

Datasheet: AAI40

Description:	GOAT ANTI PIG IgA
Specificity:	IgA
Format:	Purified
Product Type:	Polyclonal Antibody
Isotype:	Polyclonal IgG
Quantity:	1 mg

Product Details

Applications

This product has been reported to work in the following applications. This information is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for further information. For general protocol recommendations, please visit www.bio-rad-antibodies.com/protocols.

	Yes	No	Not Determined	Suggested Dilution
Flow Cytometry			▪	
Immunohistology - Frozen			▪	
Immunohistology - Paraffin			▪	
ELISA	▪			1/100 - 1/30000
Immunodiffusion	▪			

Where this antibody has not been tested for use in a particular technique this does not necessarily exclude its use in such procedures. Suggested working dilutions are given as a guide only. It is recommended that the user titrates the antibody for use in their own system using the appropriate negative/positive controls.

Target Species	Pig
Product Form	Purified IgG - liquid
Antiserum Preparation	Antisera to porcine IgA were raised by repeated immunisation of goat with highly purified antigen. Purified IgG prepared by affinity chromatography.
Buffer Solution	Phosphate buffered saline
Preservative Stabilisers	0.09% Sodium Azide
Approx. Protein Concentrations	IgG concentration 1.0 mg/ml
Immunogen	Purified porcine IgA.

Specificity

Goat anti pig IgA antibody recognizes porcine IgA and shows no cross-reactivity with other porcine immunoglobulin classes as assessed by immunoelectrophoresis. This antibody may cross-react with IgA from other species.

Goat anti Porcine IgA antibody has been successfully used for the evaluation of porcine IgA levels in body fluids of pigs by both ELISA and Western blotting.

References

1. Takahashi, M. *et al* (2005) Correlation between positivity for immunoglobulin A antibodies and viraemia of swine hepatitis E virus observed among farm pigs in Japan. [J Gen Virol. 86: 1807-13.](#)
2. Scharek, L. *et al*. (2005) Influence of a probiotic *Enterococcus faecium* strain on development of the immune system of sows and piglets. [Vet Immunol Immunopathol. 105: 151-61.](#)
3. Nakai, I. *et al*. (2006) Different fecal shedding patterns of two common strains of hepatitis E virus at three Japanese swine farms. [Am J Trop Med Hyg. 75: 1171-7.](#)
4. Zhang, L. *et al*. (2007) Intranasal administration of CpG oligonucleotides induces mucosal and systemic Type 1 immune responses and adjuvant activity to porcine reproductive and respiratory syndrome killed virus vaccine in piglets *in vivo*. [Int Immunopharmacol. 7: 1732-40.](#)
5. Bestagno, M. *et al*. (2007) Recombinant dimeric small immunoproteins neutralize transmissible gastroenteritis virus infectivity efficiently *in vitro* and confer passive immunity *in vivo*. [J Gen Virol. 88: 187-95.](#)
6. Bestagno, M. *et al*. (2007) Recombinant dimeric small immunoproteins neutralize transmissible gastroenteritis virus infectivity efficiently *in vitro* and confer passive immunity *in vivo*. [J Gen Virol. 88: 187-95.](#)
7. Picherot, M. *et al*. (2007) Swine infection with *Trichinella spiralis*: Comparative analysis of the mucosal intestinal and systemic immune responses. [Vet Parasitol. 143: 122-30.](#)
8. Scharek, L. *et al*. (2007) Impact of the probiotic bacteria *Enterococcus faecium* NCIMB 10415 (SF68) and *Bacillus cereus* var. *toyoi* NCIMB 40112 on the development of serum IgG and faecal IgA of sows and their piglets. [Arch Anim Nutr. 61: 223-34.](#)
9. Eblé, P.L. *et al*. (2007) Serological and mucosal immune responses after vaccination and infection with FMDV in pigs. [Vaccine. 25: 1043-54.](#)
10. Kang, M.L. *et al*. (2008) Chitosan microspheres containing *Bordetella bronchiseptica* antigens as novel vaccine against atrophic rhinitis in pigs. [J Microbiol Biotechnol. 18: 1179-85.](#)
11. Linghua, Z. *et al*. (2008) *In vivo* oral administration effects of various oligodeoxynucleotides containing synthetic immunostimulatory motifs in the immune response to pseudorabies attenuated virus vaccine in newborn piglets. [Vaccine. 26 \(2\): 224-33.](#)
12. Olvera, A. *et al*. (2010) Virulence-associated trimeric autotransporters of *Haemophilus parasuis* are antigenic proteins expressed *in vivo*. [Vet Res. 41: 26.](#)
13. Sheoran A *et al*. (2012) Infection with *Cryptosporidium hominis* provides incomplete protection of the host against *Cryptosporidium parvum*. [J Infect Dis. 205 \(6\): 1019-23.](#)
14. Cordes, H. *et al*. (2012) Cell-mediated and humoral immune responses in pigs following primary and challenge-exposure to *Lawsonia intracellularis*. [Vet Res. 43:9.](#)

