

## Datasheet: AAI40

**BATCH NUMBER 149207**

<b>Description:</b>	GOAT ANTI PIG IgA
<b>Specificity:</b>	IgA
<b>Format:</b>	Purified
<b>Product Type:</b>	Polyclonal Antibody
<b>Isotype:</b>	Polyclonal IgG
<b>Quantity:</b>	1 mg

### Product Details

#### Applications

This product has been reported to work in the following applications. This information is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for further information. For general protocol recommendations, please visit [www.bio-rad-antibodies.com/protocols](http://www.bio-rad-antibodies.com/protocols).

	Yes	No	Not Determined	Suggested Dilution
Flow Cytometry			▪	
Immunohistology - Frozen			▪	
Immunohistology - Paraffin			▪	
ELISA	▪			1/100 - 1/30000
Immunodiffusion	▪			

Where this antibody has not been tested for use in a particular technique this does not necessarily exclude its use in such procedures. Suggested working dilutions are given as a guide only. It is recommended that the user titrates the antibody for use in their own system using the appropriate negative/positive controls.

#### Target Species

Pig

#### Product Form

Purified IgG - liquid

#### Antiserum Preparation

Antisera to porcine IgA were raised by repeated immunisation of goat with highly purified antigen. Purified IgG prepared by affinity chromatography.

#### Buffer Solution

Phosphate buffered saline

#### Preservative Stabilisers

0.09% Sodium Azide

#### Approx. Protein Concentrations

IgG concentration 1.0 mg/ml

Immunogen	Purified porcine IgA.
RRID	AB_323041
Specificity	<p><b>Goat anti pig IgA antibody</b> recognizes porcine IgA and shows no cross-reactivity with other porcine immunoglobulin classes as assessed by immunoelectrophoresis. This antibody may cross-react with IgA from other species.</p> <p>Goat anti Porcine IgA antibody has been successfully used for the evaluation of porcine IgA levels in body fluids of pigs by both ELISA and Western blotting.</p>
References	<ol style="list-style-type: none"> <li>1. Takahashi, M. <i>et al</i> (2005) Correlation between positivity for immunoglobulin A antibodies and viraemia of swine hepatitis E virus observed among farm pigs in Japan. <a href="#">J Gen Virol. 86: 1807-13.</a></li> <li>2. Scharek, L. <i>et al</i>. (2005) Influence of a probiotic <i>Enterococcus faecium</i> strain on development of the immune system of sows and piglets. <a href="#">Vet Immunol Immunopathol. 105: 151-61.</a></li> <li>3. Nakai, I. <i>et al</i>. (2006) Different fecal shedding patterns of two common strains of hepatitis E virus at three Japanese swine farms. <a href="#">Am J Trop Med Hyg. 75: 1171-7.</a></li> <li>4. Zhang, L. <i>et al</i>. (2007) Intranasal administration of CpG oligonucleotides induces mucosal and systemic Type 1 immune responses and adjuvant activity to porcine reproductive and respiratory syndrome killed virus vaccine in piglets <i>in vivo</i>. <a href="#">Int Immunopharmacol. 7: 1732-40.</a></li> <li>5. Bestagno, M. <i>et al</i>. (2007) Recombinant dimeric small immunoproteins neutralize transmissible gastroenteritis virus infectivity efficiently <i>in vitro</i> and confer passive immunity <i>in vivo</i>. <a href="#">J Gen Virol. 88: 187-95.</a></li> <li>6. Bestagno, M. <i>et al</i>. (2007) Recombinant dimeric small immunoproteins neutralize transmissible gastroenteritis virus infectivity efficiently <i>in vitro</i> and confer passive immunity <i>in vivo</i>. <a href="#">J Gen Virol. 88: 187-95.</a></li> <li>7. Picherot, M. <i>et al</i>. (2007) Swine infection with <i>Trichinella spiralis</i>: Comparative analysis of the mucosal intestinal and systemic immune responses. <a href="#">Vet Parasitol. 143: 122-30.</a></li> <li>8. Scharek, L. <i>et al</i>. (2007) Impact of the probiotic bacteria <i>Enterococcus faecium</i> NCIMB 10415 (SF68) and <i>Bacillus cereus</i> var. <i>toyoi</i> NCIMB 40112 on the development of serum IgG and faecal IgA of sows and their piglets. <a href="#">Arch Anim Nutr. 61: 223-34.</a></li> <li>9. Eblé, P.L. <i>et al</i>. (2007) Serological and mucosal immune responses after vaccination and infection with FMDV in pigs. <a href="#">Vaccine. 25: 1043-54.</a></li> <li>10. Kang, M.L. <i>et al</i>. (2008) Chitosan microspheres containing <i>Bordetella bronchiseptica</i> antigens as novel vaccine against atrophic rhinitis in pigs. <a href="#">J Microbiol Biotechnol. 18: 1179-85.</a></li> <li>11. Linghua, Z. <i>et al</i>. (2008) <i>In vivo</i> oral administration effects of various oligodeoxynucleotides containing synthetic immunostimulatory motifs in the immune response to pseudorabies attenuated virus vaccine in newborn piglets. <a href="#">Vaccine. 26 (2): 224-33.</a></li> <li>12. Olvera, A. <i>et al</i>. (2010) Virulence-associated trimeric autotransporters of <i>Haemophilus parasuis</i> are antigenic proteins expressed <i>in vivo</i>. <a href="#">Vet Res. 41: 26.</a></li> <li>13. Sheoran A <i>et al</i>. (2012) Infection with <i>Cryptosporidium hominis</i> provides incomplete protection of the host against <i>Cryptosporidium parvum</i>. <a href="#">J Infect Dis. 205 (6): 1019-23.</a></li> <li>14. Cordes, H. <i>et al</i>. (2012) Cell-mediated and humoral immune responses in pigs</li> </ol>

