

Datasheet: AAI40B BATCH NUMBER 158140

Description:	GOAT ANTI PIG IgA:Biotin
Specificity:	IgA
Format:	Biotin
Product Type:	Polyclonal Antibody
Isotype:	Polyclonal IgG
Quantity:	1 mg

Product Details

Applications

This product has been reported to work in the following applications. This information is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for further information. For general protocol recommendations, please visit www.bio-rad-antibodies.com/protocols.

	Yes	No	Not Determined	Suggested Dilution
Flow Cytometry				
Immunohistology - Frozen				
Immunohistology - Paraffin				
ELISA	•			1:10000 - 1:100000
Western Blotting	-			1:10000 - 1:100000

Where this product has not been tested for use in a particular technique this does not necessarily exclude its use in such procedures. Suggested working dilutions are given as a guide only. It is recommended that the user titrates the product for use in their own system using appropriate negative/positive controls.

Target Species	Pig
Product Form	Purified IgG conjugated to Biotin - liquid

Antiserum Preparation Antisera to porcine IgA were raised by repeated immunisation of goat with highly purified antigen. Purified IgG was prepared by affinity chromatography.

Buffer Solution	Phosphate buffered saline
Preservative Stabilisers	0.09% Sodium Azide (NaN ₃) 0.2% Bovine Serum Albumin
Approx. Protein Concentrations	IgG concentration 1.0 mg/ml

Immunogen	Purified porcine IgA.
RRID	AB_10675496
Specificity	Goat anti pig IgA antibody recognizes porcine IgA and shows no cross-reactivity with other porcine immunoglobulin classes as assessed by immunoelectrophoresis. This antibody may cross-react with IgA from other species.
	Goat anti Porcine IgA antibody has been succesfully used for the evaluation of porcine IgA levels in body fluids of pigs by both ELISA and Western blotting.
References	1. Takahashi, M. <i>et al</i> (2005) Correlation between positivity for immunoglobulin A antibodies and viraemia of swine hepatitis E virus observed among farm pigs in Japan. J. Gen Virol. 86: 1807-13. 2. Scharek, L. <i>et al.</i> (2005) Influence of a probiotic <i>Enterococcus faecium</i> strain on development of the immune system of sows and piglets. Vet Immunol Immunopathol. 105: 151-61. 3. Nakai, I. <i>et al.</i> (2006) Different fecal shedding patterns of two common strains of hepatitis E virus at three Japanese swine farms. Am J Trop Med Hyg. 75: 1171-7. 4. Zhang, L. <i>et al.</i> (2007) Intranasal administration of CpG oligonucleotides induces mucosal and systemic Type 1 immune responses and adjuvant activity to porcine reproductive and respiratory syndrome killed virus vaccine in piglets <i>in vivo</i> . Int Immunopharmacol. 7: 1732-40. 5. Bestagno, M. <i>et al.</i> (2007) Recombinant dimeric small immunoproteins neutralize transmissible gastroenteritis virus infectivity efficiently <i>in vitro</i> and confer passive immunity <i>in vivo</i> . J Gen Virol. 88: 187-95. 6. Bestagno, M. <i>et al.</i> (2007) Recombinant dimeric small immunoproteins neutralize transmissible gastroenteritis virus infectivity efficiently <i>in vitro</i> and confer passive immunity <i>in vivo</i> . J Gen Virol. 88: 187-95.
	 Picherot, M. et al. (2007) Swine infection with <i>Trichinella spiralis</i>: Comparative analysis of the mucosal intestinal and systemic immune responses. <u>Vet Parasitol. 143: 122-30.</u> Scharek, L. et al. (2007) Impact of the probiotic bacteria <i>Enterococcus faecium</i> NCIMB 10415 (SF68) and <i>Bacillus cereus</i> var. <i>toyoi</i> NCIMB 40112 on the development of serum IgG and faecal IgA of sows and their piglets. <u>Arch Anim Nutr. 61: 223-34.</u> Eblé, P.L. et al. (2007) Serological and mucosal immune responses after vaccination

antigens as novel vaccine against atrophic rhinitis in pigs. <u>J Microbiol Biotechnol. 18:</u>
1179-85.

