al. (2010) Serodiagnosis of porcine reproductive and respiratory syndrome virus infection with the use of glycoprotein 5 antigens. Can J Vet Res. 74: 223-7. Busquets, N. et al. (2010) Experimental infection with H1N1 European swine influenza virus protects pigs from an infection with the 2009 pandemic H1N1 human influenza virus. Vet Res. 41: 74. Kick, A.R. et al. (2011) Evaluation of peripheral lymphocytes after weaning and vaccination for <i>Mycoplasma hyopneumoniae</i>. Res Vet Sci. 91 (3): e68-72. Sheoran, A. et al. (2012) Infection With <i>Cryptosporidium hominis</i> Provides Incomplete Protection of the Host Against <i>Cryptosporidium parvum</i>. J Infect Dis 205: 1019-23. Fleury, A. et al. (2015) <i>Taenia solium</i>: Development of an Experimental Model of Porcine Neurocysticercosis. PLoS Negl Trop Dis. 9 (8): e0003980. Zhang, Y. et al. (2016) Generation of <i>E. coli</i>-derived virus-like particles of porcine circovirus type 2 and their use in an indirect IgG enzyme-linked immunosorbent assay. Arch Virol. 161 (6): 1485-91. Lee, J.A. et al. (2016) <i>Mycoplasma hyorhinis</i> is a potential pathogen of porcine respiratory disease complex that aggravates pneumonia caused by porcine reproductive and respiratory syndrome virus. Vet Immunol Immunopathol. 177: 48-51. Lorenzen, E. et al. (2017) Intrauterine inoculation of minipigs with <i>Chlamydia</i> |

trachomatis during diestrus establishes a longer lasting infection compared to vaginal inoculation during estrus. [Microbes Infect. 19 \(6\): 334-42.](#)

15. Morales, J. *et al.* (2018) Persistent *Taenia solium*. Cysticercosis In the State of Morelos, Mexico: Human and Porcine Seroprevalence. [J Parasitol. 104 \(5\): 465-72.](#)

16. Zhang, G. *et al.* (2019) Identification of the B-cell epitopes on N protein of type 2 porcine reproductive and respiratory syndrome virus, using monoclonal antibodies. [Int J Biol Macromol. 130: 300-6.](#)

17. Tan, T.K. *et al.* (2021) A COVID-19 vaccine candidate using SpyCatcher multimerization of the SARS-CoV-2 spike protein receptor-binding domain induces potent neutralising antibody responses. [Nat Commun. 12 \(1\): 542.](#)

18. Lentsch, V. *et al.* (2023) "EvoVax" - A rationally designed inactivated *Salmonella typhimurium* vaccine induces strong and long-lasting immune responses in pigs. [Vaccine. 41 \(38\): 5545-52.](#)

19. Hernández-Chea, R. *et al.* (2024) Epidemiology of swine cysticercosis in two rural communities of Zacapa, Guatemala. [Vet Parasitol Reg Stud Reports. 47: 100951.](#)

20. Somda, M.B. *et al.* (2022) Evaluation of antibody responses to tsetse fly saliva in domestic animals in the sleeping sickness endemic foci of Bonon and Sinfra, Côte d'Ivoire. [Vet Parasitol Reg Stud Reports. 34: 100773.](#)

21. Stepanova, K. *et al.* (2023) Modified live vaccine strains of porcine reproductive and respiratory syndrome virus cause immune system dysregulation similar to wild strains. [Front Immunol. 14: 1292381.](#)

Storage Store at +4°C. DO NOT FREEZE.
This product should be stored undiluted. Should this product contain a precipitate we recommend microcentrifugation before use.

Guarantee 12 months from date of despatch

Health And Safety Information Material Safety Datasheet documentation #10041 available at: <https://www.bio-rad-antibodies.com/SDS/AAI41F>
10041

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