- following primary and challenge-exposure to *Lawsonia intracellularis*. [Vet Res. 43:9.](#)
15. Crisci, E. *et al.* (2014) Immune characterization of long pentraxin 3 in pigs infected with influenza virus. [Vet Microbiol. 168 \(1\): 185-92.](#)
16. Le Bourgot, C. *et al.* (2016) Short-chain fructooligosaccharide supplementation during gestation and lactation or after weaning differentially impacts pig growth and IgA response to influenza vaccination [Journal of Functional Foods. 24: 307-15.](#)
17. Lorenzen, E. *et al.* (2017) Intrauterine inoculation of minipigs with *Chlamydia trachomatis*. during diestrus establishes a longer lasting infection compared to vaginal inoculation during estrus. [Microbes Infect. 19 \(6\): 334-42.](#)
18. Williams, A.R. *et al.* (2017) Dietary cinnamaldehyde enhances acquisition of specific antibodies following helminth infection in pigs. [Vet Immunol Immunopathol. 189: 43-52.](#)
19. Williams, A.R. *et al.* (2017) A polyphenol-enriched diet and *Ascaris suum*. infection modulate mucosal immune responses and gut microbiota composition in pigs. [PLoS One. 12 \(10\): e0186546.](#)
20. López-Serrano, S. *et al.* (2021) Sow Vaccination with a Protein Fragment against Virulent *Glaesserella (Haemophilus) parasuis*. Modulates Immunity Traits in Their Offspring. [Vaccines. 9 \(5\): 534.](#)
21. Tan, T.K. *et al.* (2021) A COVID-19 vaccine candidate using SpyCatcher multimerization of the SARS-CoV-2 spike protein receptor-binding domain induces potent neutralising antibody responses. [Nat Commun. 12 \(1\): 542.](#)

<b>Storage</b>	Store at +4°C. DO NOT FREEZE. This product should be stored undiluted. Should this product contain a precipitate we recommend microcentrifugation before use.
<b>Guarantee</b>	12 months from date of despatch
<b>Health And Safety Information</b>	Material Safety Datasheet documentation #10040 available at: <a href="https://www.bio-rad-antibodies.com/SDS/AAI40">https://www.bio-rad-antibodies.com/SDS/AAI40</a> 10040
<b>Regulatory</b>	For research purposes only

## Related Products

### Recommended Secondary Antibodies

Rabbit Anti Goat IgG (Fc) (STAR122...) [FITC](#), [HRP](#)

<b>North &amp; South America</b>	Tel: +1 800 265 7376 Fax: +1 919 878 3751 Email: <a href="mailto:antibody_sales_us@bio-rad.com">antibody_sales_us@bio-rad.com</a>	<b>Worldwide</b>	Tel: +44 (0)1865 852 700 Fax: +44 (0)1865 852 739 Email: <a href="mailto:antibody_sales_uk@bio-rad.com">antibody_sales_uk@bio-rad.com</a>	<b>Europe</b>	Tel: +49 (0) 89 8090 95 21 Fax: +49 (0) 89 8090 95 50 Email: <a href="mailto:antibody_sales_de@bio-rad.com">antibody_sales_de@bio-rad.com</a>
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To find a batch/lot specific datasheet for this product, please use our online search tool at: [bio-rad-antibodies.com/datasheets](https://www.bio-rad-antibodies.com/datasheets)

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