

Datasheet: 1351115

Description:	VIVAFIX™ 498/521 CELL VIABILITY ASSAY
Name:	VIVAFIX™ ASSAY
Format:	498/521
Product Type:	Accessory Reagent
Quantity:	200 TESTS

Product Details

Applications	This product has been reported to work in the following applications. This information is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for further information. For general protocol recommendations, please visit <u>www.bio-rad-antibodies.com/protocols</u> .						
		Yes	No	Not Determined	Suggested Dilution		
	Flow Cytometry				1/500		
	Where this product has not been tested for use in a particular technique this does not necessarily exclude its use in such procedures. Suggested working dilutions are given as a guide only. It is recommended that the user titrates the product for use in their own system using appropriate negative/positive controls.						
Product Information	ion VivaFix cell viability assays are easy-to-use, versatile solutions for assessing the via of mammalian cells by flow cytometry and microscopy. In most flow cytometry experiments, the exclusion of dead cells within a sample is an important step to preve erroneous interpretation of the data caused by nonspecific fluoresence signals. Whe performing cell imaging experiments, cell viability and live:dead ratio are parameters are frequently evaluated to measure cell health.						
	The proprietary dyes in the VivaFix Cell Viability Assay can easily assist researchers with distinguishing betweeen live and dead cells by covalently binding to primary amines. In live cells, the VivaFix Dyes bind to the cell surface primary amines. In dead cells, where the plasma membrane is compromised, the dyes are able to permeate the cell and also react with intracellular primary amines. As a result, a greater number of fluorophores is associated with dead cells and at least a 100-fold difference in fluorescence intensity is measured between the live and dead cells thereby allowing an easier discrimination between two populations. VivaFix Dyes are compatible with cell fixation without any significant loss of fluorescence.						
Reagents In The Kit	Kit includes 4 x 50 assay	vials and	250 µl of	DMSO, for running 20	00 assays		
Instructions For Use	Important: Thaw all com	ponents p	prior to us	е.			

Note 1: Any buffer without sodium azide, serum or protein can be used instead of phosphate buffered saline.

Note 2: For multicolor experiments, the cells can be stained with the antibodies of your choice before or after staining with VivaFix Dye. Antibody staining conditions should be optimized for your assay.

1. Prepare a 500x stock solution by adding 50 μ l of dimethyl sulfoxide (DMSO) to a VivaFix Dye vial and mix by vortexing.

2. Wash cells once with phosphate buffered saline, then resuspend cells at 2-3 x 10^6 cells/ml in phosphate buffered saline. Use 0.5 ml of cell suspension per assay.

3. Add 1 μl of 500x dye stock solution (from step 1) to 0.5 ml cells/assay and mix by vortexing.

4. Incubate the mixture for 30 min at room temperature (RT) or in a $37^{\circ}C/5\%$ CO₂ incubator. Protect from light.

Note: The optimal dye concentration and incubation time for different cell lines should be assessed empirically.

- 5. Wash cells twice in phosphate buffered saline.
- 6. Fix cells as desired. For 3.7% formaldehyde:

-Suspend cells in 900 µl of phosphate buffered saline following step 5

-Add 100 μI of 37% of formaldehyde to the cell suspension

-Incubate for 15 min at room temperature, then wash cells twice with phosphate buffered saline

7. Resuspend cells in the appropriate flow analysis buffer.

8. Sort cells with an <u>S3e Cell Sorter</u>, analyze cells with a flow cytometer, view and capture an image of cells with the <u>ZOE Fluorescent Cell Imager</u>, or view cells with a conventional fluorescence microscope.

 References
 1. Ramendra, R. *et al.* (2021) Glutathione Metabolism Is a Regulator of the Acute Inflammatory Response of Monocytes to (1→3)-β-D-Glucan. Front Immunol. 12: 694152.
 2. Globig, P. *et al.* (2021) Slow degrading Mg-based materials induce tumor cell dormancy on an osteosarcoma-fibroblast coculture model <u>Bioactive Materials. 30 Dec [Epub ahead</u> of print].

3. Méndez-Solís, O. *et al.* (2021) Kaposi's sarcoma herpesvirus activates the hypoxia response to usurp HIF2 α -dependent translation initiation for replication and oncogenesis. <u>Cell Rep. 37 (13): 110144.</u>

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Material Safety Datasheet documentation #1351111 available at:										
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