

## Datasheet: AAI40AB

<b>Description:</b>	GOAT ANTI PIG IgA:Alk. Phos.
<b>Specificity:</b>	IgA
<b>Format:</b>	Alk. Phos.
<b>Product Type:</b>	Polyclonal Antibody
<b>Isotype:</b>	Polyclonal IgG
<b>Quantity:</b>	0.5 mg

## Product Details

### Applications

This product has been reported to work in the following applications. This information is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for further information. For general protocol recommendations, please visit [www.bio-rad-antibodies.com/protocols](http://www.bio-rad-antibodies.com/protocols).

	Yes	No	Not Determined	Suggested Dilution
Immunohistology - Frozen	▪			1/100 - 1/1000
Immunohistology - Paraffin			▪	
ELISA	▪			1/1000 - 1/10000
Western Blotting	▪			1/1000 - 1/10000

Where this antibody has not been tested for use in a particular technique this does not necessarily exclude its use in such procedures. Suggested working dilutions are given as a guide only. It is recommended that the user titrates the antibody for use in their own system using the appropriate negative/positive controls.

<b>Target Species</b>	Pig
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<b>Product Form</b>	Purified IgG conjugated to Alkaline Phosphatase - liquid
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**Antiserum Preparation** Antisera to porcine IgA were raised by repeated immunisation of goat with highly purified antigen. Purified IgG prepared by affinity chromatography.

<b>Buffer Solution</b>	50mM HEPES 0.1M NaCl 1mM MgCl <sub>2</sub> 0.1mM ZnCl <sub>2</sub>
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<b>Preservative</b>	0.09% Sodium Azide
<b>Stabilisers</b>	0.2% Bovine Serum Albumin

<b>Approx. Protein Concentrations</b>	IgG concentration 0.5mg/ml
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<b>Immunogen</b>	Purified porcine IgA.
<b>RRID</b>	AB_10846108
<b>Specificity</b>	<p><b>Goat anti pig IgA antibody</b> recognizes porcine IgA and shows no cross-reactivity with other porcine immunoglobulin classes as assessed by immunoelectrophoresis. This antibody may cross-react with IgA from other species.</p> <p>Goat anti Porcine IgA antibody has been successfully used for the evaluation of porcine IgA levels in body fluids of pigs by both ELISA and Western blotting.</p>
<b>References</b>	<ol style="list-style-type: none"> <li>1. Takahashi, M. <i>et al.</i> (2005) Correlation between positivity for immunoglobulin A antibodies and viraemia of swine hepatitis E virus observed among farm pigs in Japan. <a href="#">J Gen Virol. 86: 1807-13.</a></li> <li>2. Linghua, Z. <i>et al.</i> (2008) <i>In vivo oral administration effects of various oligodeoxynucleotides containing synthetic immunostimulatory motifs in the immune response to pseudorabies attenuated virus vaccine in newborn piglets.</i> <a href="#">Vaccine. 26 (2): 224-33.</a></li> <li>3. Olvera, A. <i>et al.</i> (2010) Virulence-associated trimeric autotransporters of <i>Haemophilus parasuis</i> are antigenic proteins expressed <i>in vivo</i>. <a href="#">Vet Res. 41: 26.</a></li> <li>4. Scharek, L. <i>et al.</i> (2005) Influence of a probiotic <i>Enterococcus faecium</i> strain on development of the immune system of sows and piglets. <a href="#">Vet Immunol Immunopathol. 105: 151-61.</a></li> <li>5. Scharek, L. <i>et al.</i> (2007) Impact of the probiotic bacteria <i>Enterococcus faecium</i> NCIMB 10415 (SF68) and <i>Bacillus cereus</i> var. <i>toyoi</i> NCIMB 40112 on the development of serum IgG and faecal IgA of sows and their piglets. <a href="#">Arch Anim Nutr. 61: 223-34.</a></li> <li>6. Eblé, P.L. <i>et al.</i> (2007) Serological and mucosal immune responses after vaccination and infection with FMDV in pigs. <a href="#">Vaccine. 25: 1043-54.</a></li> <li>7. Bestagno, M. <i>et al.</i> (2007) Recombinant dimeric small immunoproteins neutralize transmissible gastroenteritis virus infectivity efficiently <i>in vitro</i> and confer passive immunity <i>in vivo</i>. <a href="#">J Gen Virol. 88: 187-95.</a></li> <li>8. Nakai, I. <i>et al.</i> (2006) Different fecal shedding patterns of two common strains of hepatitis E virus at three Japanese swine farms. <a href="#">Am J Trop Med Hyg. 75: 1171-7.</a></li> <li>9. Kang, M.L. <i>et al.</i> (2008) Chitosan microspheres containing <i>Bordetella bronchiseptica</i> antigens as novel vaccine against atrophic rhinitis in pigs. <a href="#">J Microbiol Biotechnol. 18: 1179-85.</a></li> <li>10. Picherot, M. <i>et al.</i> (2007) Swine infection with <i>Trichinella spiralis</i>: Comparative analysis of the mucosal intestinal and systemic immune responses. <a href="#">Vet Parasitol. 143: 122-30.</a></li> <li>11. Sheoran A <i>et al.</i> (2012) Infection with <i>Cryptosporidium hominis</i> provides incomplete protection of the host against <i>Cryptosporidium parvum</i>. <a href="#">J Infect Dis. 205 (6): 1019-23.</a></li> <li>12. Bestagno, M. <i>et al.</i> (2007) Recombinant dimeric small immunoproteins neutralize transmissible gastroenteritis virus infectivity efficiently <i>in vitro</i> and confer passive immunity <i>in vivo</i>. <a href="#">J Gen Virol. 88: 187-95.</a></li> <li>13. Cordes, H. <i>et al.</i> (2012) Cell-mediated and humoral immune responses in pigs following primary and challenge-exposure to <i>Lawsonia intracellularis</i>. <a href="#">Vet Res. 43:9.</a></li> <li>14. Crisci, E. <i>et al.</i> (2014) Immune characterization of long pentraxin 3 in pigs infected with influenza virus. <a href="#">Vet Microbiol. 168 (1): 185-92.</a></li> </ol>

15. Le Bourgot, C, et al. (2016) Short-chain fructooligosaccharide supplementation during gestation and lactation or after weaning differentially impacts pig growth and IgA response to influenza vaccination [J Funct Foods. 24: 307-15.](#)
16. Lorenzen, E. et al. (2017) Intrauterine inoculation of minipigs with *Chlamydia trachomatis* during diestrus establishes a longer lasting infection compared to vaginal inoculation during estrus. [Microbes Infect. Feb 9. pii: S1286-4579\(17\)30025-4. \[Epub ahead of print\]](#)
17. Williams, A.R. et al. (2017) Dietary cinnamaldehyde enhances acquisition of specific antibodies following helminth infection in pigs. [Vet Immunol Immunopathol. 189: 43-52.](#)
18. Williams, A.R. et al. (2017) A polyphenol-enriched diet and *Ascaris suum* infection modulate mucosal immune responses and gut microbiota composition in pigs. [PLoS One. 12 \(10\): e0186546.](#)

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**Storage**

Store at +4°C.  
DO NOT FREEZE.

This product should be stored undiluted.  
Should this product contain a precipitate we recommend microcentrifugation before use.

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**Guarantee**

12 months from date of despatch

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**Health And Safety Information**

Material Safety Datasheet documentation #10089 available at:  
10089: <https://www.bio-rad-antibodies.com/uploads/MSDS/10089.pdf>

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**Regulatory**

For research purposes only

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