11. Linghua, Z. *et al.* (2008) *In vivo* oral administration effects of various oligodeoxynucleotides containing synthetic immunostimulatory motifs in the immune.

10. Kang, M.L. et al. (2008) Chitosan microspheres containing Bordetella bronchiseptica

- 11. Linghua, Z. *et al.* (2008) *In vivo* oral administration effects of various oligodeoxynucleotides containing synthetic immunostimulatory motifs in the immune response to pseudorabies attenuated virus vaccine in newborn piglets. <u>Vaccine. 26 (2):</u> 224-33.
- 12. Olvera, A. *et al.* (2010) Virulence-associated trimeric autotransporters of *Haemophilus parasuis* are antigenic proteins expressed *in vivo*. <u>Vet Res. 41: 26.</u>
- 13. Sheoran A *et al.* (2012) Infection with *Cryptosporidium hominis* provides incomplete protection of the host against *Cryptosporidium parvum*. <u>J Infect Dis. 205 (6): 1019-23.</u>
- 14. Cordes, H. et al. (2012) Cell-mediated and humoral immune responses in pigs

and infection with FMDV in pigs. Vaccine. 25: 1043-54.

following primary and challenge-exposure to Lawsonia intracellularis. Vet Res. 43:9.

- 15. Crisci, E. *et al.* (2014) Immune characterization of long pentraxin 3 in pigs infected with influenza virus. Vet Microbiol. 168 (1): 185-92.
- 16. Le Bourgot, C. *et al.* (2016) Short-chain fructooligosaccharide supplementation during gestation and lactation or after weaning differentially impacts pig growth and IgA response to influenza vaccination Journal of Functional Foods. 24: 307-15.
- 17. Lorenzen, E. *et al.* (2017) Intrauterine inoculation of minipigs with *Chlamydia trachomatis*. during diestrus establishes a longer lasting infection compared to vaginal inoculation during estrus. Microbes Infect. 19 (6): 334-42.
- 18. Williams, A.R. *et al.* (2017) Dietary cinnamaldehyde enhances acquisition of specific antibodies following helminth infection in pigs. Vet Immunol Immunopathol. 189: 43-52.
- 19. Williams, A.R. *et al.* (2017) A polyphenol-enriched diet and *Ascaris suum*. infection modulate mucosal immune responses and gut microbiota composition in pigs. <u>PLoS One.</u> 12 (10): e0186546.
- 20. López-Serrano, S. *et al.* (2021) Sow Vaccination with a Protein Fragment against Virulent *Glaesserella (Haemophilus) parasuis*. Modulates Immunity Traits in Their Offspring. <u>Vaccines. 9 (5): 534.</u>
- 21. Tan, T.K. *et al.* (2021) A COVID-19 vaccine candidate using SpyCatcher multimerization of the SARS-CoV-2 spike protein receptor-binding domain induces potent neutralising antibody responses. <u>Nat Commun. 12 (1): 542.</u>

Storage	Store at +4°C. DO NOT FREEZE. This product should be stored undiluted. Should this product contain a precipitate we recommend microcentrifugation before use.
Guarantee	12 months from date of despatch
Health And Safety Information	Material Safety Datasheet documentation #10041 available at: https://www.bio-rad-antibodies.com/SDS/AAI40B 10041

 North & South
 Tel: +1 800 265 7376
 Worldwide
 Tel: +44 (0)1865 852 700
 Europe
 Tel: +49 (0) 89 8090 95 21

 America
 Fax: +1 919 878 3751
 Fax: +44 (0)1865 852 739
 Fax: +49 (0) 89 8090 95 50

For research purposes only

To find a batch/lot specific datasheet for this product, please use our online search tool at: bio-rad-antibodies.com/datasheets 'M363651:200528'

Printed on 01 Mar 2024

© 2024 Bio-Rad Laboratories Inc | Legal | Imprint

Regulatory