15. Crisci, E. *et al.* (2014) Immune characterization of long pentraxin 3 in pigs infected with influenza virus. [Vet Microbiol. 168 \(1\): 185-92.](#)
16. Le Bourgot, C. *et al.* (2016) Short-chain fructooligosaccharide supplementation during gestation and lactation or after weaning differentially impacts pig growth and IgA response to influenza vaccination [Journal of Functional Foods. 24: 307-15.](#)
17. Lorenzen, E. *et al.* (2017) Intrauterine inoculation of minipigs with *Chlamydia trachomatis*. during diestrus establishes a longer lasting infection compared to vaginal inoculation during estrus. [Microbes Infect. 19 \(6\): 334-42.](#)
18. Williams, A.R. *et al.* (2017) Dietary cinnamaldehyde enhances acquisition of specific antibodies following helminth infection in pigs. [Vet Immunol Immunopathol. 189: 43-52.](#)
19. Williams, A.R. *et al.* (2017) A polyphenol-enriched diet and *Ascaris suum*. infection modulate mucosal immune responses and gut microbiota composition in pigs. [PLoS One. 12 \(10\): e0186546.](#)
20. López-Serrano, S. *et al.* (2021) Sow Vaccination with a Protein Fragment against Virulent *Glaesserella (Haemophilus) parasuis*. Modulates Immunity Traits in Their Offspring. [Vaccines. 9 \(5\): 534.](#)
21. Tan, T.K. *et al.* (2021) A COVID-19 vaccine candidate using SpyCatcher multimerization of the SARS-CoV-2 spike protein receptor-binding domain induces potent neutralising antibody responses. [Nat Commun. 12 \(1\): 542.](#)
22. López-Serrano, S. *et al.* (2023) Immune responses following neonatal vaccination with conserved F4 fragment of VtaA proteins from virulent *Glaesserella parasuis* adjuvanted with CAF®01 or CDA. [Vaccine X. 14: 100330.](#)
23. Michael, H. *et al.* (2020) Malnutrition Decreases Antibody Secreting Cell Numbers Induced by an Oral Attenuated Human Rotavirus Vaccine in a Human Infant Fecal Microbiota Transplanted Gnotobiotic Pig Model. [Front Immunol. 11: 196.](#)

Storage Store at +4°C. DO NOT FREEZE.
This product should be stored undiluted. Should this product contain a precipitate we recommend microcentrifugation before use.

Guarantee 12 months from date of despatch

Health And Safety Information Material Safety Datasheet documentation #10040 available at: <https://www.bio-rad-antibodies.com/SDS/AAI40>
10040

Regulatory For research purposes only

Related Products

Recommended Secondary Antibodies

Rabbit Anti Goat IgG (Fc) (STAR122...) [FITC](#), [HRP](#)

North & South Tel: +1 800 265 7376

America Fax: +1 919 878 3751

Email: antibody_sales_us@bio-rad.com

Worldwide

Tel: +44 (0)1865 852 700

Fax: +44 (0)1865 852 739

Email: antibody_sales_uk@bio-rad.com

Europe

Tel: +49 (0) 89 8090 95 21

Fax: +49 (0) 89 8090 95 50

Email: antibody_sales_de@bio-rad.com

To find a batch/lot specific datasheet for this product, please use our online search tool at: [bio-rad-antibodies.com/datasheets](https://www.bio-rad-antibodies.com/datasheets)

'M428079:240301'

